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CLIMATE CHANGE EFFECTS ON SOIL N AND P CYCLING AND GREENHOUSE GAS FLUXES IN CORK OAK FORESTS OF SOUTHERN SPAIN

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Table of Contents:

1	INTR	ODUCTION1
	1.1	BACKGROUND
	1.1.1	Soil nutrient cycling4
	1.1.2	Greenhouse gases
	1.2	OBJECTIVE
2	MAT	ERIAL AND METHODS9
	2.1	Study site
	2.2	Experimental design
	2.2.1	Establishment of Treatments
	2.3	FIELD PROCEDURES
	2.3.1	Soil Nutrient Sampling
	2.3.2	Greenhouse Gases Sampling11
	2.4	LAB PROCEDURES
	2.4.1	Soil Nutrient Analysis
	2.4.2	Greenhouse gases analysis
	2.5	Statistical Analysis
	2.5.1	
	2.5.2	Statistical Models
3	RESU	ILTS
	3.1	SOIL NUTRIENTS AVAILABILITY
	3.2	GREENHOUSE GASES FLUXES
4	DISC	USSION 24
	4.1	Soil Nutrients Availability
	4.2	GREENHOUSE GASES FLUXES
5	CON	CLUSION 29
6	REFE	RENCES
7	ANN	EX 40

List of Figures:

FIGURE 1: LOCATION OF LOS ALCORNOCALES PARK. MAP IMAGERY TAKEN FROM PROTECTED PLANET (2014)	2
FIGURE 2: SIMPLIFIED DIAGRAM OF NITROGEN CYCLE, FROM BERHANRD, 2010	4
FIGURE 3: SIMPLIFIED PHOSPHORUS CYCLE, FROM HYLAND ET AL., 2005.	6
FIGURE 4: A: EXAMPLE OF OPEN TOP CHAMBER (OTC) (WAHREN, 2019) B:RAIN EXCLUSION SETUP IN LOS ALCORNACALES	
(JIMÉINEZ ET AL 2018)	10
FIGURE 5: AVERAGE PRODUCTION RATES OF SOIL NUTRIENTS BY TREATMENT PER SEASON.	17
FIGURE 6: AVERAGE GREENHOUSE GAS FLUXES PER SEASON IN UG PER M2/DAY. LETTERS REPRESENTING SIGNIFICANT DIFFERENCES	
BETWEEN GROUPS.	19
FIGURE 7: AVERAGE GREENHOUSE GASES FLUXES BY TREATMENT IN EACH SAMPLING SEASON.	23
FIGURE 8: AVERAGE SOIL MOISTURE PERCENTAGE IN THE TOP 15CM OF SOIL OF THE STUDY SITE IN EACH TREATMENT OVER THE	
SAMPLING PERIOD.	26

List of Tables:

TABLE 1:: SOIL PHYSICAL PROPERTIES OF STUDY AREA, TAKEN FROM GÓMEZ-APARICIO ET AL (2017), WHERE VALUES REPRESENT
median [P10–P90, 10th and 90th percentiles] for 49 sampled points regularly distributed in a 70 m $ imes$ 70 m
SQUARE PLOT AT EACH SITE
TABLE 2: SUMMARY OF MODELS USED FOR ANOVA ANALYSIS FOR EACH VARIABLE. 14
TABLE 3: SUMMARY ANOVA RESULTS CONDUCTED ON GLS OF SOIL NUTRIENT CYCLING RATES. MEANS ARE
REPORTED WITH STANDARD ERRORS IN BRACKETS. ALL MEAN RATES ARE IN MG CM ² PER DAY
TABLE 4: RESULTS OF ANOVAS ON GLS MODELS OF SOIL NUTRIENT RATES BY TREATMENT AND SEASON. SIGNIFICANT RESULTS ARE
MARKED WITH '*' AND UNDERLINED. ALL MEANS ARE RATES IN UG PER CM ² PER DAY
TABLE 5: RESULTS OF ANOVAS FOR EACH GREENHOUSE GAS. MEANS ARE REPORTED IN UG PER M ² PER DAY. STANDARD ERRORS OF
THE MEANS ARE INCLUDED IN BRACKETS. SIGNIFICANT VALUES ARE MARKED WITH '*'
TABLE 6: RESULTS OF ANOVAS ON LMM MODELS OF GREENHOUSE GASES BY TREATMENT AND SEASON. AVERAGE RATES OF
EMISSION ARE EXPRESSED IN UG PER M ² PER DAY, WITH STANDARD ERROR IN BRACKETS. SIGNIFICANT RESULTS ARE MARKED
WITH '*'

Abstract

English

The Mediterranean basin is a culturally and biodiversity rich region that is expected to suffer some of the worst effects of predicted climate change with higher temperatures, significant decreases in precipitation, and more extreme climatic events. Los Alcornocales natural park in southern Spain is one of the largest protected cork oak forests in the region, with great ecological, cultural, and economic value. Its longstanding forests provide many ecosystem services but are threatened by several interacting stressors such as human disturbances, climatic changes, pathogens, and pests. Thus, we aimed to study how key ecosystem processes, soil nitrogen and phosphorus cycling and greenhouse gas fluxes, would be impacted by these predicted climatic changes. Treatments were established that increased temperature by 2-3°C, reduced rainfall by ~30%, and a combination treatment of both. We found that all soil nutrients were significantly impacted by reductions in rainfall, with altered seasonal patterns of production. Greenhouse gases fluxes were also impacted on a seasonal basis, with nitrous oxide emissions significantly increased overall by temperature treatments, carbon dioxide exhibiting variable responses, and methane fixation rates reduced under climate change conditions. We conclude that soil nutrient cycles and greenhouse gas fluxes were significantly altered by our treatments, and that climate change will have serious impacts on ecosystem functionality.

Portuguese

A bacia do Mediterrâneo é uma região rica em cultura e biodiversidade que é provável que sofra alguns dos piores efeitos das mudanças climáticas, incluindo temperaturas mais altas, diminuições significativas na precipitação e eventos climáticos mais extremos. O parque natural "Los Alcornocales", no sul da Espanha, é uma das maiores florestas de sobreiro protegidas do Mediterrâneo, com um grande valor ecológico, cultural e econômico. As suas florestas fornecem muitos serviços ecossistêmicos, mas estão ameaçadas por vários fatores de estresse interativos, como perturbações antropogénicas, mudanças climáticas, patógenos e pragas. O objetivo de esta tese é estudar como processos ecossistêmicos chave (especificamente a produção de nitrogênio e fósforo no solo e os fluxos de gases de efeito estufa) vão ser impactados por essas mudanças na temperatura e na precipitação previstas pela maioria dos modelos climáticos. Para alcançar os nossos objetivos, foram estabelecidos em parcelas naturais sistemas de manipulação climática que aumentaram a temperatura do solo em 2-3°C, reduziram a precipitação em $\widetilde{}~30\%$ (alem de um tratamento combinado dos dois anteriores). Os nossos resultados mostram que a produção de todos os nutrientes do solo considerados foi significativamente influenciada pelas reduções na precipitação, com padrões sazonais de produção também alterados. Os fluxos de gases de efeito estufa mostraram uma grande variabilidade sazonal. Apesar disso, as emissões de óxido nitroso aumentaram significativamente com o aumento datemperatura^[41], e as taxas de fixação de metano foram reduzidas sob condições de mudanças climáticas. Estes resultados permitem-nos concluir que as mudanças previstas na temperatura e na precipitação provavelmente resultaram em alterações significativas na capacidade destas florestas para disponibilizar nutrientes e trocar gases de efeito estufa com a atmosfera.

Keywords: N cycling, P cycling, greenhouse gases, climate change, Mediterranean

List of Abbreviations

CO_2	Carbon dioxide
CH_4	Methane
GHG	Greenhouse gas
Ν	Nitrogen
N_2O	Nitrous oxide
NH_4	Ammonium
$\rm NH_4NO_3$	Total inorganic nitrogen
$ m NH_4NO_3$ $ m NO_3$	Total inorganic nitrogen Nitrate
	0 0
NO_3	Nitrate
NO ₃ OTC	Nitrate Open top chamber

1 Introduction

The current climatic changes that we are experiencing are unprecedented and are expected to continue to intensify across the world in the coming years. These changes are likely to have important implications for ecosystem functioning, and consequently, human livelihoods (IPCC, 2018). This is particularly true in the case of the Mediterranean region in Europe, where 472 million called the region home in 2010, and is predicted to reach 572 million by 2030 (UN Environment Program, 2017). The Mediterranean basin may only cover 2 % of the Earth's surface, but it is one of 34 biodiversity hotspots based on the multitude of plant and animal species that inhabit it, including a high number of endemic species (Gauquelin et al., 2018; European Commission, 2009).

This culturally and biodiversity rich region is expected to suffer some of the worst effects of forecasted climate change. Average temperatures in the region have already risen by 1.4°C since the pre-industrial era, and drought severity has intensified with a loss of 15.6% precipitation with an additional 7.4% evaporative demand in the last five decades (Avila et al. 2019; Vicente-Serrano et al. 2014). The IPCC forecasts that southern Europe will continue to experience higher temperatures than any other region in Europe, with significant decreases in precipitation and more extreme climatic events. Droughts are expected to lengthen, and soil moisture is predicted to significantly decrease in response to the higher temperatures. (Grillakis, 2019; Cramer et al., 2018; Tueling, 2018; Ozturk et al., 2015; Valentini et al., 2014).

Increasing temperatures and loss of soil moisture by reductions in rainfall both have known impacts on soil functioning and plant growth. Thus, reductions in soil moisture limits plant growth and nutrient uptake, and many soil processes are dictated by complex microbial communities which are very responsive to changes in temperature and soil moisture, with studies showing reductions in soil enzymatic activity or community composition and abundance under drought conditions (Preece et al., 2019; Bastida et al., 2019; Ren et al., 2018; Naylor et al., 2018; Xu & Yuan, 2017; Meisner et al., 2013; Auyeung et al., 2013 Sardans & Peñuelas, 2008). This can lead to changes in the rate and timing at which bioavailable soil nutrients are transformed (Homyak et al., 2017; Matías et al., 2010; Sardans & Peñuelas, 2005), and in the rate of emission or fixation of greenhouse gases (Smith et al., 2018; Oertel et al., 2016; Martins et al., 2015). However, as these reactions have large spatial variability due to differences in soil characteristics and local climatic dynamic, the effects of climate change are likely to differ across localities (Reys-Garcia et al., 2016), and we are still far to fully understand how climate will impact ecosystem functioning in many areas of the planet, including Mediterranean forests.

Therefore, this study was conducted in Los Alcornocales natural park, a protected cork-oak forest in southern Spain with great ecological, cultural, and economic value. Its long-standing forests provide many ecosystem services but are threatened by several interacting stressors such as human disturbances, climatic changes, pathogens, and pests. (Costa, 2016; Kim et al., 2017; Moricca et al., 2016). These oak forests have provided economic stability for local communities for at least the past two centuries through cork-oak production, with 16,000 metric tons of cork being harvested from this area annually (Aronson et al., 2009). Other resources include are lowland pasture

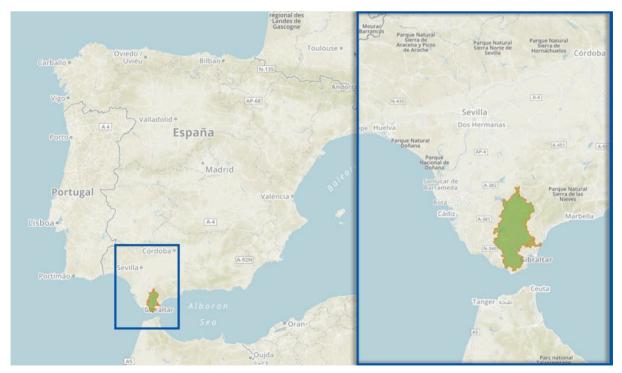


Figure 1: Location of Los Alcornocales park. Map imagery taken from Protected Planet (2014).

for free-range cattle, wild game for hunting such as red deer (*Cervus elaphus*) and roe deer (*Capreolus capreolus*), and many non-timber forest products (Costa, 2016; Kim et al., 2017, Aronson et al., 2009). Besides provisioning ecosystem services, the forest also provides regulating services such as soil conservation, water retention, watershed protection, erosion control, fire risk prevention, carbon sequestration (WWF, 2004; Aronson et al., 2009). Climate change is already directly impacting these forests with decreased precipitation and increased temperature causing physiological stress to the trees, causing limited nutrient uptake, stem mortality and seedling death (Liu et al., 2018; Costa, 2016; Pressarakli, 1999). Invasive pathogens such as *Phytophthora cinnamoni*, are spread much more quickly under increasing temperatures and longer warm seasons, and limit nutrient uptake in cork oaks causing further stress and extensive die-off (Homet et al., 2019; Gómez-Aparicio et al., 2017; Kim et al., 2017; Avila et al., 2016). Thus, the interacting challenges that these valuable forests are facing make understanding the effects of predicted climate change on soil nutrient cycling and greenhouse gases emissions even more imperative.

1.1 Background

While these forests are already adapted to nutrient poor soils, further reductions or changes in soil nutrients could be detrimental (Liu, 2010). Soil nitrogen (N) and phosphorus (P) are two of the most important soil nutrients for microorganisms and vegetation, and the available forms of these nutrients are largely produced in the soil by complex communities of soil microbes (Bardgett & van der Putten, 2014). Many of the processes involving soil nutrients are also interconnected with the exchange of greenhouse gases (GHG) between the soil and the atmosphere, so the determination of climate change effects on these soil processes are integral (Walkiewicz & Brzezińska, 2019; Oretel et al., 2016; Veraart et al., 2015; Singh, 2011; Leifeld, 2007). Decreases in soil moisture, due to decreases in precipitation, have been shown to suppress microbial activity and to change or decrease soil microbial communities that facilitate the soil nutrient cycling processes (Ren et al., 2018; Homyak et al., 2017; Bérard et al., 2011). Meanwhile, increases in temperatures are shown to stimulate microbial

Climate change effects in cork oak forests of southern Spain

communities, which could at least partially compensate the effect of decreases in precipitation. Similarly, increases in temperature have also been shown to increase the soil to atmosphere greenhouse gases fluxes (Duan et al., 2018; Schaufler et al., 2010). Thus, forecasted increases in temperature could at least partially compensate the negative effects of decreasing soil moisture on soil functioning, at least in the short term (Pold & DeAngelis, 2013). However, increasing soil temperature also increases soil evapotranspiration, which could exacerbate the negative impacts of decreasing precipitation in water-limited ecosystems (Samaniego et al., 2018; Vicente-Serrano et al., 2014)

1.1.1 Soil nutrient cycling

The soil nitrogen cycle is composed of four main steps: fixation, mineralization, nitrification, and denitrification (Figure 2). Fixation is the process by which atmospheric gaseous dinitrogen (N2) is fixed in the soil by diazotrophic bacteria, either free-living in the soil substrate or those that have a symbiotic relationship with plant roots. These bacteria use nitrogenase enzymes to transform N_2 , which is metabolically useless to all but a few microorganisms, to ammonia, which can metabolized by most organisms and incorporated into biological structures (also known as immobilization). Mineralization occurs thereafter, where the previously fixed and immobile ammonia is

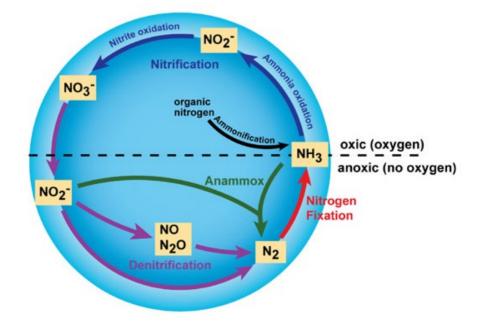


Figure 2: Simplified diagram of nitrogen cycle, from Berhanrd, 2010.

then oxidized to ammonium (NH_4^+) , becoming again bioavailable to plants and soil microorganisms. The rates of mineralization vary with soil temperature, moisture and the amount of oxygen in the soil (Fowler et al., 2013; Singh, 2011; Bernhard, 2010; Johnson et al., 2005). After mineralization, ammonium undergoes a two-step conversion by soil-living bacteria which obtain energy by converting ammonium to nitrite (NO_2) , and then to nitrate (NO_3) under a process known as nitrification. Nitrification is most rapid when soil is warm (between 28°C and 36°C), moist, and well-aerated (Bernhard, 2010; Johnson et al., 2005; IPNI, 2004a). Nitrifying bacteria are especially sensitive to changes in soil water content as it affects the equilibrium between soluble and exchangeable ammonium, the concentration of salts, and oxygen availability (IPNI, 2004a). The final step of the nitrogen cycle is denitrification, where nitrate is reduced progressively into gaseous forms of nitrogen gas (N_2) , nitric oxide (NO), or nitrous oxide (N_2O) under primarily anaerobic conditions. The determination of the ratios of these outputs depends a lot on the soil conditions, particularly soil water content. Denitrification can occur at low rates under aerobic conditions; however, it is the dominating process under anaerobic conditions. At 60% waterfilled pore space nitrous oxide is the primary byproduct, while dinitrogen gas becomes the primary product after about 75% water-filled pore space (van Groenigen et al., 2015; GRDC, 2014; Singh, 2011; Bernhard, 2010; IPNI, 2004b). Temperatures have less impact on denitrification rate than soil moisture, but ideal temperatures are between 27-38°C (GRDC, 2014)

While phosphorous usually only makes up about 0.2% of plant weight, it is also a necessary nutrient for plant growth, maturity, and reproduction. This is because phosphate is an important part of adenosine triphosphate (ATP), which is used in many biochemical processes in plants and allows them to extract nutrients from soils. It is necessary for energy transport, cellular structures, and nucleic acids (Margalef et al., 2017; Hyland et al., 2005). Bioavailable, or inorganic, phosphorous is transformed through the mineralization of organic P to $H_2PO_4^-$ or HPO_4^{-2} -(Figure 3) (Hyland et al., 2005). Mineralization is made possible by phosphatase enzymes that are produced by bacteria, fungi, and plant roots (Maragalef et al, 2017). It has been shown that microbial communities responsible for phosphorus mineralization have sensitivity to temperature and soil moisture changes, with decreases in inorganic production rates under drought conditions and increases of temperature above 40°C (Brödlin et al.,2019; Maragalef et al, 2017; Hinojosa et al., 2012; Butterly et al., 2011; Sardans & Peñuelas, 2007, 2004).

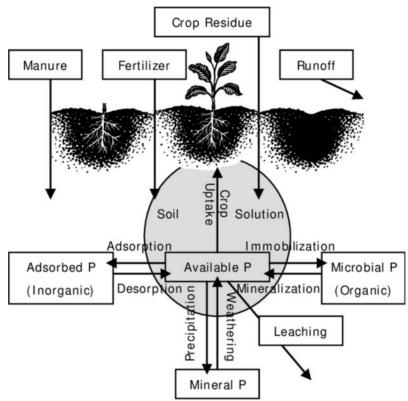


Figure 3: Simplified phosphorus cycle, from Hyland et al., 2005.

1.1.2 Greenhouse gases

Greenhouse gases are major contributors to global warming, and investigating their sources is important to properly inform global climate models for accuracy, and for management strategies to minimize their output (Oretal et al., 2016). Soils have the potential to emit GHG through transformational processes by soil microbial communities, but also to act as 'sinks' by capturing organic forms of these elements. These dynamics depend on the biotic and abiotic soil characteristics of each ecosystem. The biotic sources of greenhouse gases are driven by dynamic and complex processes and vary with environmental conditions such as plant and microbial community structure, soil organic matter, ammonium and nitrate concentrations, soil water content, and temperature (Shvaleva et al., 2011; Leifeld et al., 2007). Therefore, the fluxes of greenhouse gases can be significantly impacted by changes in climate conditions. In this study we consider the flux dynamics between soil and atmosphere for the three most detrimental greenhouse gases: carbon dioxide (CO_2), nitrous oxide (N_2O), and methane (CH_4).

Carbon dioxide is the most notorious greenhouse gas, and much of the anthropogenically caused global warming is attributed to the increases in CO_2 emissions from human activities over the past century. Carbon dioxide is emitted from soil through microbial respiration (anaerobic and aerobic) and root respiration (Rastogi et al., 2002). Emission rates of CO_2 are dependent on a number of factors such as soil texture, pH, available organic carbon, and nitrogen content of the soil, but temperature and soil moisture hold the most sway over the decomposition rates that result in CO_2 emissions (Smith et al., 2018; Rastogi et al., 2002).

Nitrous oxide gas is a long-lived relatively active greenhouse gas with an atmospheric lifetime of about 120 years. Its global warming potential is 265 times more powerful than CO₂ and is responsible for about 6% of global warming (Myhre et al., 2013; Ussiri & Lal, 2013). It also plays a significant role in the depletion of stratospheric ozone (Ussiri & Lal, 2013). Nitrous oxide is emitted by soils during nitrification (oxidation of $\rm NH_4^+$ to $\rm NO_2^-$ and $\rm NO_3^-$) and denitrification (reduction of $\rm NO_3$ to $\rm N_2O$ and $\rm N_2$). However, its mainly produced by denitrification under anaerobic conditions, when soil water content is above 50% (Oertel et al., 2016; Groenigen et al., 2015). The availability of inorganic N together with organic matter (heterotrophic transformation) or carbon dioxide and oxygen (autotrophic transformation) also determines N₂O production rates (Leifeld, 2007). Nitrifier denitrification is a relatively newly discovered source of N₂O, and there is a lot of discussion on how much is actually emitted under this process, so it's significance as a source of N₂O is still under discussion, especially in dryland ecosystems (Smith et al., 2018; Wrage-Mönnig et al., 2018; Oretel et al., 2016; Kool et al., 2011.)

Climate change effects in cork oak forests of southern Spain

The concentration of CH_4 in the atmosphere has more than doubled since the preindustrial era, with soils contributing about 40% of total emissions (Smith et al., 2018). Methane has a global warming potential of 28 times that of carbon dioxide, with an atmospheric life time of 12.4 years (Myhre et al., 2013). In anerobic soils, methane is produced through methanogenesis, and is consumed by methanotrophic microorganisms that use O_2 and CH_4 for their metabolism under aerobic conditions (Oretel et al., 2016; Smith et al., 2018; Leifeld, 2007). While most atmospheric methane is destroyed by oxidation in the atmosphere, 5–10% is diffused into aerated topsoils, where it is oxidized by soil microorganisms, this diffusion is primarily influenced by soil moisture, texture and bulk density (Shvaleva et al., 2014). Soils are CH_4 sinks under aerobic conditions, and sources in anerobic conditions. In the Mediterranean, forest soils are typically sinks of methane, not sources (Oertel et al., 2016; Shvaleva et al., 2011). However, changes in soil moisture and precipitation, as well as the forecasted more sporadic but intense precipitation events, are likely affect the net flux of CH_4 between the soil and the atmosphere in these areas.

1.2 Objective

In this study, we aimed use a manipulative field experiment to investigate the changes induced by increases in temperature and decreases in precipitation (and the combination of both) on the cycling of soil nitrogen and phosphorus, and on the fluxes of carbon dioxide, nitrous oxide, and methane in the soils of Los Alcornocales natural park. Many of these processes have been shown to be sensitive to forecasted climate changes. However, changes in ecosystem functioning are the result of complex interactions between these key processes and site-specific environmental conditions, making it difficult to forecast future scenarios based on single-factor studies or on studies carried out in other systems. Most of these studies are conducted in more temperate or tropical ecosystems, with either observational or laboratory-based manipulations that typically only include one factor. This manipulative study is novel in its field-based, full-factorial design that is inclusive of seasonal variation over one full year of sampling. Due to the variation between seasons, which is characteristic of

Mediterranean ecosystems, climate change may have different impacts on soil functioning at different times of year, and therefore this study offers insight to these interacting dynamics.

2 Material and Methods

2.1 Study site

This study was carried out at Los Alcornocales Natural Park in Southern Spain, known to be an ecological hotspot and one of the largest cork-oak forests in Western Europe, covering 170,025 hectares (Aila et al., 2017; Jiminez-Chacon et al., 2018). The climate of the region is characterized as sub-humid Mediterranean with hot, dry summers and mild, wet winters. Temperatures range between 7-10°C in winter, and 22-26°C in summer, with an annual average temperature of 15.7°C (Avila, 2017, 2019). Due to its proximity to the Mediterranean Sea, and the predominant western winds, there is a relatively high degree of humidity, with an average annual rainfall of 748-1100mm, 95% of it occurring between October and May (Jiminez, 2018; Avila, 2018; Aponte Matais et al., 2014). The study site was established in the drier lowlands of the park (36°34'07.5″, N 5°32'08.3″ W), where forest grows in acidic, nutrient-poor, sandy soils and is composed of two dominant tree types: *Quercus suber L.* and *Olea europaea var. sylvestris Brot* (Table 1) (Avila, 2017).

Table 1:: Soil physical properties of study area, taken from Gómez-Aparicio et al (2017), where values represent median [P10–P90, 10th and 90th percentiles] for 49 sampled points regularly distributed in a 70 m × 70 m square plot at each site.

Soil chemical and physical properties									
Total C (mm/g)	45.15 [35.17,55.70]								
Total N (mm/g)	$2.30 \ [1.92, 2.90]$								
Available P (mm/Kg)	7.50 [6.40-9.81]								
Ca (mm/Kg)	684.32 [467.4-865.4]								
$Mg \ (mm/Kg)$	142.23 [91.47-201.81]								
K (mm/Kg)	117.63 [81.77-146.18]								
Clay content (%)	12.84 [10.03-16.71]								
Sand content $(\%)$	73.59 [69.84-78.36]								

2.2 Experimental design

2.2.1 Establishment of Treatments

In 2015, six experimental plots (20 x 15 m) were established within the study site: three that included rain exclusion (RE) structures and three control. Each RE plot was located adjacently to a control plot to minimize edaphic variations between the two treatments. The three paired blocks were separated by a minimum of 200m. In each plot, 3 adult individuals per tree species were selected and characterized (36 individuals in total). Rainfall exclusions were created by suspending plastic white PVC channels without ultraviolet filter of 2.5 cm in diameter over the surface of the forest floor at a minimum height of one meter to minimize the microclimatic effect on the soil (Figure 4B). The structure was supported by a network of iron supports to ensure the stability of the structure in the long term and allow its slope to be adjusted to a minimum of 5% for the correct evacuation of intercepted water. Each RE plot had 24 rows of channels separated from each other by approximately 70 cm. In the control



Figure 4: A: Example of open top chamber (OTC) (Wahren, 2019) B:Rain exclusion setup in Los Alcornacales (Jiméinez et al 2018)

plots, a similar structure was built, but with the channels upside down so that possible microclimatic effects of the structure were reproduced but without intercepting rainwater. To avoid edge effects no measurements were made in the outer meter of the perimeter of each plot. All plots were fenced to avoid the passage of large herbivores that could damage the infrastructure. In addition to the rainfall exclusion treatment. а temperature increase treatment with two levels (ambient and +2-3 °C) was set up next to each selected tree since 2016. A 2-3 °C increase was

achieved by using $40 \times 50 \times 32$ cm hexagonal open top chambers (OTCs, Figure 4A). Open top chambers are a well-established method to simulate climate warming and have been successfully used to study the effect of warming from arctic and tundra plants (Yang et al, 2018; Bokhorst et al 2013) to tropical grasslands (Buhrmann et al, 2016). The OTCs were made of methacrylate and elevated 5 cm from the surface to avoid overheating. Thus, our full-factorial experimental design comprised a total of four treatments: control, rain exclusion, temperature, and a combined treatment of temperature and rain exclusion (T+D); as well as 18 sampling points per treatment (72 sampling points in total). All sampling points were sampled in summer, fall, and winter of 2017, and in spring of 2018 to account for interseasonal variability. Winter 2017 samples were omitted for soil nutrient measurements, due to sampling error.

2.3 Field procedures

2.3.1 Soil Nutrient Sampling

To assess soil total inorganic nitrogen (N) and phosphorus (P) availability, we used ion exchange membranes (IEMs). The resins used on IEMs exhibit emulate plant root surfaces, taking into account the nutrient diffusion capacity in soil. Studies have shown that the ion concentrations in IEM extracts correlate to plant uptake of nutrients, thus, these membranes have been established as an effective method for the analysis of soil nutrients availability and production (Duran et al, 2013; Qian, Schoenau, & Ziadi, 2008, Rieman & Walton, 1970). For each sampling point, two sets of cationic and anionic exchange membranes (3cm x 3cm) were attached to 30 cm rulers at 15 cm and 30 cm depth using acrylic adhesive, vertically placed in the soil leaving the first 5 cm out for removal (therefore the actual depths were 10 and 25 cm), and incubated for one month. After removal, they were kept in plastic bags in a cool dry place until lab processing.

2.3.2 Greenhouse Gases Sampling

We used the static chamber technique to measure the in-situ greenhouse gases (GHG) exchange between the soil and the atmosphere. A PVC ring (diameter = 20 cm, height = 10 cm) was inserted approximately 5 cm into the ground at the start of

the experiment at each sampling point. Immediately before each measurement, a 20 cm diameter, 9 cm high PVC chamber was placed on top of each ring and sealed with a rubber band. Each chamber had a sampling port in the top center of the chamber that allowed air sampling and was covered with reflective material to thermally isolate it during the measurement. Gas samples were collected at 0, 30, and 60 min after chamber closure using a hypodermic needle attached to a polypropylene syringe, transferred to 22 ml pre-vacuumed vials and kept at room temperature until analysis. Measurements were taken in a randomized design between 10 a.m. and 12 p.m. to minimize sharp temperature variation effects on GHG sampling.

2.4 Lab Procedures

2.4.1 Soil Nutrient Analysis

The IEMs were separated from the rulers, washed with distilled water, and air dried. Then, NH_4^+ -N, NO_3^- -N and PO_4^{-3} -P were extracted from the membranes following Rodríguez et al. (2009). In summary, IEMs were cut in half, placed in 50 ml falcon tubes, and shaken for 60 minutes at 200 rpms in 30 ml of 1M KCl. These extracts were immediately filtered through standard lab filters (125mm diameter ashless filter papers, GE Healthcare & Filter Lab) and kept in the freezer (-20°C) until analysis. The amount of inorganic N and P in the extracts was analyzed by colorimetry using a microplate reader (Sims et al., 1995). In the case of N, we used the indophenol blue method, while P was measured using the molybdenum blue method (Allen et al., 1986). Concentrations (ppm) of NH_4^+ -N, NO_3^- -N and PO_4^{-3} -P in the extracts were estimated with the help of standard curves. The ppm values were then converted to a rate of μ g of nutrient per square centimeter per day by using the following formula:

$$r = \frac{\frac{PV}{S}}{t}$$

Where the rate (r) equals the concentration of the sample in ppm (P) per the volume of KCl used to extract the sample in ml (V), divided by the surface area of the IEM in cm² (S), divided again by the incubation time in days (t). We then calculated soil total inorganic N (NH₄NO₃-N) as the sum of NH₄⁺-N and NO₃⁻-N.

2.4.2 Greenhouse gases analysis

Gas samples were analyzed at the Instituto Superior de Agronomia (Lisbon), using gas chromatography. Specifically, by electron capture, thermal conductivity and flame ionization detection in the cases of N2O, CO2 and CH4, respectively. The carrier gas used was helium in all cases. The returned chromatograph values were used to calculate the concentration of each gas in the sample using the ideal gas law equation, and to estimate net gas fluxes between the soil and the atmosphere from the linear rate of change in gas concentration, the chamber internal volume, and the soil surface area (Durán et al., 2013)

2.5 Statistical Analysis

2.5.1 Data Exploration

First, boxplots were conducted for each independent variable to identify potential outliers (determined as values three times the interquartile range) and heteroscedasticity problems. No outliers were removed from soil nutrient data, while 16 outliers were removed from greenhouse gases data (six outliers from N₂O, five each from CO_2 and CH_4). Then, we constructed histograms to check for normality. Finally, we conducted an analyses of summary statistics to get means and standard errors for each variable among other statistics.

2.5.2 Statistical Models

As most nutrient variables were not normal and showed heteroscedasticity, we used generalized least squares (GLS) models. These models allow for non-normality and they allow the use of weight and correlation arguments to account for the typical temporal dependencies derived from the use of repeated measures (Hansen, 2007). Thus, we used temperature and rain exclusion treatments, as well as season as fixed factors. *Tree* was used as a correlating factor and an interaction between treatment and a season as weighted argument to correct heteroscedasticity.

To model the impact of our treatments and account for seasonal differences in soil greenhouse gas emissions, linear mixed-effect models (LMMs) were used. These models have been often used in ecological data to account for field variations, nested, and hierarchical relationships between variables in datasets, such as repeated measurements of an individual over time (Harrison et al, 2018). Here, we used *Tree* as a random factor, to incorporate the repeated sampling dependencies. Temperature and rain exclusion treatments were included as fixed factors, with two levels each. Season was also included as a fixed factor with four levels, to evaluate inter-seasonal variation. Residuals from the models were visually tested for normality and homoscedasticity. Methane and carbon dioxide were deemed normal and homoscedastic, while nitrous oxide was able to meet these assumptions after applying a square root transformation.

Model selection was carried out through a step-wise selection procedure using the Aikake information criterion (AIC) to select the best-fitted models. No-significant factors (high p-values) remained in the models when their interaction with other factors was significant. After determining the best model, an ANOVA was conducted to evaluate the significance of treatments and interactions. A summary of all models and factors used is found in Table 2, and all model results are found in Appendix I-VIII. All statistical tests were conducted using R statistical software (R Core Team, 2019). Linear mixed models were performed using the *lmer* package (Bates et al, 2015), and GLS models performed using the *nlme* package (Pinheiro et al., 2019).

Table 2: Summary of mode	ls used for ANOVA	l analysis for each	variable.
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	Dependent Variable	Transformation	Model Type	Fixed Factors	Random Factor	Weights (GLS only)
Q 1	$\rm CO_2$	None	LMM	T, R, S		
Greenhouse Gases	NO_2	Square root	LMM	T, R, S		
Gases	CH_4	None	LMM	T, R, S		
G :1	NH_4	None	GLS	T, R, S	Tree	Tt, S
Soil	$\rm NH_4~NO_3$	None	GLS	T, R, S		Tt, S
Nutrients	NO ₃	None	GLS	T, R, S		Tt, S
	PO_4	None	GLS	T, R, S		Tt, S

Factor abbreviations: T=Temperature, R=Rain exclusion, S=Season, Tt=Treatment

Results

3 Results

3.1 Soil nutrients availability

All surrogates of soil nutrient production rates were significantly affected by the rain exclusion treatment (Table 3). Specifically, soil ammonium (NH₄), total inorganic nitrogen (NH₄ + NO₃), and inorganic phosphate (PO₄) production rates were 42%, 9%, and 79% higher in the rain exclusion plots than in the control plots. Conversely, soil nitrate (NO₃) production was significantly lower in the rain exclusion plots than in the control. Neither temperature nor the combination of temperature and rainfall exclusion significantly affected any of the overall soil nutrient production rates. Our results also revealed a significant effect of season on all the soil nutrient production rates, with values consistently lower in summer than in fall and spring (Table 4) (Figure 5). We also found significant interactions between season and the rain exclusion treatment for the production of available ammonium, nitrate, and phosphate (Figure 5A,C,D). A significant interaction between our temperature treatment and season was also found for soil ammonium production (Figure 5A).

Our rain exclusion treatment decreased ammonium production by 30% in summer but increased it by 47% in fall. Ammonium production was also was found to be significantly increased by our temperature treatment in fall by 33%. Total inorganic N production was most impacted by the rain exclusion and combination treatments in spring, with a 45% and 13% increase, respectively, as compared to the control plots. In summer the rate was reduced by 39% in the rain exclusion treatment as compared to the control plots. Nitrate production was significantly impacted by the rain exclusion in summer, where the treatment decreased the rate by 50%. When considering a 90% confidence interval, nitrate also significantly increased (p=0.06) in the spring season by 22%. It was also revealed that the rate of phosphate production increased in fall in response to the rain exclusion treatment by six times the control rate, while remaining stable in spring and summer under all treatments.

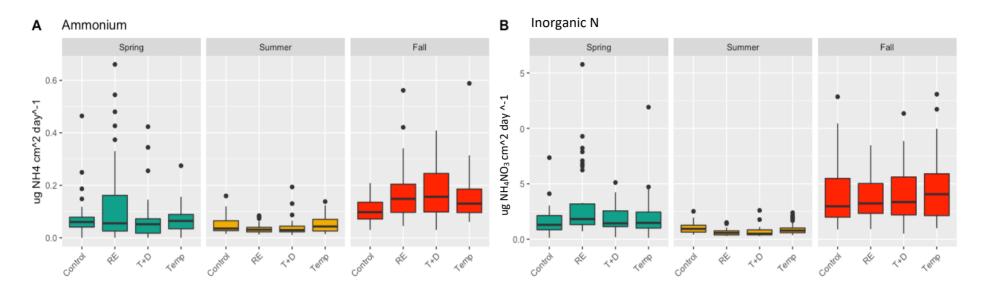
Climate change effects in cork oak forests of southern Spain

Table 3: Summary ANOVA results conducted on GLS of soil nutrient cycling rates.are reported with standard errors in brackets. All mean rates are in $\mu g \ cm^2$ per day.

ANOVA Mean Ammonium **D**.**F**. F р *Control* 0.076 (0.005) _ _ *Temperature* 0.092 (0.007) 1 1.910.167*RE* 0.108 (0.011) 1 24.39< 0.0001* $T+D \quad 0.097 \quad (0.010)$ 1 0.260.608 286.81 Season -< 0.0001* Temp:Season -20.0026*6.022RE:Season -8.53 0.0002^{*} 2T+D:Season -1.980.1399Nitrate *Control* 0.150 (0.018) _ _ *Temperature* 0.156 (0.021) 3.050.0811 0.134(0.013)1 23.72< 0.0001*RE $T+D \quad 0.126 \quad (0.016)$ 1 0.820.365 Season -286.55< 0.0001*2Temp:Season -1.560.212 2RE:Season -5.77 0.0034^{*} *Control* 0.150 (0.018) _ _ _ Inorganic N *Control* 0.228 (0.020) _ _ *Temperature* 0.258 (0.027) 0.1921 1.70RE 0.248 (0.022)25.441 < 0.0001* $T+D \quad 0.223 \ (0.021)$ 1 0.350.5532145.76< 0.0001*Season -2Temp:Season -0.823 0.440 2RE:Season 2.430.090 _ 2T+D:Season -3.081 0.047^{*} **Phosphate** Control 0.0024 (0.0003) _ *Temperature* 0.0025 (0.0003) 0.092 1 2.860.0043(0.0004)1 19.58< 0.0001*RE $T+D \quad 0.0034 \quad (0.0004)$ 0.058 1 3.60241.94 $\leq 0.0001^{*}$ Season -2Temp:Season -0.3640.695 RE:Season -211.23< 0.0001*

* Indicates a significant result

Results



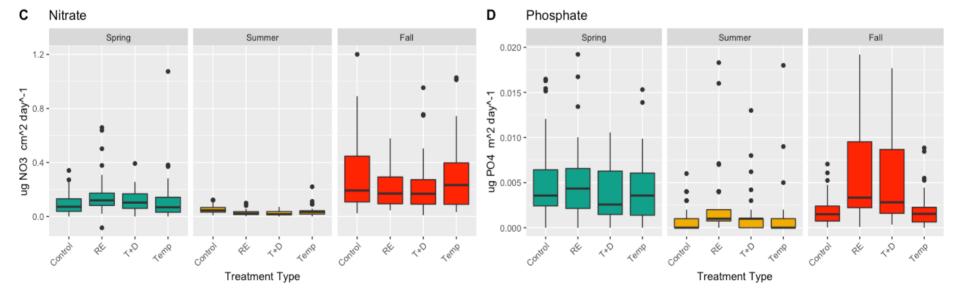


Figure 5: Average production rates of soil nutrients by treatment per season.

Table 4: Results of ANOVAs on GLS models of soil nutrient rates by treatment and season. Significant results are marked with '*' and underlined. All means are rates in ug per cm² per day.

	S	pring			Summer		Fall			
	Mean	χ^2	р	Mean	χ^2	р	Mean	χ^2	р	
Ammonium										
Control	0.074(0.01)	-	-	0.048(0.005)	-	-	0.106(0.006)	-	-	
Temp	0.069(0.01)	2.52	0.11	0.052(0.005)	1.31	0.25	0.156(0.017)	5.43	0.02*	
RE	0.124(0.03)	1.46	0.23	0.034(0.003)	3.80	0.05^{*}	0.167(0.015)	5.52	0.02*	
T+D	0.073(0.03)	1.50	0.22	0.040(0.006)	0.18	0.67	0.174(0.017)	2.80	0.09	
Nitrate										
Control	0.089(0.01)	-	-	0.053(0.004)	-	-	0.305(0.04)	-	-	
Temp	0.124(0.03)	0.092	0.76	0.041(0.006)	2.30	0.13	0.303(0.05)	0.040	0.84	
RE	0.163(0.02)	2.75	0.10	0.027(0.002)	19.03	$1.28e-5^{*}$	0.215(0.02)	2.94	0.09	
T+D	0.114(0.01)	3.61	0.06	0.026(0.003)	1.84	0.17	0.238(0.04)	0.040	0.84	
Inorganic N										
Control	0.166(0.02)	-	-	0.100(0.008)	-	-	0.412(0.04)	-	-	
Temp	0.204(0.04)	1.40	0.24	0.092(0.009)	0.10	0.75	0.472(0.06)	1.32	0.25	
RE	0.307(0.05)	4.10	0.04*	0.061(0.004)	17.0	$3.8e-5^{*}$	0.382(0.03)	0.76	0.38	
T+D	0.189(0.02)	5.12	0.02*	0.067(0.008)	1.20	0.27	0.412(0.04)	0.30	0.59	
Phosphate										
Control	0.005(7.0e-4)	-	-	0.002(2.0e-4)	-	-	0.001(3.0e-4)	-	-	
Temp	0.004(6.0e-4)	2.16	0.14	0.001(6.0e-4)	0.03	0.86	0.002(3.0e-4)	0.98	0.32	
RE	0.005(6.0e-4)	0.01	0.92	0.002(6.0e-4)	2.31	0.13	0.006(7.0e-4)	13.8	$\underline{2.07e-4^{*}}$	
T+D	0.004(5.0e-4)	0.04	0.84	0.002(4.0e-4)	2.42	0.12	0.005(7.0e-4)	2.23	0.14	

Results

3.2 Greenhouse gases fluxes

The net exchange of carbon dioxide (CO₂) and methane (CH₄) between the soil and the atmosphere was not significantly affected by any of the treatments. However, nitrous oxide (N₂O) net fluxes were significantly influenced by the temperature treatment (F=8.95, p=0.003), with a negative emission rate (net fixation) in the control plots, but a positive rate (net emission) in the temperature treatment plots

(Table 5). There were no significant interactions between season and the treatments. However, all gases fluxes significantly varied between seasons (Table 5). Carbon dioxide emissions were significantly highest in spring and lowest fall, with insignificant differences between all seasons (Figure 6A). Nitrous oxide was also found to be significantly different between all seasons (Figure 6B). Methane was fixated at the highest rate in spring (lowest rate of emission), with no significant differences between fall, winter, and summer. (Figure 6C).

Large variations in the response variable values inhibited our ability to detect statistically significant differences in GHG fluxes rates. However, observing seasonal averages yield insights

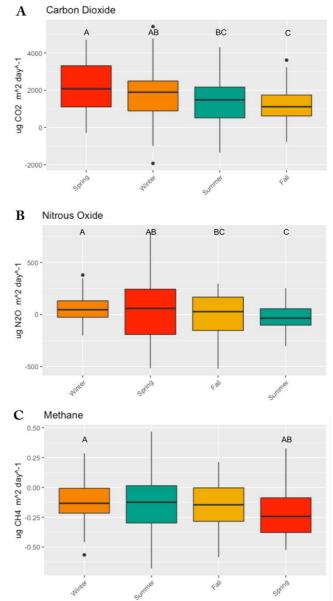


Figure 6: Average greenhouse gas fluxes per season in ug per m2/day. Letters representing significant differences between groups.

	Mean	ANOV	A	
Carbon dioxide		D . F .	F	р
Control	1781.5(174.7)	-	-	-
Temperature	1437.1(134.6)	1	2.39	0.123
RE	1772.0(152.6)	1	0.54	0.469
T+D	1665.1 (145.9)	1	0.61	0.438
Season	-	3	9.99	$3.25e-6^{*}$
Temp:Season	-	3	0.93	0.426
RE:Season	-	3	1.10	0.351
Temp:RE:Season	-	3	1.20	0.309
Nitrous oxide				
Control	-21.18(27.5)	-	-	-
Temperature	27.27(24.6)	1	8.95	<u>0.003*</u>
RE	46.65(25.4)	1	0.56	0.454
T+D	15.29(19.4)	1	1.87	0.174
Season	-	3	16.1	$5.264e-9^{*}$
Temp:Season	-	3	1.38	0.251
RE:Season	-	3	0.274	0.844
Temp:RE:Season	-	3	0.758	0.519
Methane				
Control	-0.174(0.025)	-	-	-
Temperature	-0.137(0.025)	1	0.14	0.709
RE	-0.147(0.026)	1	0.01	0.940
T+D	-0.168(0.021)	1	1.76	0.186
Season	-	3	3.19	0.024*
Temp:Season	-	3	1.19	0.314
RE:Season	-	3	0.77	0.520
Temp:RE:Season	-	3	1.57	0.197

Table 5: Results of ANOVAs for each greenhouse gas. Means are reported in ug per m^2 per day. Standard errors of the means are included in brackets. Significant values are marked with '*'

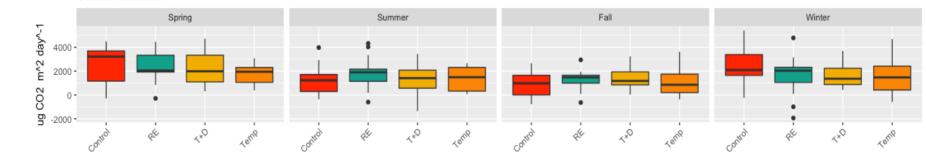
into how climate change stressors differentially impact GHG fluxes depending on the season (Table 6). Summer and fall CO_2 emissions were higher in all treatments than in control plots, while in spring and winter we observed the opposite trend, with higher rates in the control plots (Figure 7A).

Nitrous oxide fluxes were negatively impacted by temperature in spring (Figure 7B). Similarly, the rain exclusion treatment also reduced spring emissions by 67%, despite not being significantly different. Summer N₂O net fluxes rate were negative in the control plot, indicating net fixation under normal conditions. However, our temperature treatment reduced this net fixation by 49%. Furthermore, the rain exclusion treatment increased summer N₂O fluxes by 150%. As a result, the net rates were no longer negative, and the system became a net source of N₂O instead of a sink. In fall, the average flux in the control plots represented the greatest rate of fixation of any plot for the year. However, in the same season, the net N₂O flux became positive under the temperature, rain exclusion, and combination treatments, indicating a shift from fixation to source. Average winter N₂O fluxes were higher in the temperature and rain exclusion treatments than the control plots.

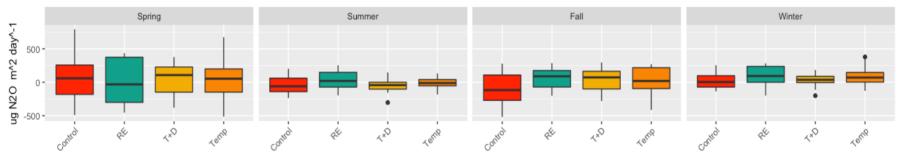
Lastly, the average CH₄ fluxes rates remained negative throughout the year under all treatments (Figure 7C). Control plots tended to show the highest rates of fixation in spring and lowest in summer. In fall, all plots were fixating around the same rate, while in winter the fixation rates under temperature (55%) and rain exclusion (34%) treatments were reduced, as compared to control plots. Table 6: Results of ANOVAs on LMM models of greenhouse gases by treatment and season. Average rates of emission are expressed in ug per m^2 per day, with standard error in brackets. Significant results are marked with '*'.

	Spring			Summer			Fall			Winter		
	Mean F p		Mean F p		Mean F p		Mean	F	р			
CO ₂												
Control	2565.73(362.01)	-	-	1222.66(268.57)	-	-	973.15(263.26)	-	-	2349.72(346.81)	-	-
Temp	1742.80(203.35)	2.59	0.117	1392.98(250.72)	0.17	0.680	1084.53(249.58)	0.32	0.577	1540.12(340.81)	1.91	0.176
RE	2403.46(289.37)	0.34	0.565	1787.36(311.80)	0.82	0.369	1275.79(189.93)	1.80	0.189	1624.20(364.15)	0.98	0.330
$T \!\! + \!\! D$	2274.01(328.00)	1.37	0.250	1369.54(350.93)	0.97	0.329	1409.66(218.98)	0.001	0.975	1590.90(222.99)	1.61	0.212
N ₂ O												
Control	48.91(85.36)	-	-	-40.33(33.23)	-	-	-97.97(56.63)	-	-	13.31(27.35)	-	-
Temp	13.85(70.87)	8.86	0.013*	-19.81(21.70)	3.51	0.073	16.94(51.12)	0.22	0.649	92.86(33.87)	1.29	0.267
RE	16.30(78.22)	0.0004	0.984	25.93(35.40)	1.53	0.228	54.25(36.66)	0.02	0.904	89.80(37.08)	0.009	0.924
$T\!\!+\!\!D$	43.92(54.58)	0.33	0.576	-43.26(27.17)	0.05	0.820	31.34(39.76)	0.48	0.504	26.55(23.04)	3.87	0.060
CH ₄												
Control	-0.253(0.050)	-	-	-0.102(0.049)	_	-	-0.198(0.057)	-	-	-0.146(0.037)	-	-
Temp	-0.224(0.052)	3.46	0.072	-0.140(0.051)	0.85	0.361	-0.111(0.042)	0.07	0.788	-0.066(0.048)	0.01	0.930
RE	-0.249(0.032)	0.93	0.343	-0.129(0.070)	0.54	0.466	-0.111(0.042)	0.05	0.828	-0.097(0.050)	0.38	0.541
$T\!\!+\!\!D$	-0.134(0.046)	1.13	0.295	-0.193(0.047)	0.06	0.813	-0.176(0.042)	3.2	0.081	-0.169(0.037)	3.12	0.086

A Carbon Dioxide



B Nitrous Oxide



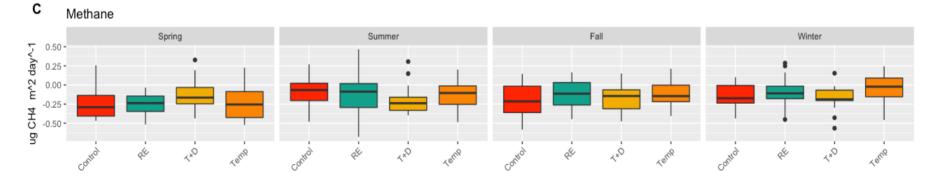


Figure 7: Average greenhouse gases fluxes by treatment in each sampling season.

4 Discussion

Los Alcornocales natural park is recognized as an ecologically, culturally, and economically valuable region and is considered and expected to be at risk under present and future climate change scenarios (Kim et al., 2017; Aronson et al., 2009). The impacts that these climatic changes will have on ecosystem functioning is difficult to forecast but of vital importance to investigate. The main aim of this study was to understand the impact of these projected changes in temperature and precipitation on important ecosystem processes, such as the rates of soil nutrient cycling and greenhouse gas fluxes. Our results demonstrate that these aforementioned alterations will result in significant changes in the availability of nitrogen and phosphorus, as well as in changes in the rate of exchange of greenhouse gases between the soil and the atmosphere, with significant increases in nitrous oxide rates, and potential decreases in sink capacity of methane, and increased emissions of carbon dioxide. Our results also provide valuable and novel information about the seasonal dynamics of these key ecosystem functions and how they will be impacted by future climate change.

4.1 Soil Nutrients Availability

Overall, soil ammonium, inorganic N, and phosphorus production significantly increased, while nitrate significantly decreased, under the rain exclusion treatment. Seasonal dynamics and the interaction with our treatments revealed more insights into how climate change may impact ecosystem functionality.

Spring is an important period for plant growth, thus changes in soil nutrient availability during this time can have important implications for forest functioning (Natalini et al., 2016; Liu, 2010). Inorganic nitrogen production was significantly increased under both rain exclusion and combination treatments, while nitrate was nearly significantly increased (p=0.06) under the combination treatment. It is likely that the increase in temperatures have caused a response in the microbial community under ideal seasonal growth conditions. However, the decrease in water availability due to the rain shelters and increased evapotranspiration (through the increase in temperatures) might have limited the capacity of plants to uptake those nutrients, leading to the observed accumulation of nutrients in the soil (Gessler et al, 2017; Homyak et al., 2017).

Summer in the south of the Iberian Peninsula is already a challenging season for both plant and soil microbial communities, with high temperatures and reduction in precipitation (Costa, 2018; Aronson et al., 2009). Our manipulative intensification in these environmental challenges resulted in a reduction in the availability of all the considered N pools. Our results support previous studies that suggested that intensified summer droughts can reduce nutrient production through alterations in the microbial communities involved in soil nutrient cycling (Bastida et al., 2019; Preece et al, 2019; Ren et al., 2017, 2018; Meisner et al., 2013; Hueso & Hernandez, 2012; Berard et al., 2012).

Interestingly, there was a remarkable increase in the phosphorus availability in fall under the rain exclusion treatment (as well as a similar but statistically nonsignificant increase under the combination treatment), together with an increase in soil ammonium production under both the rain exclusion and temperature treatments. During this sampling period, which occurred in November-December, there was a parallel increase in soil moisture after summer drought conditions (Figure 8), which likely triggered a recovery response in the microbial communities which took advantage of improving conditions and organic matter built up over the summer, resulting in a sharp release of available ammonium and phosphorus.

A response in the N cycle to drought conditions has also been documented by Homyak et al (2017), who found an increase in the soil availability of ammonium and stable levels of nitrate, and suggested the increase being due to the limited ability of plant uptake under drought conditions, and the limitation of nitrification under drought conditions to transform it. They also noted that, in drying soils, large fractions of the nitrified N can escape as gaseous N forms, which could at least partially help explain the decrease we found in nitrate in this study. Another possibility is discussed in a review by Booth et al. (2005), who suggested that decreases of nitrate in relation

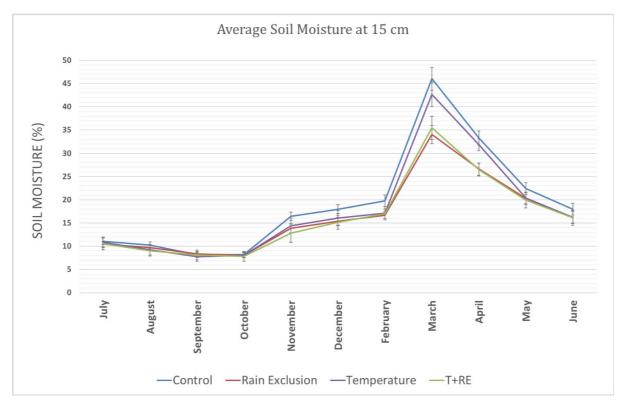


Figure 8: Average soil moisture percentage in the top 15cm of soil of the study site in each treatment over the sampling period.

to increasing amounts of ammonium might also be attributed to competition for space between mineralizing and nitrifying bacteria, as well as to interactions with soil C availability.

Another possible dynamic causing a release in soil nutrients is the so-called birch effect. Soil drying and rewetting events have also been shown to result in a release in C, N, and P as well as pulses of greenhouse gases such as CO₂ (Birch, 1958; Rodríguez et al., 2019; Morillas et al., 2017; Meisner et al 2015; Ouyang & Li, 2013; Jarvis et al, 2005). Thus, the increased availability of inorganic phosphorus and ammonium in fall also could also result from increased phosphorus and nitrogen mineralization processes due to drying-rewetting dynamics. Our field experiment supports previous lab-based studies that showed a release of inorganic phosphorus (Brodlin et al., 2019; Dihn et al. 2017; Bünemann et al. 2013; Butterly et al. 2011) and ammonium (Pezzola et al. 2019; Homyak et al. 2017) after experimental soil dryingwetting.

In summary, our results confirm that forecasted climate change-related alterations in temperature and precipitation (and its combination) might have a significant impact on the production of soil bioavailable nitrogen and phosphorus throughout the year, with noticeable releases in soil available ammonium, but likely with limited uptake by plant communities. These built-up soil nutrients, without a concomitant increase in the capacity of plants to uptake these nutrients (due to lower water availability), can result in significant losses to leaching or gaseous emissions and, on the long term, to the intensification of nutrient limitation of these already limited systems (Homyak et al., 2017; Matais et al 2010; Aronson et al., 2009). Such intensification can lead to stunted growth, and less resilience to further stressors such as prolonged or intensified droughts or pathogens such as *P. cinnamomi*, which is already severely impacting these forests (Costa, 2018; Kim et al., 2017), and even trigger a regime shift in vegetation (Rodriguez et al., 2019; Liu et al., 2018). Together, our results clearly show that soil nutrient cycling processes will not stay the same during more extreme or more frequent future droughts and increases in temperature, which alone and combined can stimulate excessive releases of soil nutrients which can be lost as leachates or gaseous emissions. This confirms that forecasted changes in temperature and precipitation will change key soil processes and put ecosystem functioning at risk for reduced resilience against continuous or additional stressors, such as further climate changes.

4.2 Greenhouse Gases Fluxes

The exchange of greenhouse gases between the soil and the atmosphere were not shown to be as sensitive to changes in temperature and rainfall as soil nutrients transformation rates, with only nitrous oxide showing a significant increase in response to our temperature treatment, particularly in spring. Our inability to detect statistically significant effects for methane and carbon dioxide is likely due to large variations in the response variable values. However, we can still evaluate some ecologically-relevant trends and differences among seasons.

Nitrous oxide can be produced by both denitrification and nitrification processes, depending on soil water content, and is stimulated by higher temperatures (IPNI, 2019; Álvaro-Fuentes et al., 2017; Homyak et al., 2017). Thus, the increases of N₂O emissions in fall and winter can be due to similar reasons as the increase in ammonium and nitrate in spring –the microbial community was stimulated by improving conditions with increased rainfall, and particularly higher temperatures under the temperature treatments. Additionally, emissions of nitrous oxide have also been shown to interact with increases in soil mineral N and C availability (Van Groenigen et al., 2015; Luo et al., 2013). Thus, as N₂O is mainly produced through heterotrophic nitrification in aerobic soils using N from ammonium pools (Butterbach-Bahl et al., 2013), it's possible that the increase in the soil ammonium availability also bolstered the increase in N_2O , particularly under the lower moisture conditions and higher temperatures of our treatments (Van Groenigen et al., 2015). In any case, our results clearly show that the forecasted changes in precipitation and temperature due to climate change can limit the capacity of these systems to uptake atmospheric nitrous oxide, and even transform the system into a net source of this powerful greenhouse gas, representing a paradigmatic example of positive biotic feedback of climate change (Durán et al. 2013).

As expected, our results confirm that these forest function as net sinks of atmospheric methane. However, in general, the amount of methane fixed by these soils was particularly high in the control plots. Our results suggest that our treatments are able to produce significant decreases in methane fixation rates throughout the year. This can be indicative of challenging conditions to methanotrophic bacterial communities (Luo et al., 2013; Ho & Frenzel, 2012). Previous studies showed a strong short-term resilience for methanotrophs under drought conditions (Ho et al., 2016). However, in the long-term, or with more extreme weather events, the structure and function of this bacterial community may be altered (Bodelier & Steenbergh, 2014; Ho & Frenzel, 2012; Mohanty et al., 2007). Our results shed some light on the potential effects of climate change on methanotrophic community. Thus, although a high temporal and spatial variability limited our capacity to consistently find significant effects, the strong and consistent observed trends suggest that under natural conditions continued increases in soil temperature and decreases in soil moisture can indeed alter the capacity of Mediterranean forests to uptake methane, at least in some seasons.

Carbon dioxide fluxes typically increase in response to variations in temperature and rainfall (Canarini et al., 2017; Smith et al., 2018; Oertel et al., 2016; Leifield, 2007). However, in our study, we did not detect this kind of response. This can be due to a few reasons. First, most studies focusing on the effects of climate change on soil respiration (i.e. soil CO_2 emission) have been carried out under controlled, and sometimes drastic lab conditions. On the contrary, our experiment was carried out under field conditions and with realistic changes in soil temperature and precipitation, so the effects in soil respiration may be more subtle or may even take more time to arise. Also, the microbial community in this region is likely to be adapted to cope with subtle and somehow transient (one year) changes in temperature and soil moisture but might still suffer in response to longer-term stressors or more extreme climatic events (Flores-Rentería et al., 2018; Curiel-Yuste et al., 2014). Indeed, this study was conducted only a year or two after the initial setup of the experiment and is only capturing the dynamics of one full year, so further monitoring of the study site may reveal longer-term changes. Likewise, all of soil nutrients and greenhouse gases could have been affected by these reasons as well. Thus, this study highlights that longer monitoring times of climate change field treatments are important to uncover longerterm trends and to account for intra and inter-annual variations.

5 Conclusion

Soil nutrient cycle integrity is important in these already nutrient-limited systems, and as this field study has shown, forecasted increases in temperature and rainfall reduction (and the combination of both) are likely to significantly impact soil nutrient cycles and greenhouse gases emissions. This study has shown that reductions in rainfall particularly impact soil nutrient availability, and possibly causes a decrease in nutrient uptake in plants leading to an accumulation of nutrients in the soil, increasing the risk of nutrient loss by leaching or gaseous emissions. Indeed, this study also demonstrate that increases in temperature and decreases in precipitation might also increase soil nitrous oxide emissions and likely decrease the capacity of this ecosystem to uptake methane from the atmosphere, which could contribute to further global warming.

This valuable cork-oak ecosystem is already under the strain of multiple stressors and understanding future challenges will better prepare managements strategies to mitigate these changes, and to understand what processes will be most damaged by climate change. Our study clearly demonstrates the capacity of climate change to alter soil functioning in these ecosystems. However, further studies on how this study site continues to react to these treatments on the long-term, how tree growth and vegetation dynamics are affected by the treatments, and an exhaustive analysis on the how the microbial communities are changing would be highly valuable and recommendable.

6 References

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7 ANNEX

I. Table summarizing compounds used in lab analyses, the components that were used to make them, and the available lot/batch information. "*" indicates information not available.

Compound/Chemical	Composition	Company	${ m Lot/Batc}$
Potassium Chloride	-	Honewell Fluka	*
Citrate	Trisodium citrate	VWR Chemicals Intl	16B29001 1
	NaOH	VWR Chemicals Intl	16C10001 8
Salicylate	Sodium salicylate	Merck KGaA	K4944580 1
	Sodium pentanitroferricyanacide (III)-dihydrate	Alfa Aesar	Y23B027
Hypochlorite	Sodium phosphate	ITW Reagents	$\begin{array}{c} 000065292 \\ 4 \end{array}$
	NaOH	VWR Chemicals Intl	16C10001 8
	Sodium hypochlorite	ITW Reagents	5Z016250
Sulfuric Acid	-	Sigma-Aldrich	SZBB187 0V
Potassium Nitrate	-	*	*
Ammonium chloride	-	*	*
Devarda's Alloy	-	Merek KGaA	K4783904 1
Ammonium Molybdate	-	*	*
Tin (II) Chloride dehydrate	-	*	*

II. GLS model results for ammonium (NH₄)

gls(NH4.N~OTC*RE*Season, weights=varComb(varPower(form=~fitted(.)|Treatment*Season)), correlation=corCompSymm(form=~1|Tree), method="REML",na.action=na.omit, data=IEMGamir **GLS Model** Variable Estimates **Std. Error P**_{GLS} Treatment + 0.1060 0.0076 0.0000 (Intercept) Season OTC 0.0173 0.0039 0.0505 RE 0.0611 0.0174 0.0005 OTC:RE -0.0411 0.0278 0.1391 Season Spring -0.0317 0.0137 0.0213 Summer -0.0578 0.0080 0.0000 Interactions RE:Summer -0.07490.0173 0.0000 OTC:Spring -0.0563 0.0229 0.0145 OTC:Summer -0.0473 0.0184 0.0105

III. GLS model results for inorganic N (NH₄NO₃)

Model Formula

Model Formula

gls(NH4NO3.N~OTC*RE*Season, weights=varComb(varPower(form=~fitted(.)|Treatment*Season)), correlation=corCompSymm(form=~1|Tree), method="REML",na.action=na.omit, data=IEMGamir)

GLS Model				
Variable		Estimates	Std. Error	P _{GLS}
Treatment +	(Intercept)	0.4118	0.045	0.000
Season	OTC	0.0575	0.067	0.391
	RE	-0.0296	0.054	0.582
	OTC:RE	-0.0287	0.084	0.732
	Season			
	Spring	-0.246	0.048	0.000
	Summer	-0.311	0.045	0.000
Interactions	RE:Spring	0.1683	0.072	0.023

IV. GLS model results for nitrate (NO₃)

Model Formula

gls(NO3~OTC*RE*Season, weights=varComb(varPower(form=~fitted(.)|Treatment*Season)), correlation=corCompSymm(form=~1|Tree), method="REML",na.action=na.omit, data=IEMGamir)

Variable		Estimates	Std. Error	P _{GLS}
v al labic		Estimates	Stu. Error	I GLS
Treatment +	(Intercept)	0.0306	0.0436	0.000
Season	OTC	-0.0077	0.0575	0.894
	RE	-0.0906	0.0495	0.066
	OTC:RE	0.0274	0.0721	0.704
	Season			
	Spring	-0.2164	0.0437	0.000
	Summer	-0.2530	0.0432	0.000
Interactions	RE:Season			

V. GLS model results for inorganic phosphate (PO₄)

Model Formula

gls(PO4.P~OTC*RE*Season, weights=varComb(varPower(form=~fitted(.)|Treatment*Season)), correlation=corCompSymm(form=~1|Tree), method="REML",na.action=na.omit, data=IEMGamir)

GLS Model				
Variable		Estimates	Std. Error	P _{GLS}
Treatment +	(Intercept)	0.0019	0.0003	0.0000
Season	OTC	0.0001	0.0004	0.8034
	RE	0.0038	0.0008	0.0000
	OTC:RE	-0.0010	0.0010	0.3258
	Season			
	Spring	0.0030	0.0007	0.0000
	Summer	-0.0012	0.0003	0.0003
Interactions	RE:Spring	-0.0038	0.0011	0.0006
	RE:Summer	-0.0023	0.0008	0.0084
	RE:Season			

Imer(CO2~	1		× · · · ·	1	1	·
Fixed Effects	var	iable	Estimates	Std. Error	D.F	Pr(> t)
	(Inte	ercept)	969.32	303.65	262.97	0.0015**
	OTC		115.21	416.29	231.50	0.782210
	RE		306.47	417.33	262.96	0.463377
	OTC	C:RE	18.65	579.95	229.86	0.974369
	Seasons					
		Spring	1597.80	422.04	233.31	0.0002***
		Summer	253.34	416.29	231.50	0.5433
		Winter	1380.44	422.04	233.31	0.0012**
Random	Var	iable	Variance	Standar	d Dev.	
Effects	Tree	2	7804	88.34		
	Resi	dual	1467461	1211.39		

VI. LMM results for carbon dioxide (CO₂)

VII. LMM results for nitrous oxide (N_2O)

	1		7		
Fixed Effects	Variable	Estimates	Std. Error	D.F	Pr(> t)
	(Intercept)	11.78	1.71	138.00	2.02e-10 ***
	OTC	-0.26	2.19	138.00	0.907
	RE	-0.0076	2.24	138.00	0.9973
	OTC:RE	-0.074	2.96	138.00	0.9802
	Seasons				
	Spring	5.431752	2.3473	138.00	0.0221 *
	Summer	-2.750228	2.424362	138.00	0.2586
	Winter	-2.616184	2.28571	138.00	0.2544
Random	Variable	Variance	Standard Dev	v.	
Effects	Tree	0.00	0.00		
	Residual	20.57	4.536		

Mixed M	odel				
Fixed	Variable	Estimates	Std. Error	D.F	Pr(> t)
Effects	(Intercept)	-0.199	0.047	256.3	3.28e-05 ***
	OTC	0.087	0.065	231.8	0.1812
	RE	0.088	0.066	256.3	0.1871
	OTC:RE	-0.153	0.091	231.2	0.0960 .
	Seasons				
	Spring	-0.055	0.065	231.8	0.4020
	Summer	0.097	0.064	230.7	0.1331
	Winter	0.0523	0.064	230.7	0.4155
Random	Variable	Variance	Standard D	ev.	
Effects	Tree	0.002674	0.05171		
	Residual	0.037025	0.19242		

VIII. LMM results for methane (CH_4)