

## Accepted Manuscript

Artificial neural network modelling of the antioxidant activity and phenolic compounds of bananas submitted to different drying treatments

Raquel P.F. Guiné, Maria João Barroca, Fernando J. Gonçalves, Mariana Alves, Solange Oliveira, Mateus Mendes

PII: S0308-8146(14)01142-X

DOI: <http://dx.doi.org/10.1016/j.foodchem.2014.07.094>

Reference: FOCH 16160

To appear in: *Food Chemistry*

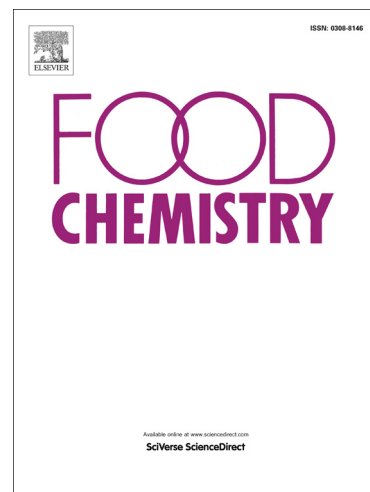
Received Date: 3 April 2014

Revised Date: 17 June 2014

Accepted Date: 17 July 2014

Please cite this article as: Guiné, R.P.F., Barroca, M.J., Gonçalves, F.J., Alves, M., Oliveira, S., Mendes, M., Artificial neural network modelling of the antioxidant activity and phenolic compounds of bananas submitted to different drying treatments, *Food Chemistry* (2014), doi: <http://dx.doi.org/10.1016/j.foodchem.2014.07.094>

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1 **Artificial neural network modelling of the antioxidant activity and phenolic**  
2 **compounds of bananas submitted to different drying treatments**

3  
4 **Running title:**

5 **MODELLING BY ANN OF THE ANTIOXIDANT ACTIVITY AND PHENOLIC**  
6 **COMPOUNDS OF BANANAS**

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10 Raquel P. F. Guiné<sup>a\*</sup>, Maria João Barroca<sup>b</sup>, Fernando J. Gonçalves<sup>c</sup>, Mariana Alves<sup>d</sup>,  
11 Solange Oliveira<sup>e</sup>, Mateus Mendes<sup>f</sup>

12  
13 <sup>a</sup> Polytechnic Institute of Viseu, Research Centre CI&DETS / Dep. Food Industry, Quinta da Alagoa,  
14 Estrada de Nelas, Ranhados, 3500-606 Viseu, Portugal. E-mail: [raquelguine@esav.ipv.pt](mailto:raquelguine@esav.ipv.pt)

15 <sup>b</sup> Polytechnic Institute of Coimbra, ISEC, DEQB, Rua Pedro Nunes, Quinta da Nora, 3030-199 Coimbra,  
16 Portugal. E-mail: [mjbarroca@gmail.com](mailto:mjbarroca@gmail.com)

17  
18 <sup>c</sup> Polytechnic Institute of Viseu, Research Centre CI&DETS / Dep. Food Industry, Quinta da Alagoa,  
19 Estrada de Nelas, Ranhados, 3500-606 Viseu, Portugal. E-mail: [fgoncalves@esav.ipv.pt](mailto:fgoncalves@esav.ipv.pt)

20  
21 <sup>d</sup> Polytechnic Institute of Viseu, Dep. Food Industry, Quinta da Alagoa, Estrada de Nelas, Ranhados,  
22 3500-606 Viseu, Portugal. E-mail: [mgalves.2100@gmail.com](mailto:mgalves.2100@gmail.com)

23  
24 <sup>e</sup> Polytechnic Institute of Viseu, Dep. Food Industry, Quinta da Alagoa, Estrada de Nelas, Ranhados,  
25 3500-606 Viseu, Portugal. E-mail: [sol\\_10890@hotmail.com](mailto:sol_10890@hotmail.com)

26  
27 <sup>f</sup> Polytechnic Institute of Coimbra – ESTGOH / Institute of Systems and Robotics of the University of  
28 Coimbra, Department of Electrical and Computer Engineering, University of Coimbra, Pinhal de  
29 Marrocos - Polo II, 3030 Coimbra, Portugal. E-mail: [mmendes@isr.uc.pt](mailto:mmendes@isr.uc.pt)

30  
31 \*Corresponding author: Raquel P. F. Guiné,  
32 Quinta da Alagoa, Estrada de Nelas, Ranhados, 3500-606 Viseu, Portugal. Tel.: +351232446641, Fax:  
33 +351232426536, e-mail: [raquelguine@esav.ipv.pt](mailto:raquelguine@esav.ipv.pt)

34  
35  
36  
37 **ABSTRACT**

38  
39 Bananas (cv. *Musa nana* and *Musa cavendishii*) fresh and dried by hot air at 50 and 70  
40 °C and lyophilisation were analysed for phenolic contents and antioxidant activity. All  
41 samples were subject to six extractions (three with methanol followed by three with  
42 acetone/water solution). The experimental data served to train a neural network  
43 adequate to describe the experimental observations for both output variables studied:  
44 total phenols and antioxidant activity. The results show that both bananas are similar  
45 and air drying decreased total phenols and antioxidant activity for both temperatures,  
46 whereas lyophilisation decreased the phenolic content in a lesser extent.

47 Neural network experiments showed that antioxidant activity and phenolic  
48 compounds can be predicted accurately from the input variables: banana variety,

49 dryness state and type and order of extract. Drying state and extract order were found to  
50 have larger impact in the values of antioxidant activity and phenolic compounds.

51

52 **Keywords:** antioxidant activity, banana, drying, neural network, phenolic compounds.

53

54

## 55 **1. Introduction**

56 The antioxidant compounds can be defined as substances that in small  
57 concentrations, compared to the oxidizable substrate, significantly delay or prevent the  
58 initiation or propagation of oxidizing chain reactions. These natural chemical  
59 compounds are generally aromatic and contain at least one hydroxyl group and are  
60 called bioactive substances, including, among others, phenolic compounds that are part  
61 of the constitution of various foods. Phenolic compounds are widely present in the  
62 plant kingdom, have simple or complex structures, and are essential for growth and  
63 reproduction of plants, besides being responsible for the colour, astringency and aroma  
64 in several foods (Sharma, 2014). These compounds, being antioxidants, fight free  
65 radicals (Rodrigo & Gil-Becerra, 2014), prevent heart diseases (Jiang, 2014; Khoo &  
66 Falk, 2014), neurodegenerative disorders (Hamaguchi, Ono, Murase, & Yamada, 2009),  
67 circulatory problems (Medina-Remón, Tresserra-Rimbau, Valderas-Martinez, Estruch,  
68 & Lamuela-Raventos, 2014), cancer (Fernández-Arroyo et al., 2012), inflammation  
69 (Wen, Chen, & Yang, 2012), and inhibit lipid oxidation (Maqsood & Benjakul, 2010).  
70 Thermal processing may destroy the amount or the bioavailability of these compounds,  
71 thus reducing beneficial health effects (Agcam, Akyıldız, & Akdemir Evrendilek, 2014;  
72 Al Bittar, Périno-Issartier, Dangles, & Chemat, 2013).

73 Bananas belong to the genus *Musa* from the family Musaceae and are one of the  
74 most popular fruits worldwide. They have a strong ability to protect themselves from  
75 the oxidative stress caused by intense sunshine and high temperature by increasing their  
76 antioxidant levels. Bananas contain vitamins (A, B, C and E),  $\beta$ -carotene and phenolic  
77 compounds, such as catechin, epicatechin, lignin, tannins and anthocyanins (Huang et  
78 al., 2014; Sulaiman et al., 2011), and are notably perishable, as they ripen rapidly  
79 causing significant changes of physicochemical, biochemical and sensory attributes  
80 (Huang et al., 2014). Hence drying represents one of the possible preservation methods  
81 to prevent deterioration and extend the shelf life.

82       Drying is a very ancient way of preserving foods, and is still in use nowadays due to  
83 its ability to inhibit microbial growth and enzymatic modifications, owing to the low  
84 moisture and water activity of the dried products. However, the advantages of drying  
85 surpass the preservation capacity (Guiné, Pinho, & Barroca, 2011). Drying, and  
86 particularly air drying, usually implies an exposure to high temperature for some time,  
87 and that may affect the product properties, either at the physical or chemical levels  
88 (Coimbra, Nunes, Cunha, & Guiné, 2011; Guiné, 2011). Polyphenols, which are  
89 sensitive to high temperatures, may be affected by heat treatment, leading to some  
90 reduction on their content and antioxidant capacity (Ahmad-Qasem et al., 2013).

91       Artificial neural networks have been used in the past years for modelling many  
92 processes in food engineering. Behroozi Khazaeia et al. (2013) used neural networks to  
93 model and control the drying process of grapes. Aghbashlo et al. (2012) used artificial  
94 neural networks to predict exergetic performance of the spray drying process for fish  
95 oil and skimmed milk powder. Kerdpi boon et al. (2006) used artificial neural network  
96 analysis to predict shrinkage and rehydration of dried carrots. Hernández-Pérez et al.  
97 (2004) proposed a predictive model for heat and mass transfer using artificial neural  
98 networks to obtain on-line prediction of temperature and moisture kinetics during the  
99 drying of cassava and mango.

100       The present study was undertaken to investigate the impact of drying conditions on  
101 the total phenolic compounds and antioxidant activity in bananas from two cultivars, as  
102 well as to model the process variables by means of artificial neural networks.

103

## 104 **2. Materials and methods**

105

### 106 *2.1. Sampling*

107       In this work samples from two varieties of banana, *Musa nana* (MN) and *Musa*  
108 *cavendishii* (MC) were used. The bananas were obtained from a local supermarket and  
109 then were peeled and cut into slices 8 mm thick before submitting them to the drying  
110 process. The initial moisture content of the bananas was calculated as an average of  
111 three tests made with a halogen Moisture Analyser (Operating parameters: temperature  
112 = 130 °C, rate = 3). For *Musa nana* the initial moisture content was 67.37±2.65 % (wet  
113 basis), and for *Musa cavendishii* it was 72.32±2.36 % (wet basis).

114

### 115 *2.2. Processing*

116 The convective drying was undertaken in an electrical FD 155 Binder drying  
117 chamber with an air flow of 0.2 m/s and over perforated trays. The samples were dried  
118 until a final moisture content lower than 10% (wet basis) was reached, in order to ensure  
119 good preservation characteristics as well as good final physical and chemical properties.  
120 The drying experiments were conducted at a constant temperature, having been tested  
121 two different temperatures: 50 and 70 °C. The drying of the bananas of cv. *Musa nana*  
122 at 50 °C lasted 525 min and the obtained final moisture content (wet basis) was 9.36%,  
123 whereas the drying at 70 °C was faster, lasting only 270 min and the final moisture  
124 obtained was 4.71%. For *Musa cavendishii* dried at 50 °C the process lasted 450 min  
125 and the final moisture content (wet basis) was 6.37%, while at 70 °C the process lasted  
126 300 min and the final moisture was 8.83%.

127 Lyophilization was made using a Freeze Dryer TDF 5505 (Uniequip, Germany).  
128 The samples were frozen in a conventional kitchen freezer, and then left in the freeze-  
129 drier for 96 hours at a temperature ranging from -52 °C to -49 °C and a pressure 0.7 Pa.  
130 The final moisture content was 2.32 and 2.14 % (wet basis) for *Musa nana* and *Musa*  
131 *cavendishii*, respectively.

132

### 133 2.3. Analysis of total phenolic compounds and antioxidant activity

134 In the present work the extraction of phenolic compounds was performed in  
135 multiple successive steps, namely three times with methanol solutions followed by three  
136 times with acetone/water solutions. This procedure was adopted so as to extract the  
137 highest possible quantity of the phenolic compounds present in the original sample.  
138 Each sample was used to obtain extracts, rich in phenolic compounds, according to the  
139 method described by Soutinho, Guiné, Jordão, & Gonçalves, (2013). Each of the  
140 samples was macerated and successively submitted to multiple extractions: first with a  
141 solution of methanol: three times with acetic acid (98:2), and following with an  
142 acetone/water solution (60:40) also three times. For each of the 6 extractions performed,  
143 the sample was left for 1 hour in an ultrasonic bath at room temperature. This procedure  
144 resulted in three methanol extracts (M1, M2 and M3) and three acetone extracts (A1, A2  
145 and A3).

146 The phenolic compounds were determined by means of the Folin-Ciocalteu  
147 reagent, using gallic acid as a standard, according to the conditions described by  
148 Gonçalves et al. (2012). The results were expressed as milligrams of gallic acid  
149 equivalents (GAE) per gram of dried sample mass. The expression of the results in

150 terms of dry mass instead of whole sample allows the direct comparisons of the results  
151 of the different samples, because in that way the effect of water content was eliminated.

152 The antioxidant activity was determined by the method based on the radical  
153 ABTS (2,2'-azino-bis(3-ethylbenzthiazoline-6-sulphonic acid)), as described by Miller  
154 et al. (1993). The results were expressed as micromoles of Trolox per gram of dried  
155 sample mass.

156

#### 157 *2.4. Artificial neural network modelling*

158 Artificial Neural Networks (ANN) models come from Artificial Intelligence,  
159 where they were first proposed for learning and function approximation. ANNs are an  
160 interconnected assembly of simple processing elements, known as artificial neurons.  
161 Each artificial neuron aims to mimic the functioning of a human neuron. The input for  
162 each neuron is one or more weighted variables, and the output is a linear or non-linear  
163 function of the weighted inputs. Neurons learn by adjusting the weights of the input  
164 variables. Those weights are adjusted in a way to minimise the error between the  
165 neuron's expected output and the measured output value.

166 In the present work, experimental data were modelled using artificial neural  
167 networks, trained and simulated in Matlab™<sup>1</sup>.

168

##### 169 *2.4.1. Data encoding and modelling*

170

171 For ANN modelling, the data were first encoded in a manner suitable for ANN  
172 processing. Variety *Musa nana* was encoded as 1, variety *Musa cavendishii* was  
173 encoded as 2. Banana state values 'fresh', 'dehydrated at 50 °C', 'dehydrated at 70 °C'  
174 and 'lyophilized' were encoded with integers from 1 to 4. Methanol and acetone  
175 extracts were encoded as 1 and 2, respectively.

176 The number of samples available from the experimental data to train and  
177 validate the neural networks was 264 for the output variable 'antioxidant activity' and  
178 277 for the output variable 'phenolic compounds content'. To facilitate training and the  
179 analysis of the results, each output variable was processed separately. This  
180 simplification does not imply any loss of generality, for it is always possible to simulate  
181 a smaller neural network using a larger neural network with sufficient neurons.

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<sup>1</sup> Matlab is a registered trademark of Mathworks. [www.mathworks.com](http://www.mathworks.com).

182 The ANN used was a feed forward model, created using the Matlab fitnet  
183 function in a Matlab script. The ANN used the Levenberg-Marquardt method for  
184 training and the Mean Squared Error (MSE) method for performance assessment.

185 Each network was created with just four inputs (variety, state/dehydration  
186 method, extract type and extract order) and one output, for each of the variables  
187 separately. Each network had just one hidden layer with ten neurons and one output  
188 neuron. There is no general rule accepted for calculating the number of neurons in the  
189 hidden layer, although the common recommendation  $N_{\text{hidden}} = 2/3 (N_{\text{inputs}} + N_{\text{outputs}})$  is  
190 often followed. In the present work, empirical evidence showed that 10 neurons could  
191 produce better results than smaller numbers, without over fitting. Thus, all the results  
192 shown in Table 3 (section 3.4) were obtained using ten neurons in the hidden layer.

193 For training and testing the neural networks, two smaller datasets were created,  
194 one for each output variable. Invalid rows, i.e., rows where there was no valid output,  
195 were removed from the smaller datasets. For each run, the Matlab script randomly  
196 selected 70% of the samples for the train subset and 15% for the validate subset. The  
197 remainder samples were used for the test subset.

198  
199

### 200 3. Results and discussion

201

#### 202 3.1. Phenolic compounds

203 Table 1 shows the amount of phenolic compounds present in the three methanol  
204 extracts and in the three acetone extracts, for all samples at study: fresh, dried at 50 and  
205 70 °C and lyophilized. Regarding variety *M. nana*, the amount of phenolic compounds  
206 present in the first methanol and acetone extracts represented between 52 and 60%, and  
207 between 53 and 76% of the sum of compounds extracted with methanol and with  
208 acetone, respectively. For variety *M. cavendishii*, the amount of phenolics present in  
209 the first methanol and acetone extracts represented between 43 and 69% and between 42  
210 and 64% relative to the total extracted with methanol and with acetone, respectively.  
211 The results showed that the amount of compounds extracted diminished from the first to  
212 the second and again to the third extracts, either in methanol or in acetone, for both  
213 varieties and all stages (fresh or processed). Although most of the compounds were  
214 effectively recovered in the first extraction, the results also show that the first extract  
215 itself would account for an insufficient amount of the phenolics present (42-69%), the

216 second extraction accounting for 13-32% and the third still recovering 11-26%. This  
217 confirms the usefulness of the procedure adopted, by performing successive extractions  
218 with each of the solvents used. The relative percentage of phenolics extracted with  
219 methanol is on average higher (60.0%) than that of the acetone extracts (40.0%). The  
220 type of phenols soluble in each of the solvents tested is different, because phenols  
221 include one or more hydroxyl groups (polar part) attached directly to an aromatic ring  
222 (non polar part). This stereochemistry distinguishes phenols according to their  
223 polarity variance, which influences the recovery of phenols from natural sources, when  
224 accomplished with solvent extraction, being the yield of the process strongly dependent  
225 on the nature of the solvent (Meneses, Martins, Teixeira, & Mussatto, 2013).

226 Flavonoids are in the soluble polar fraction and can therefore easily be extracted  
227 with a polar solvent such as methanol (Risipail, Morris, & Webb, 2005). Methanol has  
228 been generally found to be more efficient in the extraction of lower molecular weight  
229 polyphenols while the higher molecular weight flavonols are better extracted with  
230 aqueous acetone (Dai & Mumper, 2010). However, the solubility of the phenols in  
231 each solvent is very much dependent on the food matrix at study (Michiels, Kevers,  
232 Pincemail, Defraigne, & Dommes, 2012; Tomson, Kruma, & Galoburda, 2012; Zhou  
233 & Yu, 2004).

234 Figure 1 shows the amount of phenolic compounds present in the extracts of  
235 methanol and acetone, as a whole, expressed as gallic acid equivalents (GAE) per gram  
236 of dry matter, for the fresh samples and after the different drying treatments. Looking at  
237 the graph, it can be seen that, in general, the amount of phenolic compounds in the  
238 methanol extracts was higher than in the acetone extracts. The only exception was the  
239 lyophilized sample of the variety *Musa cavendishii*. Considering the total phenolic  
240 compounds quantified in the two groups, the sample *M. nana* fresh had the largest  
241 quantified amount, 6.91 mg GAE/g (dry basis), 60% more than in sample *M.*  
242 *cavendishii* fresh (4.17 mg GAE/g db). These values stand in the same range of those  
243 reported by Sulaiman et al. (2011) for total phenolic content in eight banana cultivars,  
244 varying from 3.98 to 13.00 mg GAE/g dry weight.

245 For variety *M. nana*, the amount of total phenolic compounds of the dried samples  
246 ranged between 3.79 and 6.91 mg GAE/g dry matter, in all extracts. For variety *M.*  
247 *cavendishii*, the amount of total phenolic compounds of the dried samples ranged  
248 between 3.52 and 6.27 mg GAE/g dry matter, in all extracts. The convective drying  
249 originated in all cases a reduction in the total phenolic compounds present, relatively to



250 the fresh sample. For bananas of variety *M. cavendishii* the reduction was 20% at 50 °C  
251 and 15% at 70°C, while for variety *M. nana* the reduction was larger, 43% at 50 °C and  
252 45% at 70°C. The lyophilized samples showed a better preservation of the phenolic  
253 compounds in the bananas of variety *M. nana* (4.71 mg GAE/g dry matter), even  
254 increasing in the case of variety *M. cavendishii* (6.27 mg GAE/g dry matter).

255 Many authors have previously reported that polyphenolics are heat sensitive and that  
256 prolonged heat treatment causes irreversible chemical changes to phenol contents (Lin,  
257 D. Durance, & Scaman, 1998; Mejia-Meza et al., 2008), this being attributed to  
258 different phenomena occurring during heat treatment. According to Martín-Cabrejas *et*  
259 *al.* (2009) and Qu *et al.* (2010), this may be attributed to the binding of polyphenols  
260 with other compounds or to alterations in the chemical structure of polyphenols.  
261 Julkunen-Tiitto and Sorsa (2001) observed a destruction of flavonoids and tannins  
262 during drying. Other authors suggested that another factor contributing to the  
263 degradation of polyphenols may be the activity of polyphenol oxidase, organic acid  
264 content, sugar concentration, and pH (de Ancos, Ibañez, Reglero, & Cano, 2000;  
265 Nicolas, Richard-Forget, Goupy, Amiot, & Aubert, 1994; Yousif, Durance, Scaman, &  
266 Girard, 2000).

267

### 268 3.2. Antioxidant activity

269 Table 2 shows values of the antioxidant activity for the three extracts of methanol  
270 and acetone. The values for the antioxidant activity of the first methanol extract of fresh  
271 samples represent 74 and 65% of the sum of the three extracts, respectively for *M. nana*  
272 and *M. cavendishii*. Dehydration of the samples resulted in a decrease of this value to  
273 about 50%, on average. As to the acetone extracts, the first represents 53% on average  
274 of all samples and both varieties, while the second extract represents 31% and the third  
275 17%. Once again the results show that the procedure of making multiple extractions is  
276 adequate since the last extract still represented 19% and 17% of the total antioxidant  
277 activity measured in the methanol and acetone extracts.

278 Figure 2 shows the antioxidant activity in  $\mu\text{mol Trolox/g}$  (expressed on dry basis)  
279 as determined by the ABTS method in the banana samples subjected to different  
280 treatments. The results show that the antioxidant activity for the fresh bananas, and for  
281 those dried at 50 and 70 °C, is always higher in the methanol extracts than in the acetone  
282 extracts. On the other hand, the lyophilized samples from both varieties show an

283 opposite trend, with the antioxidant activity quantified in the acetone extracts larger  
 284 than in the methanol extracts. These results indicate that the phenolic compounds  
 285 present in the acetone extract of the freeze-dried samples had higher antioxidant activity  
 286 when compared with those present in the methanol extracts. The antioxidant capacity of  
 287 phenolic compounds depends on their conformational chemical structure, namely on  
 288 their ability to donate a hydrogen or an electron as well as on their ability to delocalise  
 289 the unpaired electron within the aromatic structure. For instance, procyanidins are the  
 290 most protective when the oxidant agent is the thermo-labile free radical ABTS (Lotito et  
 291 al., 2000).

292 Comparing the two varieties under study, it was found that the fresh sample of *M.*  
 293 *nana* had an antioxidant activity of 16.0  $\mu\text{mol Trolox/g}$  dry matter, higher than that of  
 294 the *M. cavendishii*, 13.7  $\mu\text{mol Trolox/g}$  dry matter. Sulaiman et al. (2011) reported  
 295 values of antioxidant activity for pulp of eight banana cultivars ranging between 1.12  
 296 and 12.83 mgTE/g dry weight.

297 The convective drying at both temperatures induced a decrease in the phenolic  
 298 compound, up to 40% in the methanol extracts and 22% in the acetone extracts.  
 299 Furthermore, the extension of the reduction in antioxidant activity was higher for  
 300 variety *M. nana* than *M. cavendishii*. Lyophilization did not induce a reduction in the  
 301 total antioxidant activity, so that the values in the lyophilized samples were 14.1 and  
 302 16.4  $\mu\text{mol Trolox/g}$  respectively for *M. nana* and *M. cavendishii*.

303

### 304 3.3. Correlation between antioxidant activity and phenolic compounds

305 The concentrations of the phenolic compounds in the extracts were correlated with  
 306 the antioxidant activity, this being done for the methanol and acetone extracts separately  
 307 and for the whole data together. The results obtained are expressed by the following  
 308 equations:

309 For methanol extracts data:  $AA = 0.4490 + 2.1325 TP$  ;  $R = 0.8258$  (1)

310 For acetone extracts data:  $AA = 0.4472 + 2.9597 TP$  ;  $R = 0.7992$  (2)

311 For all data:  $AA = 0.6563 + 2.1452 TP$  ;  $R = 0.7638$  (3)

312 where AA is antioxidant activity ( $\mu\text{mol Trolox/g}$  dry basis) and TP is total phenols  
 313 content (mg GAE/g dry basis).

314 The results show a good correlation between the two parameters, with correlation  
315 coefficients ranging from 0.7638 to 0.8258, being the correlation stronger in the  
316 methanol extracts than in the acetone extracts. These results are consistent with those  
317 described by different authors that reported a positive correlation between the  
318 concentration of phenolic compounds and antioxidant activity in foods (Katalinić,  
319 Milos, Modun, Musić, & Boban, 2004; Sulaiman et al., 2011).

320 Empirical evidence showed that it is possible to train a neural network, with the  
321 same characteristics as described in Section 2.4 and one neuron in the hidden layer, to  
322 predict antioxidant activity based on the phenolic content with  $R = 0.90$  for the whole  
323 dataset. Predicting phenolic contents from antioxidant activity is a harder problem, as  
324 the neural network with the same characteristics can only predict with  $R = 0.85$ .

325

#### 326 *3.4. Artificial neural network modelling*

327 Table 3 shows the results obtained for function approximation using the neural  
328 networks. Columns 2 to 5 show the R value for the linear regression between the ANN  
329 predicted values and the experimental results, for train, validation and test sets and for  
330 the whole datasets of each variable. Column 6 shows the performance as measured  
331 Mean Squared Error (MSE). Columns 7 and 8 show the average and standard deviation  
332 (STD) calculated for each variable, using the experimental data.

333

334  
335 As the results in Table 3 show, the artificial neural network learns to predict the  
336 phenolic content and antioxidant activity with very high accuracy, approximating the  
337 experimental data with a very small error. Training was performed based on  
338 experimental data, which is not free from noise and outliers. Even so, it is clear from the  
339 results that the neural network was able to abstract an accurate model. Further  
340 comparison of the values predicted by the neural network with the experimental values  
341 showed, for the whole dataset, four errors greater than 75% for the antioxidant activity  
342 and two errors greater than 75% for the phenolic contents. These are with high  
343 probability outliers in the experimental data. This is mentioned just for completeness,  
344 since the impact of those apparent outliers seems negligible, for the present work they  
345 were not removed from the datasets.

346

### 347 *3.5. Neuron weights analysis*

348

349 One interesting characteristic of neural networks is that some information about the  
350 data can be discovered by analysis of the weights of each input. In the present work, a  
351 simplified version of the neural network was implemented. The neural networks were  
352 reconfigured with just one neuron in the hidden layer. Those simplified networks are  
353 not as stable as the networks with 10 neurons in the hidden layer, which showed to be  
354 excellent predictors, as described in the previous section. However, it was still possible  
355 to make the simplified neural networks converge and fit the data with  $R > 0.85$  for most  
356 of the experiments. The advantage of approximating the variables with these simplified  
357 networks is that they have only one weight for each input variable, to weigh the value  
358 fed to the hidden neuron. That weight is a direct indication of the relevance of each  
359 variable to the neuron and, further, to the output function.

360 Table 4 shows the weights of each input for the single neuron in the hidden layer of  
361 the neural networks used, for selected experiments where data was fit with  $R > 0.85$ . As  
362 the table shows, the order of the extract is the most important predictor both for  
363 antioxidant activity and phenolic content. In other words, the variables drying method,  
364 extract type and variety are all less important than extract order. Still, the state is the  
365 second best predictor, which confirms that both phenolic content and antioxidant  
366 activity are greatly affected when the fruits are dried.

367 The weights also show that both banana types are very similar: the weight of the  
368 variety variable is negligible, compared to their state and extract order. Variety is  
369 actually the least important predictor of all.

370 Another important confirmation is that the extract type indeed affects the results,  
371 specially the amount of phenolic contents measured. For the antioxidant activity, it is  
372 the least important predictor, but for the phenolic contents it is more important than  
373 variety by a factor of almost 6.

374

#### 375 **4. Conclusions**

376

377 In general, the phenolic compounds present in all banana samples were recovered  
378 preferentially in the methanol extracts.

379 The results obtained during the present study showed that the drying processes  
380 resulted in bananas with lower phenolic compounds content and antioxidant activity  
381 expressed in a dry basis, when compared with the fresh fruits.

382 The lyophilization process showed to be a drying process that preserves in higher  
383 extent the original properties of fresh bananas than the drying in a ventilated chamber.

384 Neural network modelling showed that the antioxidant activity and phenolic  
385 compounds contents can be predicted with high accuracy from banana variety, drying  
386 state and extract type, using simple neural networks. Antioxidant activity can also be  
387 predicted from the phenolic contents. Neuron weight analysis indicated that the order of  
388 the extract is the most important factor to predict the amount of phenolic contents and  
389 antioxidant activity measured, and both banana varieties show similar properties in the  
390 analysis.

391

#### 392 **Acknowledgment**

393 The authors thank financial support from CI&DETS research centre through project  
394 PROJ/CI&DETS/2012/0001 and FCT (Fundação para a Ciência e Tecnologia) through  
395 project PEst-OE/CED/UI4016/2011.

396

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559 **Tale captions**

560

561 **Table 1.** Phenolic compounds in the different methanol and acetone extracts for both  
562 varieties of banana studied.

563

564 **Table 2.** Antioxidant activity in the different methanol and acetone extracts for both  
565 varieties of banana studied.

566

567 **Table 3.** Results obtained for approximating the variables using neural networks with  
568 ten neurons in the hidden layer.

569

570 **Table 4.** Input variable weights for each variable, obtained for networks with just 1  
571 neuron in the hidden layer and which fit the output variables with  $R = 0.86$ .

572

573

574 **Figure captions**

575

576 **Figure 1.** Total phenolic compounds considering the total among the methanol and  
577 acetone extracts for both varieties of banana studied.

578

579 **Figure 2.** Antioxidant activity considering the total among the methanol and acetone  
580 extracts for both varieties of banana studied.

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**Table 1.** Phenolic compounds in the different methanol and acetone extracts for both varieties of banana studied.

<b>Total phenolic compounds (mg GAE/g dry solids)</b>						
<i>Musa nana</i>						
	<b>M1</b>	<b>M2</b>	<b>M3</b>	<b>A1</b>	<b>A2</b>	<b>A3</b>
Fresh	2.82±0.15	1.07±0.05	0.82±0.03	1.25±0.05	0.69±0.19	0.25±0.03
50 °C	1.25±0.04	0.66±0.04	0.49±0.05	1.17±0.07	0.20±0.07	0.17±0.01
70 °C	1.29±0.10	0.63±0.03	0.47±0.02	0.87±0.04	0.33±0.01	0.21±0.03
Lyophilized	1.57±0.07	0.75±0.01	0.51±0.04	1.00±0.16	0.57±0.09	0.31±0.04
<i>Musa cavendishii</i>						
	<b>M1</b>	<b>M2</b>	<b>M3</b>	<b>A1</b>	<b>A2</b>	<b>A3</b>
Fresh	1.81±0.20	0.44±0.15	0.39±0.10	0.64±0.08	0.49±0.07	0.41±0.10
50 °C	0.85±0.03	0.63±0.05	0.52±0.16	0.79±0.03	0.34±0.08	0.17±0.02
70 °C	1.16±0.02	0.61±0.12	0.44±0.15	0.84±0.04	0.30±0.03	0.17±0.08
Lyophilized	1.66±0.12	0.75±0.18	0.33±0.07	2.04±0.16	0.91±0.10	0.58±0.07

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593 **Table 2.** Antioxidant activity in the different methanol and acetone extracts for both  
 594 varieties of banana studied.

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<b>Antioxidant activity (<math>\mu\text{mol Trolox/g dry solids}</math>)</b>						
<i>Musa nana</i>						
	<b>M1</b>	<b>M2</b>	<b>M3</b>	<b>A1</b>	<b>A2</b>	<b>A3</b>
Fresh	6.8 $\pm$ 0.6	1.5 $\pm$ 0.1	0.9 $\pm$ 0.2	3.6 $\pm$ 0.2	2.6 $\pm$ 0.9	0.7 $\pm$ 0.2
50 °C	3.4 $\pm$ 0.1	1.6 $\pm$ 0.1	1.2 $\pm$ 0.1	3.6 $\pm$ 0.4	1.1 $\pm$ 0.0	0.6 $\pm$ 0.1
70 °C	2.6 $\pm$ 0.3	1.7 $\pm$ 0.1	1.3 $\pm$ 0.2	3.1 $\pm$ 0.2	1.4 $\pm$ 0.1	0.8 $\pm$ 0.1
Lyophilized	3.3 $\pm$ 0.5	2.1 $\pm$ 0.1	1.2 $\pm$ 0.1	3.4 $\pm$ 0.4	2.9 $\pm$ 0.0	1.2 $\pm$ 0.3
<i>Musa cavendishii</i>						
	<b>M1</b>	<b>M2</b>	<b>M3</b>	<b>A1</b>	<b>A2</b>	<b>A3</b>
Fresh	4.8 $\pm$ 0.3	1.4 $\pm$ 0.4	1.2 $\pm$ 0.2	3.2 $\pm$ 0.2	2.0 $\pm$ 0.9	1.2 $\pm$ 0.4
50 °C	2.7 $\pm$ 0.1	2.8 $\pm$ 0.3	2.0 $\pm$ 0.1	2.5 $\pm$ 0.3	1.6 $\pm$ 0.3	0.8 $\pm$ 0.1
70 °C	3.8 $\pm$ 0.2	1.8 $\pm$ 0.1	1.4 $\pm$ 0.2	3.7 $\pm$ 0.2	1.4 $\pm$ 0.2	0.9 $\pm$ 0.3
Lyophilized	3.5 $\pm$ 0.4	2.1 $\pm$ 0.1	1.3 $\pm$ 0.4	3.3 $\pm$ 0.2	3.3 $\pm$ 0.0	2.8 $\pm$ 0.1

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598 **Table 3.** Results obtained for approximating the variables using neural networks with  
 599 ten neurons in the hidden layer.

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	<i>R value for each subset</i>				<i>Train error</i>	<i>Whole Dataset</i>	
	<b>Train</b>	<b>Validation</b>	<b>Test</b>	<b>All</b>		<b>Average*</b>	<b>STD*</b>
Phenolic content	0.99	0.98	0.98	0.99	57.85	56.80	40.51
Antioxidant activity	0.98	0.97	0.97	0.98	721.06	176.01	102.68

601 \*The Average and STD shown refer to all the samples in the dataset and are useful to interpret the train  
 602 MSE shown in column 6.

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606 **Table 4.** Input variable weights for each variable, obtained for networks with just 1607 neuron in the hidden layer and which fit the output variables with  $R = 0.86$ .

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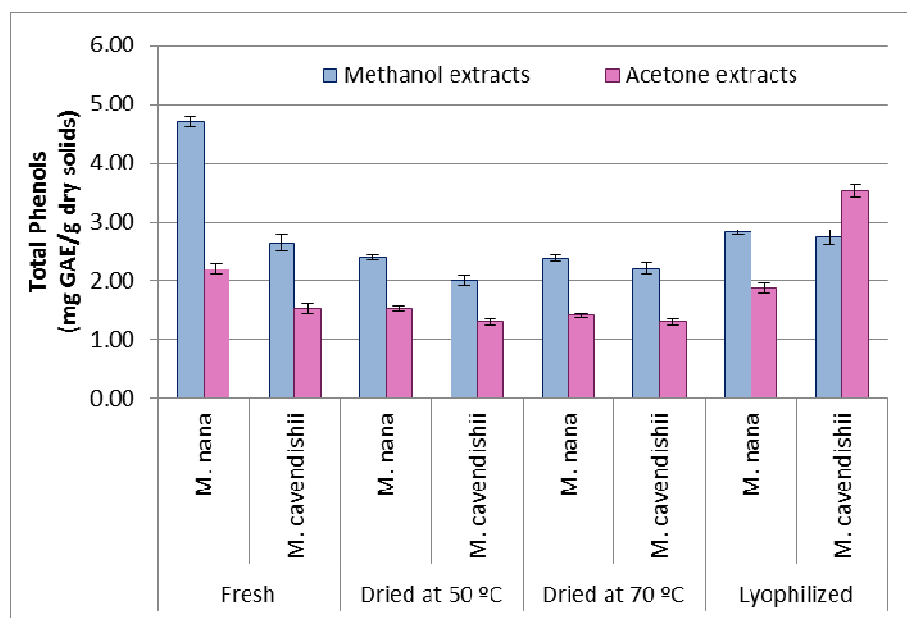
<i>Input variable</i> \ <i>Output variable</i>	<i>Phenolic content</i>	<i>Antioxidant activity</i>
Variety	-0.034	-0.034
State	0.556	-0.669
Extract type	-0.199	0.032
Extract order	-0.923	0.780

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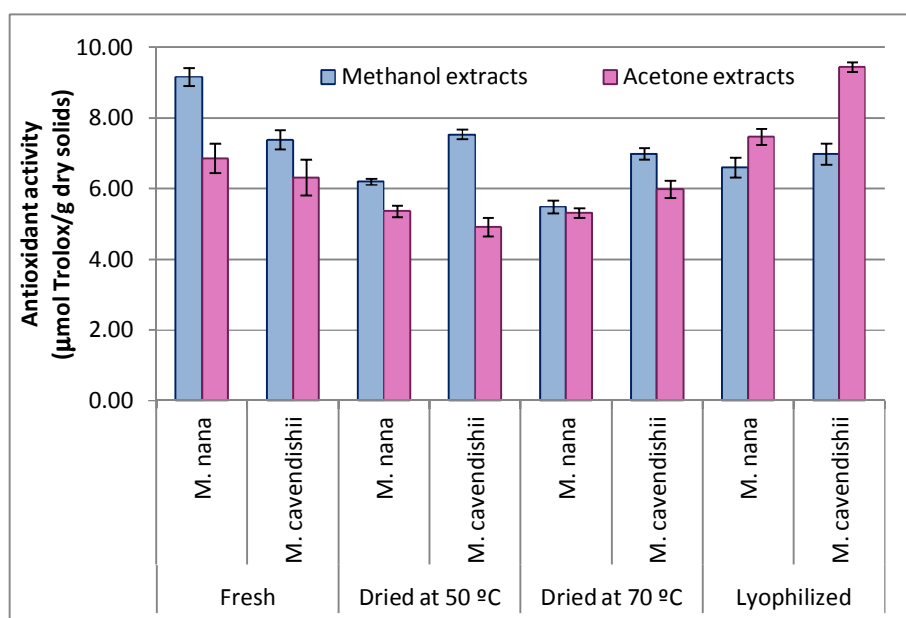
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**Figure 1.** Total phenolic compounds considering the total among the methanol and acetone extracts for both varieties of banana studied.

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**Figure 2.** Antioxidant activity considering the total among the methanol and acetone extracts for both varieties of banana studied.

628 **ARTIFICIAL NEURAL NETWORK MODELLING OF THE ANTIOXIDANT**  
629 **ACTIVITY AND PHENOLIC COMPOUNDS OF BANANAS SUBMITTED TO**  
630 **DIFFERENT DRYING TREATMENTS**

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633

- Bananas fresh and dried were analysed for phenols (TP) and antioxidant activity (AA)

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635

- Different consecutive extraction solutions were used

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- The data trained a neural network (ANN) for data analysis and variable prediction

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- Phenols and antioxidant activity decreased with drying for all treatments

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- ANN showed that TP and AA can be predicted from the input variables tested

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