



Review

The Lethal and Sub-Lethal Effects of Fluorinated and Copper-Based Pesticides—A Review

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Abstract: In recent decades, pollution levels have increased, mainly as a result of the intensive anthropogenic activities such industrial development, intensive agricultural practices, among others. The impact of metals and organic contaminants is, nowadays, a great concern to the scientific and political communities. Copper compounds are the main sold pesticides in Europe, as well as herbicides, including glyphosate. Diphenyl ethers are the second ones most sold. Glyphosate and copper compounds are intensively studied, but the opposite is seen in the case of diphenyl ethers, including fluorinated pesticides (e.g., oxyfluorfen). Some research has been performed to increase the knowledge about these contaminants, daily inputted on the aquatic systems and with dangerous effects at physical and biochemical levels on the organisms. A wide range of biomarkers (e.g., growth, survival, reproductive success, enzymatic activity, lipid metabolism) has been applied to determine the potential effects in many species. This review intends to: (a) perform a compilation of the knowledge in previous research about the action mode of organic (fluorinated-based herbicide) and inorganic (copper-based pesticides) contaminants; (b) carry out an information survey about the lethal and sub-lethal effects of the fluorinated-based pesticides, namely the oxyfluorfen and the copper-based pesticides, on aquatic species from different trophic levels, according to in vitro and in vivo studies; (c) understand the impact of oxyfluorfen and copper-based pesticides, considering their effects reported in in vitro studies and, simultaneously, the authorized concentrations by legal organizations and the effective concentrations of each pollutant found in the environment. The literature analyzed revealed noxious effects of Cu and oxyfluorfen to aquatic organisms, including freshwater and marine species, even when exposed to the reference as well as to environmental concentrations, thus highlighting the importance of more monitoring and ecotoxicological studies, to chemical pollutants and different species from different ecological niches, to sustain and improve the legislation.

Keywords: copper sulphate; oxyfluorfen; ecotoxicological effects; aquatic species; biomarkers



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1. Introduction

Human populations are increasing and, consequently, increasing the food production need. To suppress the production needs, pesticides and fertilizers are widely used in the agricultural practices [1], and according to the CAS (Chemical Abstract Service), there are more than eighty-nine million reported chemical compounds. Daily, an increase in the number of pollutants from anthropogenic activities on the aquatic systems is reported due to the input of novel products [2,3]. The trace concentrations together with the large number and types of pollutants represent a challenge in recognition and remediation [4].

According to [5], in the last 3 decades, pesticides use has increased about 1 million tons. In addition, the Eurostat agency reported 310,739 tons of sold pesticides in Europe in 2019, with the main pesticides categories being fungicides and bactericides (39.81%), and herbicides, haulm destructors, and moss killers (31.75%). Moreover, among the fungicides

and bactericides, the greatest part is from inorganic origin (54%), including mainly copper compounds, as well as inorganic sulfur and other inorganic fungicides. The herbicides, haulm destructors, and moss killers category was composed mostly of 'Other herbicides' (61.8%), including glyphosate (a well-studied pesticide) and fluorinated herbicides [6], a group with an increasing application, as it is seen as an alternative to control glyphosate-resistant weeds [7].

Fluorinated pesticides may act by different modes on the organisms: (a) inhibition of the acetolactate synthase, an enzyme with an important role on the synthesis of ramified chain amino acids [8,9]; (b) inhibition of Acetyl-CoA Carboxylase, which catalyzes the first step of the fatty acids' biosynthesis on plants [10,11]; (c) inhibition of mitosis and consequent cell division [10]; (d) inhibition of protoporphyrinogen oxidase or 'Protox', a key enzyme that catalyzes the oxidation of protoporphyrinogen IX into protoporphyrin IX, essential on the chlorophyll biosynthetic pathway [10,12,13]; (e) inhibition of 4-hydroxyphenylpyruvate dioxygenase, an enzyme involved in the tyrosine biosynthesis, affecting the chlorophyll production [14,15]; (f) inhibition of very long-chain fatty acids elongase, with effects on the biosynthesis of these fatty acids [16–18]. According to the different action modes of these pesticides, detrimental effects such as inhibition of photosynthesis and synthesis of photosynthetic pigments, amino acids synthesis inhibition, and changes in the fatty acids and protein metabolism have been reported [19–23]. Thus, the fatty acids and amino acids determination, as well as the assessment of lipid peroxidation and photosynthetic pigments, may be valuable biomarkers on the monitoring and health status of the aquatic organisms.

Some researchers have also been dedicated to understanding the copper action mode. Currently, three processes have been reported that may explain the mode of action of copper ions—(a) inactivation of proteins due to the interactions between copper (II) and thiol-, imidazole-, and carboxyl-groups of amino acids; (b) formation of copper (I), through the interactions between copper (II) and deoxidants; the copper (I) may act as a catalyzation agent on the formation of hydroxyl radicals, a reactive oxygen species (ROS); (c) copper (II) and copper (I) may replace essential cations from specific binding sites [24,25]. Consequently, several detrimental effects on the photosynthesis process, fatty acids metabolism, carbohydrates synthesis [18,26–29], cell respiration, ATP production, pigments synthesis, and inhibition of cell division [30] have been associated with the exposure at high copper concentrations, with consequences on the function and structure of the cell membrane, loss of key nutrients, and metabolic imbalance [31].

UNESCO reported an input of pollutants into the aquatic systems of 730 million tons per year [32]. Marine systems have a key role in human life, as these systems provide food sources and natural resources with application in different sectors such as pharmaceutical, cosmetic, agriculture and animal feeding, and recreational and professional activities. However, these systems are often under anthropogenic pressures such as sewage water discharges, input of contaminants from industrial and agricultural activities, and plastic pollution. These continuous anthropogenic pressures affect the water quality and, thus, the aquatic communities [33].

First, changes in the structure or function of microbial communities, which is very sensitive to the changes in the water quality, may affect the entire aquatic ecosystem [34]. The quality and abundance of bacterial communities, and micro- and macroalgae depend on the environmental conditions [35]. These organisms are at the base of the trophic chain acting as primary producers, presenting a key role in the trophic chain. Zooplankton acts like links between primary producers and secondary consumers. The exposure to stressors may affect its nutritive value, with effects on the lipid metabolism [24]. In the case of bivalves, which are greatly appreciated as a food source by human beings, chemical stressors may affect greatly their behavioral (feeding, growth, reproduction, cardiac activity, and maturation) [36] and the biochemical pathways (decrease in the ATP; lipid peroxidation and cell glutathione imbalance; inhibition of respiratory process; changes in the enzymatic activity, proteins synthase, and lipids profile) [37]. Fish are the first source of highly unsaturated fatty acids to the human population. These organisms are greatly

affected by metals, such as copper, and organic contaminants, such as fluorinated pesticides, with consequences on the enzymatic activity, oxidative stress, and changes in the lipid metabolism and fatty acids profile with a decrease in its nutritive value (decrease in omega-3 and -6, and increase in saturated fatty acids) [27]. Considering the increased production and input of the pollutants into the environment, as well as the several adverse effects on the ecosystems and communities, it becomes very important to monitor and to diagnose the ecosystems exposure to these organic and inorganic pollutants by applying biomarker tools.

According to the studies conducted thus far about both chemicals, it is possible to verify that the exposure to these compounds comprises effects on the behavior (e.g., with alteration in the growth), lethal [18,20,29,38–44], and biochemical levels (e.g., with changes in the enzymatic activities, lipid metabolism, and proteins expression) [18–23,29,41,44–54].

Metals and organic pollutants have been reported as inducers of reactive oxygen species (ROS) formation [55,56]. ROS are the group of superoxide, hydroxyl, and non-radical oxygen derivatives (e.g., hydrogen peroxide and singlet oxygen) [57]. When the antioxidant system does not act efficiently against the ROS action, the organisms suffer oxidative stress caused by biological, chemical, and physical stress [58]. Thus, to fight the oxidative stress, aerobic cells have developed antioxidant defense systems or a redox balance [59]. Among several mechanisms and agents of antioxidant defense, metallothioneins (MTs), catalase (CAT), glutathione peroxidase (GPx), glutathione S-transferase (GST), superoxide dismutase (SOD), species reactive of thiobarbituric acid (TBARS), and cholinesterase (ChE) are mostly the biomarkers used in the environmental assessment [60–62]. Moreover, reduced glutathione (GSH) may act as a substrate of GST and also takes part in the xenobiotics conjugation and detoxification [61]. Lactate dehydrogenase (LDH) plays a key role in the anaerobic metabolism [63,64]. ROS action, when it is non-controlled, may have deleterious effects on pigments, proteins, lipids, DNA, RNA, ribosomal synthesis, and enzymatic systems [65,66]. The induction of enzymatic or non-enzymatic antioxidant defenses can provide relevant information on the study of the effects of chemical compounds on many organisms [67]. Thus, biomarkers may be defined as powerful tools and endpoints in toxicological studies to assess the organisms' health status and consequently to detect changes and stress conditions on the ecosystems.

Fatty acids (FAs) and carbohydrates have also been used as biomarkers to assess the organisms' health status [24,68] and exposure to stress conditions, such as organic and inorganic pollution, by copper and fluorinated pesticides [18,22,29,35,41,45,52].

The determination of changes in the fatty acids profile, such as inhibition of elongase of very long-chain fatty acids induced by fluorinated-based pesticides, or effects on the esterification, desaturation, and mobilization of triacylglycerols associated with copper exposure [69], is revealed to be an important point in the assessment of the aquatic organisms health status, as fatty acids are essential components in the cell membrane with crucial physiological functions (e.g., permeability). Moreover, FAs are one of the main lipid constituents of the cell membranes and are used as fuel for the metabolic processes at all trophic levels [70]. Furthermore, FAs are the most important molecules transferred along the trophic chain (from primary producers to consumers) and, as the animals are not able to synthesize some of them, the so-called essential FAs (EFAs) are essentially acquired by food sources. Thus, FAs have been used as trophic markers [29,71–73] and as qualitative markers allowing for determining predator–prey relationships [74–76].

Some researchers have also reported effects on carbohydrates composition to different organisms, because of copper and fluorinated pesticides exposure. However, the response depends on the organisms. There are studies that report a decrease in carbohydrates to *Eisenia foetida* [77] and to *Chaetoceros calcitrans* and *Nitzschia closterium* [78], as a consequence of the exposure to low concentrations of a fluorinated herbicide (oxyfluorfen). Other studies highlight an increase in the glucose levels on *Thalassiosira weissflogii* after exposure to copper and oxyfluorfen [18] and the accumulation of non-structural carbohydrates, at high levels of copper on *Citrus grandis* and *Citrus sinensis* [79]. Carbohydrates have

a key role in vital functions, such as in cell homeostasis maintenance in the immunity defense process. In addition, they are the main and speedy energy source to deal with stress conditions. However, pollutants may cause great energetic costs to the immunity balance [80]. Moreover, changes in these biomolecules content or storage may affect several cell functions, such as carbon fixation, lipid accumulation, or photosynthetic efficiency [81]. Polysaccharides are carbohydrate polymers composed of long-chain monosaccharides and they have been reported as protectors of the bacterial cells against metals, organic compounds, and many other environmental perturbations [82–85]. Therefore, this review aims to: (a) perform a collection of the literature about the action mode of organic (fluorinated herbicide) and inorganic (copper pesticides) contaminants; (b) assess the relevance of the lethal and sub-lethal effects of the fluorinated-based pesticides, namely the oxyfluorfen and the copper-based pesticides, on aquatic species from different trophic levels, reported by *in vitro* studies, but considering the existent environmental data; (c) understand the danger of oxyfluorfen and copper-based pesticides to aquatic systems, considering their effects reported in *in vitro* studies and, simultaneously, the authorized concentrations by legal identities and the concentrations of each pollutant found in the environment. Moreover, this review highlights the toxicological and biochemical effects as a consequence of the fluorinated and copper-based pesticides already studied in several species along the trophic food chain and ecological niches. It also emphasizes the harmful effects of these pollutants on the ecosystems, once concentrations reported in the environment are higher than those used in *in vitro* bioassays, which have had dangerous effects on freshwater and marine organisms, such as algae, zooplankton, macroinvertebrates, and fish species.

2. Methodology

This review article performs a compilation of the information from 236 sources. The literature review was based on the main keywords: pesticides, herbicides, organic pollutants, inorganic pollutants, metals, fluorinated based herbicides, oxyfluorfen, copper, aquatic organisms, toxicological effects. Some research platforms were used, namely Google, Google scholar, B-on, Science Direct, Pesticides Data Base, European Commission site, the USEPA site, and the World Health Organization site. Moreover, a literature search for privileged recent works from 2000 to 2023 was performed; however, some knowledge bases crucial to this work clarification required the use of older sources, namely studies since 1981.

3. Organic Contaminants—A General Characterization

Many works have reported the organic contaminants as an increasing concern given its deleterious effects on the environment [86–88]. Numerous organic contaminants are reported as hardly biodegraded and lipophilic compounds, therefore, have a great bioaccumulation ability [86,89,90]. Moreover, these contaminants are described as mutagenic, carcinogenic, endocrine disruptors, immune system depressors, reproductive impairment originators, hyperthyroidism causers, neurotoxic, and responsible for changes in skeletal growth and ontogenetic development [86,89–91].

Organic pollutants include phenolic compounds, agrochemicals (organic pesticides and herbicides), industrial chemicals (e.g., polychlorinated biphenyls—PCBs), and products from industrial processes (e.g., polychlorinated dibenzo-p-dioxins—PCDDs, polychlorinated dibenzofurans—PCDFs, and polycyclic aromatic hydrocarbons—PAHs) [86–88,92]. Many of these compounds are persistent, designated as persistent organic pollutants (POPs). POPs are chemical substances that, due to its persistence, may bioaccumulate through the trophic chain and be carried and reach other systems far from its source, with consequences on the environmental and public health at the global level, even in places that have never produced or been used [27].

3.1. Pesticides

Pesticides comprise a large spread of environmental contamination. The human population is increasing exponentially, and the fertilizers and pesticides used have become

essential to the agriculture practices, in order to suppress the food production needs. However, the excessive use of these compounds may comprise deleterious effects to the ecosystems (e.g., biodiversity loss and damage to public health) [93,94] and has become unescapable on freshwater and marine ecosystems [95]. Some studies about pesticides have been developed, as one of the main harms of this chemicals is its non-selectivity, disturbing non-target organisms and consequently the normal function and maintenance of the terrestrial and aquatic systems [96]. Moreover, when applied in soils and plants and depending on the persistence on the environment, the pesticides are subjected to many processes that may involve degradation and/or conveyance by drift, leaching, and run off to several compartments (soil, water, sediment) [97].

According to the World Health Organization (WHO), pesticides are defined as chemical compounds whose main aim is to kill pests, including insects, fungi, rodents, and weeds [98]. Nonetheless, according to the Environmental Protection Agency of the United States (USEPA), pesticides are any substance used to prevent, destroy, repel, or mitigate any pests, with pests being characterized as any animal, plant, or microorganism that affects human food, health, or wellness [99].

Pesticides may be classified according to its target (herbicides, fungicides, insecticides, repellents, avicides, acaricides, bactericides, viricides, etc.), chemical composition of the active ingredients (carbamates, organochlorine, organophosphorus, pyrethrin, pyrethroids, etc.), and according to the risk (extreme danger, high danger, moderate danger, and slight danger). The exposure pathways may be by ingestion (of contaminated water and food), inhalation (of contaminated water, dust, soil, and industrial vapors), and dermal contact [100].

Several pesticides are recognized as endocrine disruptors—exogenous substances (natural or synthetic) that affect the endocrine system and may cause damage to the physiological pathways of the organism and/or on the next generations, with deleterious effects on the development, growth, and reproduction. As the transcriptional activity of nuclear receptors is one of the first targets of the endocrine disruptors, many pesticides (organochlorine, diphenyl ethers, organophosphorus, carbamates, acid amines, ureas, pyrethroids . . .) were designed to act against these nuclear receptors [101].

Pesticides exposure is associated with some diseases in humans such as Hodgkin's disease, non-Hodgkin lymphoma [102,103], Parkinson's [104,105], endocrine disruption [106,107], breath and reproductive problems [108], lung cancer [109], and brain tumors [110]; damage to the lymphatic tissues, liver, thyroid, uterus, and mammary gland (according to animal tests of low concentrations exposure—0.1 ppm of Dieldrin) [111]; increased risk of prostate cancer [112].

3.2. Herbicides—A Main Concern

Herbicides are the most used pesticides, responsible for 40% of the world's pesticides production [113], followed by insecticides, fungicides, and other pesticides [99]. One of the greatest apprehensions about the contamination by herbicides is due to its ability of bioaccumulation on primary producers and consequent proliferation along of the trophic chain [114].

According to the Weed Science Society of America (WSSA), the herbicides may be classified into 28 groups consistent with their action mode, which often includes target metabolic enzymes, carriers, hormonal regulators, cofactors, proteins, or other cell biomolecules [115]. The herbicides action mode has a key role on its effectiveness. Furthermore, the damage severity and quickness are linked to the herbicide power against the target, as well as the target biological relevance [116].

3.2.1. Fluorinated-based herbicides

Fluorinated organic products are present mostly on the agrochemical products [117–119]. About 25% (56/229) of compounds have at least one fluorine atom, often present as substituents aryl-F, aryl-CF₃, and aryl-OCF₃ [120]. In organic chemistry, the C-F bonds are the strongest and fluorine is able to bind covalently to the next smallest carbon atom.

Therefore, the change from F to H has a negligible steric perturbation and leads to a stable derivative. The polarity linked to C-F bonds [121,122] will bring dipoles with consequent changes to the conformation, leading to a better bonds target. Moreover, the incorporation of fluorine may modify the surrounding protic functional groups acidity, namely of the groups OH, NRH, and CO₂H. Thus, considering that many fluorine substituents usually increase the lipophilicity and the molecules bioavailability to cross the cell membranes, and specific sites of the fluorine or fluorinated groups may be used in the protection against or suppress in vivo metabolism, fluorine may comprise several effects such as inhibition of biological processes (e.g., cell division, fatty acids biosynthesis including very long-chain fatty acids), inhibition of chlorophyll production, or further degradation, leading to leaf bleaching or generation of oxygen singlets and overproduction of oxygen reactive species, resulting in lipid peroxidation, which supports the development of compounds with a great efficacy [1].

The fluorinated-based herbicides can be classified according to its action mode, as different herbicides may act on different targets, as described in Table 1.

Table 1. Fluorinated-based herbicides and its action modes.

Inhibitors Type	Target Characterization	Action Mode	Chemical Compounds	Aim/Application
Acetolactate Synthase (ALS) Inhibitor	ALS is a flavin enzyme involved in the biosynthesis of the ramified chain of the amino acids L-valine, L-leucine, and L-isoleucine [10,15]	Act over the ALS, leading to the decrease in the synthesis of the amino-acids-ramified chain, essential to the early tissues growth [8,9]	Sulphonamides; Flumetsulam; Florasulam; Penoxsulam; Piroxsulam.	Fight against broadleaf weeds on wheat crops and other cereals [123]
Acetyl-CoA Carboxylase (ACC) Inhibitors	ACC catalyzes the 1st step of the fatty acids' biosynthesis on plants. ACC catalyzes the conversion from Acetyl-CoA to Malonil-CoA—with a key role on the saturated fatty acids assembly in the plastids.	ACC inhibition prevents the fatty acids biosynthesis and drains the Malonil-CoA levels in the cell to the additional elongation of SFAs, when they are transported from the plastids to the cytosol [10,11] Chemicals enter the cell and establish a link with tubulin, causing disruption to the microtubules formation and, therefore, inhibition of mitosis [10] Auxins are applied and adsorbed by the leaf and quickly distributed through the plant.	Clodinafop-propargil; Fluazifop; Fluazifop-P-butyl; Haloxifop.	Cereal crops
Mitosis Inhibitors	Cell division	Trifluralin; Ethalfuralin.	Prevent the growth of weeds on the crops	
Synthetic Auxins	Plant hormones	They induce auxins as a response, leading to the atypical development of morphologies [124].	Fluoroxipir	
Protoporphyrinogen Oxidase (PPO) or 'Protox' Inhibitors	PDSs are involved in the carotenoids biosynthesis; Catalyzes the conversion from phytoene to carotene and phytofluene in plants	PDS inhibition causes chlorophyll degradation and consequent bleaching of the leaves [125,126]	Diflufenican	
	PPO is a key enzyme that catalyzes the oxidation of protoporphyrinogen IX into protoporphyrin IX, an intermediate essential to the chlorophyll biosynthetic pathway [10,12] and to the chloroplasts, and then it is distributed through the chloroplast membrane [127]	PPO inhibitors block the protoporphyrin production; Accumulation of protoporphyrinogen on the chloroplasts; Leakage of protoporphyrinogen to the cytosol and non-enzymatic oxidation; Massive production of singlet oxygen and lipid peroxidation [13]	Oxifluorfen; Fomesafen; Lactofen.	
4-Hydroxyphenylpyruvate dioxygenase (HPPD) Inhibitors	HPPD is an enzyme involved on the tyrosine biosynthesis; HPPD converts tyrosine into homogentisate, a metabolic precursor to plastoquinone and tocopherol in plants.	HPPD inhibitors affects the chlorophyll production, causing bleaching [14,15]	Isoxaflutole; Pyrasulfatole; Tembotrione.	Fight against broadleaf and grass weeds on rice and maize farms.
Very Long-Chain Fatty Acids (VLCFAs) Elongase Inhibitors	VLCFAs have an essential importance in the formation of glycosylphosphatidyl-inositol anchors and sphingolipids [128].	Inhibition of the elongase that catalyzes the very long-chain fatty acids biosynthesis [16,17]	Flufenacet.	Prevent weeds on cereals and corn crops.

3.2.2. Oxyfluorfen and Its Effects

Oxyfluorfen was introduced by Dow in 1976 and has been widely used in rice crops due to its high efficiency as a herbicide and low application range. However, the intensive use may affect environmental and human health [129]. The oxyfluorfen application has increased on pre-emergence and early post-emergence, as, nowadays, this non-selectivity and the broad-spectrum herbicide are seen as an alternative to control the glyphosate-resistant weeds; however, the decrease in oxyfluorfen efficiency is also expected [7]. According to ECHA [130], maximum limits of oxyfluorfen in different food types are defined from 0.05 mg L⁻¹ to 0.1 mg L⁻¹; however, environmental studies have reported higher environmental concentrations, up to 23.6 mg L⁻¹ in Nile River, Egypt [131].

This diphenyl ester, such as all herbicides of fast performance that act by inhibition of PPO activity, belong to group 14 of the WSSA [116]. The PPO activity inhibition and consequent accumulation of protoporphyrinogen in the chloroplasts with leakage to the cytosol, which is non-enzymatically oxidized in protoporphyrin, lead to a massive production of singlet oxygen (Table 1), and hydrogen of the unsaturated lipids is extracted; then, lipid radicals are produced, and a chain reaction of lipid peroxidation begins. Proteins and lipids are oxidized with peroxidation damage on the cell membrane and membrane permeability disruption, causing fast disintegration of organelles and cells and, in the last state, cell death [132–138].

According to our knowledge, few studies have been dedicated to the evaluation of oxyfluorfen's effects on the different species, so there is a lack of information in this field mainly in marine systems. Among oxyfluorfen's known effects, there are the inhibition of photosynthetic pigments, photosynthetic electrons transport, acetylcholinesterase activity, growth, and phosphate and nitrogen fixation ability; induction of superoxide radicals and hydrogen peroxide levels, and activity of antioxidant enzymes and the stress protein Hsp 70; changes in fatty acid profiles with the increase in saturated and monounsaturated fatty acids content. Table 2 shows a compilation of the effects already reported in some studies to freshwater organisms, such as inhibition of photosynthetic pigments and activity and growth rate in microalgae species, as well as changes in enzymatic biomarkers response in microalgae and fish species, and changes in the nutritive value of fish species [19–23]. Moreover, considering the great oxyfluorfen concentrations above reported for aquatic systems, the concentrations used in the in vitro studies and the observed effects represent a major concern, with real impacts to the aquatic organisms and consequently to the ecosystems function and structure.

Table 2. Effects of the oxyfluorfen on freshwater species evaluating parameters, such as growth inhibition, photosynthetic and enzymatic activities, and fatty acids profile.

Concentration/Duration	Species	Effects	References
0–20 µg/mL (72 h)	<i>Nostoc muscorum</i>	Significant inhibition of the photosynthetic pigments concentrations (chlorophyll a, carotenoids, and phycocyanin) and photosynthetic activity at all treatments; Significant inhibition of the photosynthetic electrons transport activity (PS II and whole chain) to both treatments; Significant growth inhibition; Significant decrease in the ability of NO ₃ ⁻ and PO ₄ ³⁻ fixation;	[19]
	<i>Phormidium foveolarum</i>	Significant reduction of the acid phosphatase, alkaline phosphatase, and nitrate reductase to both treatments, except on <i>P. foveolarum</i> , which reported a significant increase in the nitrate reductase activity to both treatments. Significant induction in the superoxide radicals and hydrogen peroxide levels; Changes in the antioxidant enzymatic activity (catalase—CAT, superoxide dismutase—SOD, and peroxidase—POD).	

Table 2. Cont.

Concentration/Duration	Species	Effects	References
0–30 µg/L (24 h)	<i>Scenedesmus obliquos</i>	Significant growth inhibition at 20 µg/L, with IC ₅₀ = 15 µg/L; Significant induction of the enzymatic activity: Glutathione reductase up to 53% at 22.5 µg/L; Glutathione S-transferase until 76% at 22.5 µg/L; Ascorbate peroxidase up to 29% at 22.5 µg/L; Catalase until 96% at the same concentration (22.5 µg/L).	[20]
4.3 mg/L (6 days)—acute bioassay	<i>Gambusia affinis</i>	Significant inhibition of acetylcholinesterase (AChE) activity (36.7%—2 days and 13.2%—6 days);	[21]
1.43 mg/L (15 days)—sub-acute bioassay		Significant inhibition of AChE activity between 15.7% (5 days) and 30.64% (15 days);	
0.43 mg/L (30 days)—chronic bioassay		Significant inhibition of AChE activity between 24.5% (10 days) and 25.17% (20 days); Non-significant inhibition of AChE activity at the end of 30 days (decrease of 20.22%).	
0–0.6 mg/L (21 days)	<i>Oreochromis niloticus</i>	Significant increase in the liver total protein content at the lower concentration (0.3 mg/L), with no significant changes at 0.6 mg/L; Significant increase in the CAT activity to both treatments at 7 and 14 days, and at 21 days to the lower concentration; Opposite trend of SOD activity that showed a significant activity inhibition for both treatments on the different days; Significant induction of GR activity; Significant induction of GST activity at 7 days, followed by a significant inhibition at 14 and 21 days; Changes in the fatty acids' profiles to both treatments, with the most abundant being C16:0 and C18:0 (saturated fatty acids) and C18:1 and C24:1 (unsaturated fatty acids).	[22]
3 mg/L (6 days)	<i>Oreochromis niloticus</i>	Significant inhibition of AChE activity up to 54.5% (2 days); Significant inhibition of AChE activity between 52.7% (5 days) and 81.28% (15 days);	[21]
1mg/L (15 days)		Significant inhibition of AChE activity between 19.7% (10 days), 54.48% (20 days), and 65.9% (30 days).	
0.3 mg/L (30 days)		Induction of the stress protein family Hsp70.	[23]

4. Metal Contamination

Metals can have several sources; they may be from anthropogenic activities such as industrial and miner activity, metal plating activities, rainwater runoff, and sewage [139–141], or by other sources as metals such as copper, cadmium, zinc, and lead may also be released by natural sources [43]. Moreover, metals may be divided into two categories—essential and non-essential elements. Essential elements, such as copper, iron, magnesium, and zinc, have a biological function to the organisms [142,143] but may become toxic at great concentrations [144]. On the other hand, non-essential elements (e.g., mercury, lead, cadmium, and arsenic) have no role on the metabolic processes [145,146], being toxic even at very low concentrations [144].

Although some metals have a key role on the normal organisms' metabolic maintenance, at excessive concentrations, all are toxic [147]. Despite metals being naturally present in the environment, human activities have contributed to the increase in its concentrations [148]. Furthermore, high levels of CO₂ and low pH may lead to the increase in metals solubility or to the changes in their speciation, therefore contributing to the increase in metals bioavailability [149–152]. Additionally, great levels of CO₂ may influence the accumulation and intracellular link between metals and marine organisms [153–155].

Moreover, the metal concentrations may fluctuate seasonally due to the runoff variations and also according to the tidal and deep cycles; spatially according to the salinity, organic matter, temperature, and availability of other contaminants [156]; as a function of the life-cycle stage and physicochemical form of the metal [157].

Metals are persistent pollutants and exhibit hard degradation [158]. Aquatic organisms are permanently exposed to its action, as metals are present in the water column and sediments. Thus, when the metals enter in the system, they may affect, directly, the organisms by direct pathways with cell proteins, resulting in non-specific bonds, enzymatic co-factors, or transcription factors replacement and discharging of stored metals. On the other hand, the metals are able to affect indirectly the organisms by oxidative damage on the proteins or other key cell constituents [155,159–163]. Metals are also present in many chemical formulations, such as pesticides, with the ability to accumulate on the organisms by different processes (e.g., bioaccumulation, bioconcentration, proliferation along of the food web) [164]. The level of metals on the organisms may be considerably greater than on the environment [165], with adverse effects at biochemical and physiological pathways for the organisms, due to the overproduction of oxygen radicals, which will affect cellular processes through the oxidative damage caused by these radicals, and leading, for example, to lipid peroxidation occurrence and damage to biological membranes [166], affecting its properties, namely the permeability and fluidity [167]. In the ultimate analysis, metals may have dangerous effects on human health, as human beings by consuming these organisms may accumulate the metals present on this food resource [168]. Some authors have reported damage to organs such as the brain, liver, and kidney, with the excessive exposure to copper [169]. Cardiovascular health problems, cancer, and negative impacts on renal tubular function to the reabsorption of amino acids, proteins, and sugars, with the exposure to cadmium [170] and embryo toxicity, allergic reactions, and dermatitis [171] were also related to the exposure to copper.

Metals toxicity is often related to an increase in the free radicals' levels that can interact with biomolecules such as DNA bases, lipases, and protein thiols, as well as to genotoxicity [172,173] and biological homeostasis disruption [174]. Therefore, metals are able to compromise growth, development, reproductive success, and survival of the organisms and consequently have an impact on the structure and function of the populations and communities to which they belong [144,166,175–178]. Thus, the indiscriminate use of metals comprises a marked environmental risk [179] with effects on the ecosystems and their biota.

4.1. Copper—From Essential to a Toxic Element

Copper is known as an essential element, with a key role on several biological functions such as mitochondrial and cell respiration, neurotransmitter biosynthesis, and free radical detoxification [180]. It is a cofactor of many enzymes (e.g., Cu/Zn superoxide dismutase (SOD), cytochrome c oxidase, amino oxidase, laccase, plastocyanin, polyphenol oxidase) [181,182] and the copper deficiency may result in a decrease in the enzymatic activity involved in the antioxidant defense system and, consequently, in an increase in reactive oxygen species (ROS) [183,184]. This metal also participates in hemoglobin level regulation and embryonic development [180]. Despite the key work of this micronutrient, at excessive concentrations, copper may become toxic to the organisms, as reported by several researchers.

Excessive metal concentrations are toxic to the organisms and the copper is not an exception. Several studies have shown that copper toxicity is dependent on its speciation, with the Cu⁺ being the most bioavailable, so the most toxic ions [185] and the content of free ions are dependent on the physicochemical properties of the water as well as of the organic ligands' bioavailability [186,187].

Copper is reported as a promotor of oxidative stress, as it catalyzes ROS production, when at excessive concentrations, by the generation of oxidant radicals [55,188,189], with consequences on membrane lipids and, consequently, with cell damage [56]. Similarly, this

metal has been described with inhibitory effects on the acetylcholinesterase activity [190] and on the phagocytic function, by stimulation or inhibition of the ROS production, according to the concentration [191]. Moreover, copper exposure has revealed impacts at the biochemical level, with alterations on the fatty acids profile, namely the increase in saturated fatty acids and the decrease in unsaturated fatty acids content, including the decrease in essential fatty acids such as EPA. These molecules are a key structural constituent of the cell membrane phospholipid bilayer (whose variations may affect the membrane properties such as the fluidity and density); of marine species from different trophic levels, such as the diatom *Thalassiosira weissfloggi*, the zooplanktonic species *Acartia tonsa* and *Artemia franciscana*, and the bivalves *Cerastoderma edule* and *Scrobicularia plana* [18,27,29]; of the enzymatic activity, with the inhibition of digestive gland hexokinase activity associated with a decrease in GSH content on *Mytilus galloprovincialis* [192]. De Almeida et al. [193] observed an inhibition of GPx activity on the bivalves *Perna perna*, also associated with a decrease in GSH levels and with consequent increase in lipid peroxidation occurrence. Doyotte et al. [194] also reported a decrease in GSH content on *Unio tumidus* when exposed to copper. Maria and Bebianno [195] tested the effects of copper exposure on *Mytilus galloprovincialis* and observed different responses according to the analyzed tissue, verifying the activation of antioxidant enzymes, namely glutathione reductase, glutathione S-transferase, glutathione peroxidase, catalase, and metallothionines on gills and the inhibition on the digestive gland; only SOD activity was inhibited on both tissues. Mesquita et al. [42] evaluated the effects of copper exposure on the antioxidant defense system of two bivalves species (*Cerastoderma edule* and *Scrobicularia plana*) considering two different size classes, namely large and small organisms. In this study, an increase in the GST and GR activity at the lowest concentration was observed, with a return to the basal activity on the remaining concentrations and a gradual increase in GPx activity across the range of concentrations, to the large organisms of both species. In the case of the small organisms, an opposite trend was observed with *C. edule* and *S. plana* exhibiting the inhibition of GPx activity, keeping the activities of the remaining enzymes relatively constant. Moreover, effects of copper exposure have also been reported at the physiological level, such as the reduction in the abundance, growth inhibition, and no reproductive success of the marine organisms [196,197]. Moreover, copper has showed effects on the macrofouling invertebrates' assemblages [198], as it is a main contaminant of antifouling paints formulations [199].

4.2. Copper Applications and Copper Sulfate Effects on Aquatic Organisms

Copper is a metal with ecological relevance and is involved in coastal pollution processes. This metal can be employed in several ways, for example, as pentahydrate copper (II) sulphate, copper (II) sulphate, dihydrate copper (II) chloride, copper (II) chloride, copper oxide, or copper (II) [27]. Copper-derived compounds are used in formulations of fungicides, bactericides, herbicides, and anti-vegetative paints [48,196,200–219]. Moreover, the use of copper nanoparticles has increased in several products, namely agrochemicals formulations, paints, antimicrobial products, catalyzers, semiconductors compounds, and sensors, with a consequent increase in the copper release to the environment, and in the terrestrial and in the aquatic systems [220–224]. As this metal has many applications, industrial discharges [196], urban runoff, sewers discharges [225], and antifouling biocides [226,227] are most often the contributors of copper pollution [199].

The WHO [228] defined a reference value of 2.0 mg L⁻¹ to Cu; however, the same identity described the occurrence of higher values until 30 mg L⁻¹ in drinking water. Moreover, environmental studies have reported Cu concentrations of about 10 mg L⁻¹ in aquatic systems near urban areas, achieving 100 mg L⁻¹ in aquatic bodies surrounding mining areas [229].

Copper sulphate is a chemical-based copper used in industrial and agricultural activities as an ingredient in pesticides formulations (bactericides, herbicides, fungicides, etc.). Several studies have reported the effects of this chemical at different levels, such as reproductive impairments, reduced growth, or behavioral changes [38–40,43,230]. More-

over, copper can lead to physiological changes and proteins dysfunction [47,230]. In fish, excessive exposure to copper may cause dangerous effects on gills, guts, and the sensorial system [40]. Effects reported in in vivo studies should alert people to the Cu toxicity, mainly because the tested concentrations are lower or similar to the reference value established for Cu and significantly lower than the concentrations found in the aquatic systems. Thus, a compilation of copper effects can be analyzed in Table 3.

Table 3. Effects of copper under the form of copper sulphate on several species, to evaluate different parameters, such as lethality, behavior, or biochemical indicators.

Concentration/ Duration	Species/ Community	Effects	References
0.01–1.00% (w/v)	<i>Rhodococcus erythropolis</i>	Effects on the bacterial cytoplasmic membrane; Decrease in the percentage of adapted cells with polarized membranes; Alteration to the fatty acids profile—increase in the saturated fatty acids and decrease in monounsaturated and polyunsaturated fatty acids. Effects on the biomass and metabolic activities of bacteria associated with the sediment; Changes in the community structure;	[45]
30.2–603.4 mg/kg of wet sediment (10 days)	Microbial community of marine sediment	Effects on activity and survival of marine metazoan fauna; Impacts on the bioturbation ability; Effects on fatty acids profile—Decrease in SFA (C14:0, C15:0, C16:0, C17:0, C18:0), MUFA (C16:1n5), and PUFA (C20:4n5, 8, 11, 14; C20:5n3) at 30 mg/kg; increase in SFA (C15:0, C17:0, C19:0) and MUFA (C17:1n7, C18:1n7) at 90 mg/kg Decrease in the growth rate, with the increase in the concentration and exposure time; Loss of the intact structure at 840 µg/L;	[41]
180–840 µg/L (96 h)	<i>Chaetoceros calcitrans</i>	Significant changes in the chlorophyll a content (increase at 180 µg/L—96 h, and decrease at 530 µg/L (75%) and at 840 µg/L (94%)) Changes in the enzymatic activity of catalase and superoxide dismutase, establishing a positive correlation with the concentrations 50, 180, and 450 µg/L.	[38]
	<i>Phaeodactylum tricomutum</i>	Lipid peroxidation: Significant increase in the MDA concentration at 10 µg/L (p < 0.05). Significant increase in the catalase activity (p < 0.05), with the difference being dependent on the concentrations. Significant reduction in glutathione peroxidase activity to all treatments (p < 0.05).	
5 and 10 µg/L (24 h)	<i>Rhodomonas salina</i>	Significant increase in CAT activity at 5 µg/L. Significant increase in GPx activity at the lower concentration Significant inhibition of ascorbate peroxidase activity to all treatments.	[48]
	<i>Cylindrotheca closterium</i>	Increase in the superoxide dismutase activity, but non-significant.	
200 ppb	<i>Gracilaria tenuistipitata</i>	Effects in photosynthesis process; Induction of oxidative stress; Changes in fatty acids profile—increase in SFA (C14:0, C16:0, C18:0) and MUFA (C18:1n7, C18:1n9) and decrease in PUFA (C18:2n6, C18:3n6, C18:5n4, C20:4n6, C20:5n3, 22:6n3). Changes in carbohydrates—increase in lactate levels and decrease in glycogen and pyruvate levels on every treatment;	[35]
Sub-lethal concentrations (72 h)	<i>Lamellidens marginalis</i>	Inhibition of the oxidative metabolism on the tissues—decrease in the succinate dehydrogenase and malate dehydrogenase activities; increase in the glucose-6-phosphate dehydrogenase activity.	[52]
	Phototrophic organisms and macroinvertebrates	Decrease in photosynthetic health—copper’s disruptive influence on the electrons-carrying system on photosystem II [231,232]; Restructuration effect on biofilms chemical composition—dangerous effects on biofilms function [233].	[50]
0.00–2.10 mg/L (96 h)	<i>Cerastoderma edule</i>	Large Size LC50 = 0.818 (0.595–0.987) mg/L Increase in SFA and PUFA until 31.78% and 16.60%, respectively; Decrease in MUFA and HUFA until 4.65% and 37.73%, respectively.	
		Small Size LC50 = 1.129 (0.968–1.289) mg/L Decrease in SFA, MUFA, and PUFA until 15.43%, 11.71%, and 4.69%, respectively; Increase in HUFA up to 31.82%.	[29]
0.00–4.00 mg/L (96 h)	<i>Scrobicularia plana</i>	Large Size LC50 = 2.563 (2.229–2.903) mg/L Increase in SFA up to 16.98% and maintenance of the levels of unsaturated fatty acids. Small Size LC50 = 4.705 (3.540–12.292) mg/L Decrease in SFA, MUFA, and PUFA until 27.14%, 13%, and 4.69%. Increase up to 24.94%.	

Table 3. Cont.

Concentration/ Duration	Species/ Community	Effects	References
0.00–2.10 mg/L (96 h)	<i>Cerastoderma edule</i>	Large Size Biphasic response of GR and GST activity Increase in GPx and TBARS levels, indicating the possible occurrence of lipid peroxidation Small Size Decrease in GR, GST, and GPx activity, and TBARS levels	[29]
0.00–4.00 mg/L (96 h)	<i>Scrobicularia plana</i>	Large Size Biphasic response of GR and GST activity Increase in GPx activity and TBARS levels, indicating the possible occurrence of lipid peroxidation Small Size Decrease in GR and GPx activity Maintenance of GST activity Biphasic response regarding TBARS levels and consequently lipid peroxidation occurrence Damage on DNA strongly dependent on copper concentration—Significant increase in % tail DNA compared with control, to all treatments and different damage levels among all treatments.	[29]
0–100 µg/L (5 days)	<i>Mytilus edulis</i>	Increase in the total glutathione levels on adductor muscle to all treatments, being significant to the organisms exposed at 32 and 56 µg/L. Histological abnormalities on adductor muscle—increase in the myocytes size and loss of the myocytes bundle structure. Histological changes in gills—hypoplasia (loss of cilia) to all treatments Loss of the digestive tubes definitions, likely by necrosis. LC50 = 1.717 (1.571–1.873) mg/L.	[54]
0.00–3.00 mg/L (96 h)	<i>Ctenopharyngodon idella</i>	Behavioral changes—anxiety, spasms, breathing difficulties, fast and erratic swimming, abrupt change in position and orientation. Fast opercular movement, frequent air intake, and remaining on one side before death, observed on the initial exposure stages, becoming occasional. LC50 = 2.310 (2.165–2.463) mg/L	[43]
Lethal effects: 0.00–3.50 mg/L (96 h)	<i>Rutilus frisii</i>	Damage on the growth parameters, such as significant differences on body weight to all treatments, specific growth rate to weight showing significant differences at 0.23 mg/L. Food conversion ratio and survival rate with significant differences at 0.23 mg/L.	[39]
Sub-lethal effects: 0.00, 0.11, 0.23 mg/L (60 days)	<i>Danio rerio</i> (embryonic stage)	Fewer functional neuromast and inability of the larvae to orient themselves Mortality, hatching inhibition, and impairment of the larvae development.	[40]
68–244 µg/L (from egg fertilization for 120 h)	<i>Danio rerio</i> (larvae from 2 to 5 days)	Neuromast cell damage, apoptosis, and loss of ciliated cell markers.	[47]
0.15–2.50 mg/L (5 days)	<i>Clarias gariepinus</i> (embryonic stage)	Decrease in pigmentation (from 15% to 70%). Increase in the lipid peroxidation products regarding the control ($p < 0.0001$). Significant increase in the catalase activity to all times except to 24 and 48 h (non-significant increase).	[234]
0.1641 ppm (from 24 h to 30 days)	<i>Penaeus indicus</i>	Increase in the superoxide dismutase activity (from 24 h to 10 days) and decrease at 20 and 30 days. Significant decrease in phagocytosis to all treatments. Significant decrease in superoxide dismutase activity to the organisms exposed to 60 and 110 µg/L.	[51]
0–110 µg/L -copper chloride (7 days)	<i>Ruditapes philippinarum</i>	Significant increase in the hemocytes percentage showing positivity to cytochrome oxidase to the organisms exposed to 60 µg/L.	[49]

5. Conclusions

The utilization of chemical compounds has increased in recent decades mainly related to anthropogenic activities, becoming crucial to monitoring their impacts on the ecosystems and in the communities. This review highlights the dangerous effects of organic and inorganic pollutants, with a focus on two pesticides widely used in the world: the organic herbicide—oxyfluorfen, and the inorganic pesticide—copper sulphate. Despite the different action modes of these two chemicals, both are reported as dangerous to non-target species, and its application has increased to provide well-being and development of human activities. After an intensive analysis about the known effects of these two chemicals, reported in many studies, this review highlights a higher sensitivity of the primary producers to copper and oxyfluorfen, when compared to the consumers. Oxyfluorfen is shown to be more dangerous than copper to the organisms from different trophic levels.

Despite the reference levels established to Cu (2.0 mg L⁻¹) and oxyfluorfen (from 0.05 mg Kg⁻¹ to 0.1 mg Kg⁻¹ in food wells), the highest concentrations have been reported in environmental systems (up to 100 mg L⁻¹ to Cu, and up to 26.3 mg L⁻¹ to oxyfluorfen). Moreover, according to the in vitro studies, both reference and environmental concentrations to each contaminant can comprise a harmful effect to aquatic organisms; namely,

the induction of ROS production, with changes in the antioxidant system defense; the occurrence of lipid peroxidation, with consequent changes to the fatty acids profile; the changes to the photosynthetic pathway, with a decrease in the photosynthetic pigments concentrations and in the photosynthetic carriers, are the most reported consequences of the exposure to both pesticides, with the inhibition of the acetylcholinesterase activity also having an effect often reported but associated with the oxyfluorfen exposure. Then, these impacts at the organisms' level should lead to negative impacts on the structure and function of the ecosystems.

This review also emphasizes the lack of information regarding the oxyfluorfen consequences on marine systems and on non-target species, with further research about this chemical at the biological level being of extreme importance, considering the known effects, as its usage is increasing daily, as well as its discharges on the aquatic systems with dangerous effects on these ecosystems and communities. It is imperative to monitor programs to aquatic systems and to assess the effects of contaminants considering lethal and sub-lethal effects, namely in terms of reproduction, growth rate, nutritive value, antioxidant defense, or neurotoxic effects, and based on these assessments to implement mitigation plans, improve the legislation, and work on the restoration of the ecosystems' health status.

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References

1. Fujiwara, T.; O'Hagan, D. Successful fluorine-containing herbicide agrochemicals. *J. Fluor. Chem.* **2014**, *167*, 16–29. [[CrossRef](#)]
2. Luo, Y.L.; Guo, W.S.; Ngo, H.H.; Nghiem, L.D.; Hai, F.I.; Zhang, J.; Liang, S.; Wang, X.C. A review on the occurrence of micropollutants in the aquatic environment and their fate and removal during wastewater treatment. *Sci. Total Environ.* **2014**, *473–474*, 619–641. [[CrossRef](#)] [[PubMed](#)]
3. Schwarzenbach, R.P.; Egli, T.; Hofstetter, T.B.; Von Gunten, U.V.; Wehrli, B. Global Water Pollution and Human Health. *Annu. Rev. Environ. Resour.* **2010**, *35*, 109–136. [[CrossRef](#)]
4. Loos, R.; Locoro, G.; Comero, S.; Contini, S.; Schwesig, D.; Werres, F.; Balsaa, P.; Gans, O.; Weiss, S.; Blaha, L.; et al. Pan-European survey on the occurrence of selected polar organic persistent pollutants in ground water. *Water Res.* **2010**, *44*, 4115–4126. [[CrossRef](#)]
5. FAO. *Pesticides Use*; FAO: Rome, Italy, 2020.
6. Eurostat. Agri-Environmental Indicator—Consumption of Pesticides—Statistics Explained. Available online: https://ec.europa.eu/eurostat/statistics-explained/index.php?title=Agri-environmental_indicator_-_consumption_of_pesticides (accessed on 11 March 2022).
7. Fernández, P.; Alcántara, R.; Osuna, M.D.; Vila-Aiub, M.M.; De Prado, R. Forward selection for multiple resistance across the non-selective glyphosate, glufosinate and oxyfluorfen herbicides in *Lolium* weed species. *Pest Manag. Sci.* **2017**, *73*, 936–944. [[CrossRef](#)] [[PubMed](#)]
8. Corbett, C.-A.L.; Tardif, F.J. Detection of resistance to acetolactate synthase inhibitors in weeds with emphasis on DNA-based techniques: A review. *Pest Manag. Sci.* **2006**, *62*, 584–597. [[CrossRef](#)]

9. Singh, B.K.; Shaner, D. Biosynthesis of Branched Chain Amino Acids: From Test Tube to Field. *Plant Cell* **1995**, *7*, 935–944. [[CrossRef](#)]
10. Duke, S.O. Overview of herbicide mechanisms of action. *Environ. Health Perspect.* **1990**, *87*, 263–271. [[CrossRef](#)]
11. Polyak, S.W.; Abell, A.D.; Wilce, M.C.J.; Zhang, L.; Booker, G.W. Structure, function and selective inhibition of bacterial acetyl-coa carboxylase. *Appl. Microbiol. Biotechnol.* **2012**, *93*, 983–992. [[CrossRef](#)]
12. Hao, G.-F.; Zuo, Y.; Yang, S.-G.; Yang, G.-F. Protoporphyrinogen Oxidase Inhibitor: An Ideal Target for Herbicide Discovery. *Chim. Int. J. Chem.* **2011**, *65*, 961–969. [[CrossRef](#)]
13. Park, J.; Ahn, Y.O.; Nam, J.-W.; Hong, M.-K.; Song, N.; Kim, T.; Yu, G.-H.; Sung, S.-K. Biochemical and physiological mode of action of tiafenacil, a new protoporphyrinogen IX oxidase-inhibiting herbicide. *Pestic. Biochem. Physiol.* **2018**, *152*, 38–44. [[CrossRef](#)] [[PubMed](#)]
14. Ahrens, H.; Lange, G.; Müller, T.; Rosinger, C.; Willms, L.; Van Almsick, A. 4-Hydroxyphenylpyruvate Dioxygenase Inhibitors in Combination with Safeners: Solutions for Modern and Sustainable Agriculture. *Angew. Chem. Int. Ed.* **2013**, *52*, 9388–9398. [[CrossRef](#)] [[PubMed](#)]
15. Wakabayashi, K.; Böger, P. Target sites for herbicides: Entering the 21st century. *Pest Manag. Sci.* **2002**, *58*, 1149–1154. [[CrossRef](#)] [[PubMed](#)]
16. Gupta, S.; Gajbhiye, V.T. Effect of concentration, moisture and soil type on the dissipation of flufenacet from soil. *Chemosphere* **2002**, *47*, 901–906. [[CrossRef](#)]
17. Lechelt-Kunze, C.; Meissner, R.C.; Drewes, M.; Tietjen, K. Flufenacet herbicide treatment phenocopies the fiddlehead mutant in *Arabidopsis thaliana*. *Pest Manag. Sci.* **2003**, *59*, 847–856. [[CrossRef](#)]
18. Mesquita, A.; Gonçalves, F.; Rocha, C.; Marques, J.; Gonçalves, A. Biochemical Effects of Two Pesticides in Three Different Temperature Scenarios on the Diatom *Thalassiosira weissflogii*. *Processes* **2021**, *9*, 1247. [[CrossRef](#)]
19. Sheeba; Singh, V.P.; Srivastava, P.K.; Prasad, S.M. Differential physiological and biochemical responses of two cyanobacteria *Nostoc muscorum* and *Phormidium foveolarum* against oxyfluorfen and UV-B radiation. *Ecotoxicol. Environ. Saf.* **2011**, *74*, 1981–1993. [[CrossRef](#)]
20. Geoffroy, L.; Teisseire, H.; Couderchet, M.; Vernet, G. Effect of oxyfluorfen and diuron alone and in mixture on antioxidative enzymes of *Scenedesmus obliquus*. *Pestic. Biochem. Physiol.* **2002**, *72*, 178–185. [[CrossRef](#)]
21. Hassanein, H.M.A. toxicological effects of the herbicide oxyfluorfen on acetylcholinesterase in two fish species: *Oreochromis niloticus* and *Gambusia affinis*. *J. Environ. Sci. Health Part A Toxic/Hazard. Subst. Environ. Eng.* **2002**, *37*, 521–527. [[CrossRef](#)]
22. Peixoto, F.; Alves-Fernandes, D.; Santos, D.; Fontainhas-Fernandes, A. Toxicological effects of oxyfluorfen on oxidative stress enzymes in tilapia *Oreochromis niloticus*. *Pestic. Biochem. Physiol.* **2006**, *85*, 91–96. [[CrossRef](#)]
23. Hassanein, H.M.A.; Banhaway, M.A.; Soliman, F.M.; Abdel-Rehim, S.A.; Müller, W.E.G.; Schröder, H.C. Induction of Hsp70 by the Herbicide Oxyfluorfen (Goal) in the Egyptian Nile Fish, *Oreochromis niloticus*. *Arch. Environ. Contam. Toxicol.* **1999**, *37*, 78–84. [[CrossRef](#)]
24. Maazouzi, C.; Masson, G.; Izquierdo, M.S.; Pihan, J.-C. Chronic copper exposure and fatty acid composition of the amphipod *Dikerogammarus villosus*: Results from a field study. *Environ. Pollut.* **2008**, *156*, 221–226. [[CrossRef](#)]
25. Ritter, A.; Dittami, S.M.; Goulitquer, S.; Correa, J.A.; Boyen, C.; Potin, P.; Tonon, T. Transcriptomic and metabolomic analysis of copper stress acclimation in *Ectocarpus siliculosus* highlights signaling and tolerance mechanisms in brown algae. *BMC Plant Biol.* **2014**, *14*, 116. [[CrossRef](#)]
26. Ritter, A.; Goulitquer, S.; Salaün, J.-P.; Tonon, T.; Correa, J.A.; Potin, P. Copper stress induces biosynthesis of octadecanoid and eicosanoid oxygenated derivatives in the brown algal kelp *Laminaria digitata*. *New Phytol.* **2008**, *180*, 809–821. [[CrossRef](#)] [[PubMed](#)]
27. Filimonova, V.; Gonçalves, F.; Marques, J.C.; De Troch, M.; Gonçalves, A.M. Biochemical and toxicological effects of organic (herbicide Primextra® Gold TZ) and inorganic (copper) compounds on zooplankton and phytoplankton species. *Aquat. Toxicol.* **2016**, *177*, 33–43. [[CrossRef](#)]
28. Filimonova, V.; Nys, C.; De Schampelaere, K.A.C.; Gonçalves, F.; Marques, J.C.; Gonçalves, A.M.M.; De Troch, M. Ecotoxicological and biochemical mixture effects of an herbicide and a metal at the marine primary producer diatom *Thalassiosira weissflogii* and the primary consumer copepod *Acartia tonsa*. *Environ. Sci. Pollut. Res.* **2018**, *25*, 22180–22195. [[CrossRef](#)]
29. Mesquita, A.; Gonçalves, F.; Verdelhos, T.; Marques, J.; Gonçalves, A. Fatty acids profiles modifications in the bivalves *Cerastoderma edule* and *Scrobicularia plana* in response to copper sulphate. *Ecol. Indic.* **2018**, *85*, 318–328. [[CrossRef](#)]
30. Sibi, G.; Anuraag, T.; Bafila, G. Copper stress on cellular contents and fatty acid profiles in chlorella species. *Online J. Biol. Sci.* **2014**, *14*, 209–217. [[CrossRef](#)]
31. Gomes, D.G.; Lopes-Oliveira, P.J.; Debiassi, T.V.; da Cunha, L.S.; Oliveira, H.C. Regression models to stratify the copper toxicity responses and tolerance mechanisms of *Glycine max* (L.) Merr. plants. *Planta* **2021**, *253*, 1–14. [[CrossRef](#)]
32. Tagliaferro, M.; Gonçalves, A.M.; Bergman, M.; Sobral, O.; Graça, M.A. Assessment of metal exposure (uranium and copper) by the response of a set of integrated biomarkers in a stream shredder. *Ecol. Indic.* **2018**, *95*, 991–1000. [[CrossRef](#)]
33. EEA. *EEA Water 2012 Report: European Waters—Assessment of Status and Pressures*; EEA: Boston, MA, USA, 2012.
34. Littlefield-Wyer, J.; Brooks, P.; Katouli, M. Application of biochemical fingerprinting and fatty acid methyl ester profiling to assess the effect of the pesticide Atradox on aquatic microbial communities. *Environ. Pollut.* **2008**, *153*, 393–400. [[CrossRef](#)]

35. Pinto, E.; Carvalho, A.P.; Cardozo, K.H.M.; Malcata, F.; dos Anjos, F.M.; Colepicolo, P. Effects of heavy metals and light levels on the biosynthesis of carotenoids and fatty acids in the macroalgae *Gracilaria tenuistipitata* (var. *liui* Zhang & Xia). *Rev. Bras. Farm.* **2011**, *21*, 349–354. [[CrossRef](#)]
36. Fokina, N.N.; Ruokolainen, T.R.; Nemova, N.N.; Bakhmet, I.N. Changes of Blue Mussels *Mytilus edulis* L. Lipid Composition Under Cadmium and Copper Toxic Effect. *Biol. Trace Element Res.* **2013**, *154*, 217–225. [[CrossRef](#)] [[PubMed](#)]
37. Chelomin, V.; Belcheva, N. Alterations of microsomal lipid synthesis in gill cells of bivalve mollusc *Mizuhopecten yessoensis* in response to cadmium accumulation. *Comp. Biochem. Physiol. Part C Comp. Pharmacol.* **1991**, *99*, 1–5. [[CrossRef](#)]
38. Anu, P.; Nandan, S.B.; Jayachandran, P.; Xavier, N.D. Toxicity effects of copper on the marine diatom, *Chaetoceros calcitrans*. *Reg. Stud. Mar. Sci.* **2016**, *8*, 498–504. [[CrossRef](#)]
39. Gharedaashi, E.; Nekoubin, H.; Imanpoor, M.R.; Taghizadeh, V. Effect of copper sulfate on the survival and growth performance of Caspian Sea kutum, *Rutilus frisii kutum*. *Springerplus* **2013**, *2*, 498. [[CrossRef](#)] [[PubMed](#)]
40. Johnson, A.; Carew, E.; Sloman, K. The effects of copper on the morphological and functional development of zebrafish embryos. *Aquat. Toxicol.* **2007**, *84*, 431–438. [[CrossRef](#)]
41. Mayor, D.J.; Gray, N.B.; Elver-Evans, J.; Midwood, A.J.; Thornton, B. Metal-Macrofauna Interactions Determine Microbial Community Structure and Function in Copper Contaminated Sediments. *PLoS ONE* **2013**, *8*, e64940. [[CrossRef](#)]
42. Mesquita, A.F.; Marques, S.M.; Marques, J.C.; Gonçalves, F.J.M.; Gonçalves, A.M.M. Copper sulphate impact on the antioxidant defence system of the marine bivalves *Cerastoderma edule* and *Scrobicularia plana*. *Sci. Rep.* **2019**, *9*, 1–11. [[CrossRef](#)]
43. Nekoubin, H.; Gharedaashi, E.; Hatefi, S.; Sudagar, M.; Shahriari, R.; Asgharimoghadam, A. Determination of LC 50 of Copper Sulfate and Lead (II) Nitrate and Behavioral Responses of Grass Carp (*Ctenopharyngodon idella*). *Walailak J. Sci. Technol. WJST* **2012**, *9*, 333–340.
44. Mesquita, A.F.; Abrantes, N.; Campos, I.; Nunes, C.; Coimbra, M.A.; Gonçalves, F.J.; Marques, J.C.; Gonçalves, A.M. Effects of wildfire ash on the growth and biochemical profiles of the aquatic macrophyte *Lemna minor*. *Aquat. Toxicol.* **2022**, *250*, 106245. [[CrossRef](#)] [[PubMed](#)]
45. de Carvalho, C.C. Adaptation of *Rhodococcus erythropolis* cells for growth and bioremediation under extreme conditions. *Res. Microbiol.* **2012**, *163*, 125–136. [[CrossRef](#)] [[PubMed](#)]
46. Gutiérrez, I.B.; Mesquita, A.F.; Nunes, C.; Coimbra, M.A.; Gonçalves, F.J.; Marques, J.C.; Gonçalves, A.M. Impacts of S-metolachlor and terbuthylazine in fatty acid and carbohydrate composition of the benthic clam *Scrobicularia plana*. *Ecotoxicol. Environ. Saf.* **2019**, *173*, 293–304. [[CrossRef](#)]
47. Hernández, P.P.; Moreno, V.; Olivari, F.A.; Allende, M. Sub-lethal concentrations of waterborne copper are toxic to lateral line neuromasts in zebrafish (*Danio rerio*). *Hear. Res.* **2006**, *213*, 1–10. [[CrossRef](#)] [[PubMed](#)]
48. Lozano, P.; Trombini, C.; Crespo, E.; Blasco, J.; Moreno-Garrido, I. ROI-scavenging enzyme activities as toxicity biomarkers in three species of marine microalgae exposed to model contaminants (copper, Irgarol and atrazine). *Ecotoxicol. Environ. Saf.* **2014**, *104*, 294–301. [[CrossRef](#)]
49. Matozzo, L.B.V.; Ballarin, L.; Pampanin, D.M.; Marin, M.G. Effects of Copper and Cadmium Exposure on Functional Responses of Hemocytes in the Clam, *Tapes philippinarum*. *Arch. Environ. Contam. Toxicol.* **2001**, *41*, 163–170. [[CrossRef](#)]
50. McElroy, D.J.; Hochuli, D.F.; Doblin, M.A.; Murphy, R.J.; Blackburn, R.J.; Coleman, R.A. Effect of copper on multiple successional stages of a marine fouling assemblage. *Biofouling* **2017**, *33*, 904–916. [[CrossRef](#)]
51. Paila, R.V.; Yallapragada, P.R. Antioxidant Responses and Lipid Peroxidation of *Penaeus Indicus* Postlarvae Subjected to Sublethal Copper Exposure. *Crustaceana* **2011**, *84*, 1197–1210. [[CrossRef](#)]
52. Satyaparameshwar, K.; Reddy, T.R.; Kumar, N.V. Study of carbohydrate metabolism in selected tissues of freshwater mussel, *Lamellidens marginalis* under copper sulphate toxicity. *J. Environ. Biol.* **2006**, *27*, 39–41.
53. Jesus, F.; Mesquita, F.; Aldama, E.V.; Marques, A.; Gonçalves, A.M.M.; Magalhães, L.; Nogueira, A.J.A.; Ré, A.; Campos, I.; Pereira, J.L.; et al. Do Freshwater and Marine Bivalves Differ in Their Response to Wildfire Ash? Effects on the Antioxidant Defense System and Metal Body Burden. *Int. J. Environ. Res. Public Health* **2023**, *20*, 1326. [[CrossRef](#)]
54. Al-Subiai, S.N.; Moody, A.J.; Mustafa, S.A.; Jha, A.N. A multiple biomarker approach to investigate the effects of copper on the marine bivalve mollusc, *Mytilus edulis*. *Ecotoxicol. Environ. Saf.* **2011**, *74*, 1913–1920. [[CrossRef](#)]
55. Livingstone, D. Contaminant-stimulated Reactive Oxygen Species Production and Oxidative Damage in Aquatic Organisms. *Mar. Pollut. Bull.* **2001**, *42*, 656–666. [[CrossRef](#)]
56. Van der Oost, R.; Beyer, J.; Vermeulen, N.P.E. Fish bioaccumulation and biomarkers in environmental risk assessment: A review. *Environ. Toxicol. Pharmacol.* **2003**, *13*, 57–149. [[CrossRef](#)] [[PubMed](#)]
57. Scalbert, A.; Johnson, I.T.; Saltmarsh, M. Polyphenols: Antioxidants and beyond. *Am. J. Clin. Nutr.* **2005**, *81*, 215S–217S. [[CrossRef](#)]
58. Sies, H. *Oxidative Stress*; Nova Science Publishers: Orlando, FL, USA; Academic Press: London, UK, 1985.
59. Wang, G.; Liu, B.; Tang, B.; Zhang, T.; Xiang, J. Pharmacological and immunocytochemical investigation of the role of catecholamines on larval metamorphosis by β -adrenergic-like receptor in the bivalve *Meretrix meretrix*. *Aquaculture* **2006**, *258*, 611–618. [[CrossRef](#)]
60. Amiard, J.; Amiard-Triquet, C. *Les Biomarqueurs Dans L'évaluation de L'état Écologique Des Milieux Aquatiques*; Editions Technique & Doc; Lavoisier: Paris, France, 2008.
61. Verlecar, X.; Jena, K.; Chainy, G. Modulation of antioxidant defences in digestive gland of *Perna viridis* (L.), on mercury exposures. *Chemosphere* **2008**, *71*, 1977–1985. [[CrossRef](#)]

62. Amiard, J.-C.; Caquet, T.; Lagadic, L. *Use of Biomarkers for Environmental Quality Assessment*; CRC Press: London, UK, 2021. [[CrossRef](#)]
63. Gagnon, M.M.; Holdway, D.A. Metabolic Enzyme Activities in Fish Gills as Biomarkers of Exposure to Petroleum Hydrocarbons. *Ecotoxicol. Environ. Saf.* **1999**, *44*, 92–99. [[CrossRef](#)]
64. Diamantino, T.C.; Almeida, E.; Soares, A.M.; Guilhermino, L. Lactate dehydrogenase activity as an effect criterion in toxicity tests with *Daphnia magna* straus. *Chemosphere* **2001**, *45*, 553–560. [[CrossRef](#)] [[PubMed](#)]
65. Watanabe, K.; Ohori, Y.; Sato, Y.; Böger, P.; Wakabayashi, K. Changes in Fatty Acid Composition of Neutral Lipid in Mung Bean Cotyledons by Oxyfluorfen-Induced Peroxidation. *Pestic. Biochem. Physiol.* **2001**, *69*, 166–173. [[CrossRef](#)]
66. Bagchi, D.; Bagchi, M.; Hassoun, E.A.; Stohs, S.J. Cadmium-induced excretion of urinary lipid metabolites, DNA damage, glutathione depletion, and hepatic lipid peroxidation in sprague-dawley rats. *Biol. Trace Element Res.* **1996**, *52*, 143–154. [[CrossRef](#)] [[PubMed](#)]
67. Lei, Y.; Zhang, W.; Xu, W.; Zhang, Y.; Zhou, H.; Mai, K. Effects of waterborne Cu and Cd on anti-oxidative response, lipid peroxidation and heavy metals accumulation in abalone *Haliotis discus hannai* ino. *J. Ocean Univ. China* **2015**, *14*, 511–521. [[CrossRef](#)]
68. Ramírez, B.; Montero, D.; Izquierdo, M.; Haroun, R. Aquafeed imprint on bogue (Boops boops) populations and the value of fatty acids as indicators of aquaculture-ecosystem interaction: Are we using them properly? *Aquaculture* **2013**, *414–415*, 294–302. [[CrossRef](#)]
69. Engle, T.; Fellner, V.; Spears, J. Copper Status, Serum Cholesterol, and Milk Fatty Acid Profile in Holstein Cows Fed Varying Concentrations of Copper. *J. Dairy Sci.* **2001**, *84*, 2308–2313. [[CrossRef](#)] [[PubMed](#)]
70. Neves, M.; Castro, B.; Vidal, T.; Vieira, R.; Marques, J.; Coutinho, J.; Gonçalves, F.; Gonçalves, A. Biochemical and populational responses of an aquatic bioindicator species, *Daphnia longispina*, to a commercial formulation of a herbicide (Primextra® Gold TZ) and its active ingredient (S-metolachlor). *Ecol. Indic.* **2015**, *53*, 220–230. [[CrossRef](#)]
71. De Troch, M.; Boeckx, P.; Cnudde, C.; Van Gansbeke, D.; Vanreusel, A.; Vincx, M.; Caramujo, M. Bioconversion of fatty acids at the basis of marine food webs: Insights from a compound-specific stable isotope analysis. *Mar. Ecol. Prog. Ser.* **2012**, *465*, 53–67. [[CrossRef](#)]
72. Gonçalves, A.; Mesquita, A.; Verdelhos, T.; Coutinho, J.; Marques, J.; Gonçalves, F. Fatty acids' profiles as indicators of stress induced by of a common herbicide on two marine bivalves species: *Cerastoderma edule* (Linnaeus, 1758) and *Scrobicularia plana* (da Costa, 1778). *Ecol. Indic.* **2016**, *63*, 209–218. [[CrossRef](#)]
73. Kelly, J.R.; Scheibling, R.E. Fatty acids as dietary tracers in benthic food webs. *Mar. Ecol. Prog. Ser.* **2012**, *446*, 1–22. [[CrossRef](#)]
74. Grahl-Nielsen, O.; Andersen, M.; DeRoche, A.; Lydersen, C.; Wiig, Ø.; Kovacs, K. Fatty acid composition of the adipose tissue of polar bears and of their prey: Ringed seals, bearded seals and harp seals. *Mar. Ecol. Prog. Ser.* **2003**, *265*, 275–282. [[CrossRef](#)]
75. Iverson, S.; Field, C.; Bowen, W.D.; Blanchard, W. Quantitative fatty acid signature analysis: A new method of estimating predator diets. *Ecol. Monogr.* **2004**, *74*, 211–235. [[CrossRef](#)]
76. Budge, S.M.; Iverson, S.J.; Koopman, H.N. Studying trophic ecology in marine ecosystems using fatty acids: A primer on analysis and interpretation. *Mar. Mammal Sci.* **2006**, *22*, 759–801. [[CrossRef](#)]
77. Begul, P.K.; More, B.C.; Patole, S.S. Studies on Sub Lethal Effect of Cypermethrin and Oxyfluorfen on Biochemical Parameters of Earthworm Species, *Eisenia Foetida* Savigny, 1826. *Int. J. Innov. Res. Sci.* **2017**, *6*, 20437–20440. [[CrossRef](#)]
78. Neethu, K.V.; Saranya, K.S.; Krishna, N.G.A.; Praved, P.H.; Aneesh, B.P.; Nandan, S.B.; Marigoudar, S.R. Toxicity of copper on marine diatoms, *Chaetoceros calcitrans* and *Nitzschia closterium* from Cochin estuary, India. *Ecotoxicology* **2021**, *30*, 783–793. [[CrossRef](#)]
79. Li, Q.; Chen, H.-H.; Qi, Y.-P.; Ye, X.; Yang, L.-T.; Huang, Z.-R.; Chen, L.-S. Excess copper effects on growth, uptake of water and nutrients, carbohydrates, and PSII photochemistry revealed by OJIP transients in Citrus seedlings. *Environ. Sci. Pollut. Res.* **2019**, *26*, 30188–30205. [[CrossRef](#)] [[PubMed](#)]
80. Lochmiller, R.L.; Deerenberg, C. Trade-offs in evolutionary immunology: Just what is the cost of immunity? *Oikos* **2000**, *88*, 87–98. [[CrossRef](#)]
81. Hildebrand, M.; Manandhar-Shrestha, K.; Abbriano, R. Effects of chrysolaminarin synthase knockdown in the diatom *Thalassiosira pseudonana*: Implications of reduced carbohydrate storage relative to green algae. *Algal Res.* **2017**, *23*, 66–77. [[CrossRef](#)]
82. Davey, M.E.; O'Toole, G.A. Microbial Biofilms: From Ecology to Molecular Genetics. *Microbiol. Mol. Biol. Rev.* **2000**, *64*, 847–867. [[CrossRef](#)]
83. Hung, C.-C.; Santschi, P.H.; Gillow, J.B. Isolation and characterization of extracellular polysaccharides produced by *Pseudomonas fluorescens* Biovar II. *Carbohydr. Polym.* **2005**, *61*, 141–147. [[CrossRef](#)]
84. Sutherland, I.W. Microbial Polysaccharides from Gram-Negative Bacteria. *Int. Dairy J.* **2001**, *11*, 663–674. [[CrossRef](#)]
85. Kazy, S.K.; Sar, P.; Singh, S.P.; Sen, A.K.; D'Souza, S.F.D. Extracellular Polysaccharides of Copper-Sensitive and Copper-Resistant *Pseudomonas Aeruginosa* Strain: Synthesis, Chemical Nature and Copper Binding. *World J. Microbiol. Biotechnol.* **2002**, *18*, 583–588. [[CrossRef](#)]
86. Chen, B.; Yuan, M.; Liu, H. Removal of polycyclic aromatic hydrocarbons from aqueous solution using plant residue materials as a biosorbent. *J. Hazard. Mater.* **2011**, *188*, 436–442. [[CrossRef](#)] [[PubMed](#)]
87. Nagda, G.K.; Diwan, A.M.; Ghole, V.S. Potential of Tendu Leaf Refuse for Phenol Removal in Aqueous Systems. *Appl. Ecol. Environ. Res.* **2007**, *5*, 1–9. [[CrossRef](#)]

88. Nanseu-Njiki, C.P.; Dedzo, G.K.; Ngameni, E. Study of the removal of paraquat from aqueous solution by biosorption onto Ayous (*Triplochiton schleroxylon*) sawdust. *J. Hazard. Mater.* **2010**, *179*, 63–71. [[CrossRef](#)] [[PubMed](#)]
89. Abdeen, Z.; Mohammad, S.G. Study of the Adsorption Efficiency of an Eco-Friendly Carbohydrate Polymer for Contaminated Aqueous Solution by Organophosphorus Pesticide. *Open J. Org. Polym. Mater.* **2014**, *4*, 16–28. [[CrossRef](#)]
90. Valili, S.; Siavalas, G.; Karapanagioti, H.K.; Manariotis, I.D.; Christanis, K. Phenanthrene removal from aqueous solutions using well-characterized, raw, chemically treated, and charred malt spent rootlets, a food industry by-product. *J. Environ. Manag.* **2013**, *128*, 252–258. [[CrossRef](#)] [[PubMed](#)]
91. Waugh, C.A.; Nichols, P.D.; Schlabach, M.; Noad, M.; Nash, S.B. Vertical distribution of lipids, fatty acids and organochlorine contaminants in the blubber of southern hemisphere humpback whales (*Megaptera novaeangliae*). *Mar. Environ. Res.* **2014**, *94*, 24–31. [[CrossRef](#)] [[PubMed](#)]
92. Ahmed, S. Egyptian Apricot Stone (*Prunus armeniaca*) as a Low Cost and Eco-friendly Biosorbent for Oxamyl Removal from Aqueous Solutions. *Am. J. Exp. Agric.* **2014**, *4*, 302–321. [[CrossRef](#)]
93. Daam, M.A.; Brink, P.J.V.D. Implications of differences between temperate and tropical freshwater ecosystems for the ecological risk assessment of pesticides. *Ecotoxicology* **2010**, *19*, 24–37. [[CrossRef](#)]
94. Carvalho, F.P. Agriculture, pesticides, food security and food safety. *Environ. Sci. Policy* **2006**, *9*, 685–692. [[CrossRef](#)]
95. Singh, A.K.; Singh, P.P.; Tripathi, V.; Verma, H.; Singh, S.K.; Srivastava, A.K.; Kumar, A. Distribution of cyanobacteria and their interactions with pesticides in paddy field: A comprehensive review. *J. Environ. Manag.* **2018**, *224*, 361–375. [[CrossRef](#)]
96. Schreinemachers, P.; Tipraqsa, P. Agricultural pesticides and land use intensification in high, middle and low income countries. *Food Policy* **2012**, *37*, 616–626. [[CrossRef](#)]
97. Wilson, P.C.; Foos, J.F. Survey of carbamate and organophosphorous pesticide export from a south Florida (USA) agricultural watershed: Implications of sampling frequency on ecological risk estimation. *Environ. Toxicol. Chem.* **2006**, *25*, 2847–2852. [[CrossRef](#)] [[PubMed](#)]
98. World Health Organization. *Guidelines for Drinking-Water Quality*; World Health Organization: Geneva, Switzerland, 2011.
99. Sabarwal, A.; Kumar, K.; Singh, R.P. Hazardous effects of chemical pesticides on human health—Cancer and other associated disorders. *Environ. Toxicol. Pharmacol.* **2018**, *63*, 103–114. [[CrossRef](#)]
100. Li, Z.; Jennings, A. Worldwide Regulations of Standard Values of Pesticides for Human Health Risk Control: A Review. *Int. J. Environ. Res. Public Health* **2017**, *14*, 826. [[CrossRef](#)]
101. Kojima, H.; Sata, F.; Takeuchi, S.; Sueyoshi, T.; Nagai, T. Comparative study of human and mouse pregnane X receptor agonistic activity in 200 pesticides using in vitro reporter gene assays. *Toxicology* **2011**, *280*, 77–87. [[CrossRef](#)] [[PubMed](#)]
102. Luo, D.; Zhou, T.; Tao, Y.; Feng, Y.; Shen, X.; Mei, S. Exposure to organochlorine pesticides and non-Hodgkin lymphoma: A meta-analysis of observational studies. *Sci. Rep.* **2016**, *6*, 25768. [[CrossRef](#)] [[PubMed](#)]
103. Wiklund, K.; Dich, J.; Holm, L.E. Risk of malignant lymphoma in Swedish pesticide applicators. *Br. J. Cancer* **1987**, *56*, 505–508. [[CrossRef](#)]
104. Brouwer, M.; Huss, A.; van der Mark, M.; Nijssen, P.C.G.; Mulleners, W.M.; Sas, A.M.G.; van Laar, T.; de Snoo, G.R.; Kromhout, H.; Vermeulen, R.C.H. Environmental exposure to pesticides and the risk of Parkinson’s disease in the Netherlands. *Environ. Int.* **2017**, *107*, 100–110. [[CrossRef](#)]
105. Wang, A.; Cockburn, M.; Ly, T.T.; Bronstein, J.M.; Ritz, B. The association between ambient exposure to organophosphates and Parkinson’s disease risk. *Occup. Environ. Med.* **2014**, *71*, 275–281. [[CrossRef](#)]
106. Freire, C.; Koifman, R.J.; Sarcinelli, P.N.; Rosa, A.C.S.; Clapauch, R.; Koifman, S. Long-term exposure to organochlorine pesticides and thyroid status in adults in a heavily contaminated area in Brazil. *Environ. Res.* **2013**, *127*, 7–15. [[CrossRef](#)]
107. Mazur, C.S.; Marchitti, S.A.; Zastre, J. P-glycoprotein inhibition by the agricultural pesticide propiconazole and its hydroxylated metabolites: Implications for pesticide–drug interactions. *Toxicol. Lett.* **2015**, *232*, 37–45. [[CrossRef](#)]
108. Kirkhorn, S.R.; Schenker, M.B. Current Health Effects of Agricultural Work: Respiratory Disease, Cancer, Reproductive Effects, Musculoskeletal Injuries, and Pesticide-Related Illnesses. *J. Agric. Saf. Health* **2002**, *8*, 199–214. [[CrossRef](#)]
109. Thongprakaisang, S.; Thiantanawat, A.; Rangkadilok, N.; Suriyo, T.; Satayavivad, J. Glyphosate induces human breast cancer cells growth via estrogen receptors. *Food Chem. Toxicol.* **2013**, *59*, 129–136. [[CrossRef](#)] [[PubMed](#)]
110. Musicco, M.; Sant, M.; Molinari, S.; Filippini, G.; Gatta, G.; Berrino, F. A case-control study of brain gliomas and occupational exposure to chemical carcinogens: The risk to farmers. *Am. J. Epidemiol.* **1988**, *128*, 778–785. [[CrossRef](#)]
111. Norman, C. EPA halts dieldrin production. *Nature* **1974**, *250*, 528. [[CrossRef](#)]
112. Ansbaugh, N.; Shannon, J.; Mori, M.; Farris, P.E.; Garzotto, M. Agent Orange as a risk factor for high-grade prostate cancer. *Cancer* **2013**, *119*, 2399–2404. [[CrossRef](#)] [[PubMed](#)]
113. Hardersen, S.; Wratten, S.D. The Effects of Carbaryl Exposure of the Penultimate Larval Instars of *Xathocnemis Zealandica* on Emergence and Fluctuating Asymmetry. *Ecotoxicology* **1998**, *7*, 297–304. [[CrossRef](#)]
114. Galhano, V.; Santos, H.; Oliveira, M.M.; Gomes-Laranjo, J.; Peixoto, F. Changes in fatty acid profile and antioxidant systems in a *Nostoc muscorum* strain exposed to the herbicide bentazon. *Process. Biochem.* **2011**, *46*, 2152–2162. [[CrossRef](#)]
115. Dayan, F.E.; Duke, S.O. Natural Compounds as Next-Generation Herbicides. *Plant Physiol.* **2014**, *166*, 1090–1105. [[CrossRef](#)]
116. Duke, S.O. Why have no new herbicide modes of action appeared in recent years? *Pest Manag. Sci.* **2012**, *68*, 505–512. [[CrossRef](#)]
117. Giornal, F.; Pazenok, S.; Rodefeld, L.; Lui, N.; Vors, J.-P.; Leroux, F.R. Synthesis of diversely fluorinated pyrazoles as novel active agrochemical ingredients. *J. Fluor. Chem.* **2013**, *152*, 2–11. [[CrossRef](#)]

118. Jeschke, P. The unique role of halogen substituents in the design of modern agrochemicals. *Pest Manag. Sci.* **2010**, *66*, 10–27. [[CrossRef](#)]
119. Theodoridis, G. Chapter 4 Fluorine-Containing Agrochemicals: An Overview of Recent Developments. *Adv. Fluor. Sci.* **2006**, *2*, 121–175. [[CrossRef](#)]
120. MacDougall, P. Agri-Service Report. 2010. Available online: <https://www.spglobal.com/en/> (accessed on 25 February 2019).
121. O'Hagan, D. Understanding organofluorine chemistry. An introduction to the C–F bond. *Chem. Soc. Rev.* **2008**, *37*, 308–319. [[CrossRef](#)]
122. Dunitz, J.D. Organic Fluorine: Odd Man Out. *Chembiochem* **2004**, *5*, 614–621. [[CrossRef](#)]
123. Yu, Q.; Nelson, J.K.; Zheng, M.Q.; Jackson, M.; Powles, S. Molecular characterisation of resistance to ALS-inhibiting herbicides in *Hordeum leporinum* biotypes. *Pest Manag. Sci.* **2007**, *63*, 918–927. [[CrossRef](#)] [[PubMed](#)]
124. Grossmann, K. Auxin herbicides: Current status of mechanism and mode of action. *Pest Manag. Sci.* **2009**, *66*, 113–120. [[CrossRef](#)] [[PubMed](#)]
125. Arias, R.S.; Dayan, F.E.; Michel, A.; Howell, J.; Scheffler, B.E. Characterization of a higher plant herbicide-resistant phytoene desaturase and its use as a selectable marker. *Plant Biotechnol. J.* **2006**, *4*, 263–273. [[CrossRef](#)]
126. Arias, R.S.; Netherland, M.D.; Scheffler, B.; Puri, A.; Dayan, F. Molecular evolution of herbicide resistance to phytoene desaturase inhibitors in *Hydrilla verticillata* and its potential use to generate herbicide-resistant crops. *Pest Manag. Sci.* **2005**, *61*, 258–268. [[CrossRef](#)]
127. Aizawa, H.; Brown, H.M. Metabolism and Degradation of Porphyrin Iosynthesis Herbicides. In *Peroxidizing Herbicides*; Böger, P., Wakabayashi, K., Eds.; Springer: Berlin/Heidelberg, Germany, 1999; pp. 438–481.
128. Böger, P.; Matthes, B.; Schmalfuß, J. Towards the Primary Target of Chloroacetamides -New Findings Pave the Way. *Pest Manag. Sci.* **2000**, *56*, 497–508. [[CrossRef](#)]
129. Zhao, H.; Xu, J.; Dong, F.; Liu, X.; Wu, Y.; Wu, X.; Zheng, Y. Characterization of a novel oxyfluorfen-degrading bacterial strain *Chryseobacterium aquifrigidense* and its biochemical degradation pathway. *Appl. Microbiol. Biotechnol.* **2016**, *100*, 6837–6845. [[CrossRef](#)]
130. ECHA. *Annexes II, III, IV, VII—Defined & Temporary Maximum Residue Levels*; ECHA: Helsinki, Finland, 2022.
131. Ibrahim, A.M.; Sayed, D.A. Toxicological impact of oxyfluorfen 24% herbicide on the reproductive system, antioxidant enzymes, and endocrine disruption of *Biomphalaria alexandrina* (Ehrenberg, 1831) snails. *Environ. Sci. Pollut. Res.* **2019**, *26*, 7960–7968. [[CrossRef](#)]
132. Matringe, M.; Scalla, R. Studies on the Mode of Action of Acifluorfen-Methyl in Nonchlorophyllous Soybean Cells: Accumulation of Tetrapyrroles. *Plant Physiol.* **1988**, *86*, 619–622. [[CrossRef](#)]
133. Matringe, M.; Camadro, J.-M.; Labbe, P.; Scalla, R. Protoporphyrinogen oxidase as a molecular target for diphenyl ether herbicides. *Biochem. J.* **1989**, *260*, 231–235. [[CrossRef](#)] [[PubMed](#)]
134. Witkowski, D.A.; Halling, B.P. Inhibition of Plant Protoporphyrinogen Oxidase by the Herbicide Acifluorfen-Methyl. *Plant Physiol.* **1989**, *90*, 1239–1242. [[CrossRef](#)]
135. Hallahan, B.J.; Camilleri, P.; Smith, A.; Bowyer, J.R. Mode of Action Studies on a Chiral Diphenyl Ether Peroxidizing Herbicide: Correlation between Differential Inhibition of Protoporphyrinogen IX Oxidase Activity and Induction of Tetrapyrrole Accumulation by the Enantiomers. *Plant Physiol.* **1992**, *100*, 1211–1216. [[CrossRef](#)]
136. Duke, S.O.; Lydon, J.; Becerril, J.M.; Sherman, T.D.; Lehen, L.P., Jr.; Matsumoto, H. Protoporphyrinogen Oxidase-Inhibiting Herbicides. *Weed Sci.* **1991**, *39*, 465–473. [[CrossRef](#)]
137. Scalla, R.; Matringe, M.; Camadro, J.-M.; Labbe, P. Recent Advances in the Mode of Action of Diphenyl Ethers and Related Herbicides. *Z. Für Nat. C* **1990**, *45*, 503–511. [[CrossRef](#)]
138. Camadro, J.-M.; Matringe, M.; Scalla, R.; Labbe, P. Kinetic studies on protoporphyrinogen oxidase inhibition by diphenyl ether herbicides. *Biochem. J.* **1991**, *277*, 17–21. [[CrossRef](#)] [[PubMed](#)]
139. Mance, G. Introduction. In *Pollution Threat of Heavy Metals in Aquatic Environments*; Springer: Dordrecht, The Netherlands, 1987; pp. 1–8. [[CrossRef](#)]
140. El-Rayis, O.A.; Abouldahab, O.; Halim, Y.; Riley, J.P. Levels of Trace Metals in Some Food Chain Organisms from El-Mex Bay, West of Alexandria, Egypt. In *Proceedings of the 7th International Conference: Environment Protection is a Must*, Alexandria University and USPD, Alexandria, Egypt, 21–23 May 1997; pp. 20–22.
141. Soualili, D.; Dubois, P.; Gosselin, P.; Pernet, P.; Guillou, M. Assessment of seawater pollution by heavy metals in the neighbourhood of Algiers: Use of the sea urchin, *Paracentrotus lividus*, as a bioindicator. *ICES J. Mar. Sci.* **2008**, *65*, 132–139. [[CrossRef](#)]
142. Simkiss, K. Cellular Discrimination Processes in Metal Accumulating Cells. *J. Exp. Biol.* **1981**, *94*, 317–327. [[CrossRef](#)]
143. Williams, R.J.P.; Coombs, T.L.; Tinker, P.B. Physico-chemical aspects of inorganic element transfer through membranes. *Philos. Trans. R. Soc. B Biol. Sci.* **1981**, *294*, 57–74. [[CrossRef](#)]
144. Bae, J.H.; Lim, S.Y. Heavy Metals and Biochemical Composition of Four Sea Bream Species (*Acanthopagrus Schlegelii* Bleeker, *Pagrus Major* Temminck & Schlegel, *Oplegnathus Fasciatus* Krøyer and *Girella Punctata* Gray). *Philipp. Agric. Sci.* **2012**, *95*, 185–191.
145. Dallinger, R. Metabolism an Toxicity of Metals: Metallothionines and Metal Elimination. In *Cell Biology in Environment Toxicology*; Cajaraville, M.P., Ed.; Universidade del Pais Vasco: Bilbao, Spain, 1995; pp. 171–190.

146. Suzuki, K.T.; Suzuki, T. An Introduction to Clinical Aspects of Toxicity. In *Toxicology of Metals*; Chang, W., Ed.; LCRC Lewis Publishers: Boca Raton, FL, USA, 1996; pp. 333–335.
147. Engel, D.W.; Sunda, W.G. Toxicity of cupric ion to eggs of the spot *Leiostomus xanthurus* and the Atlantic silverside *Menidia menidia*. *Mar. Biol.* **1979**, *50*, 121–126. [[CrossRef](#)]
148. De Souza Machado, A.A.; Spencer, K.; Kloas, W.; Toffolon, M.; Zarfl, C. Metal fate and effects in estuaries: A review and conceptual model for better understanding of toxicity. *Sci. Total Environ.* **2016**, *541*, 268–281. [[CrossRef](#)]
149. Ardelan, M.V.; Steinnes, E.; Lierhagen, S.; Linde, S.O. Effects of experimental CO₂ leakage on solubility and transport of seven trace metals in seawater and sediment. *Sci. Total Environ.* **2009**, *407*, 6255–6266. [[CrossRef](#)]
150. Millero, F.; Woosley, R.J.; Ditrolio, B.; Waters, J. Effect of Ocean Acidification on the Speciation of Metals in Seawater. *Oceanography* **2009**, *22*, 72–85. [[CrossRef](#)]
151. López, I.R.; Kalman, J.; Vale, C.; Blasco, J. Influence of sediment acidification on the bioaccumulation of metals in *Ruditapes philippinarum*. *Environ. Sci. Pollut. Res.* **2010**, *17*, 1519–1528. [[CrossRef](#)]
152. Millero, F.J.; Ditrolio, B.R. Use of Thermodynamics in Examining the Effects of Ocean Acidification. *Elements* **2010**, *6*, 299–303. [[CrossRef](#)]
153. Lacoue-Labarthe, T.; Martin, S.; Oberhänsli, F.; Teyssié, J.-L.; Markich, S.; Jeffree, R.; Bustamante, P. Effects of increased pCO₂ and temperature on trace element (Ag, Cd and Zn) bioaccumulation in the eggs of the common cuttlefish, *Sepia officinalis*. *Biogeosciences* **2009**, *6*, 2561–2573. [[CrossRef](#)]
154. Pascal, P.-Y.; Fleeger, J.W.; Galvez, F.; Carman, K.R. The toxicological interaction between ocean acidity and metals in coastal meiobenthic copepods. *Mar. Pollut. Bull.* **2010**, *60*, 2201–2208. [[CrossRef](#)]
155. Ivanina, A.V.; Beniash, E.; Etkorn, M.; Meyers, T.B.; Ringwood, A.H.; Sokolova, I.M. Short-term acute hypercapnia affects cellular responses to trace metals in the hard clams *Mercenaria mercenaria*. *Aquat. Toxicol.* **2013**, *140–141*, 123–133. [[CrossRef](#)]
156. Myint, U.M.; Tyler, P.A. Effects of temperature, nutritive and metal stressors on the reproductive biology of *Mytilus edulis*. *Mar. Biol.* **1982**, *67*, 209–223. [[CrossRef](#)]
157. Parry, H.; Pipe, R. Interactive effects of temperature and copper on immunocompetence and disease susceptibility in mussels (*Mytilus edulis*). *Aquat. Toxicol.* **2004**, *69*, 311–325. [[CrossRef](#)] [[PubMed](#)]
158. de Mora, S.; Fowler, S.W.; Wyse, E.; Azemard, S. Distribution of heavy metals in marine bivalves, fish and coastal sediments in the Gulf and Gulf of Oman. *Mar. Pollut. Bull.* **2004**, *49*, 410–424. [[CrossRef](#)] [[PubMed](#)]
159. Valko, M.; Morris, H.; Cronin, M.T.D. Metals, Toxicity and Oxidative Stress. *Curr. Med. Chem.* **2005**, *12*, 1161–1208. [[CrossRef](#)] [[PubMed](#)]
160. Martelli, A.; Rousset, E.; Dycke, C.; Bouron, A.; Moulis, J.-M. Cadmium toxicity in animal cells by interference with essential metals. *Biochimie* **2006**, *88*, 1807–1814. [[CrossRef](#)] [[PubMed](#)]
161. Cherkasov, A.A.; Overton, R.A.; Sokolov, E.P.; Sokolova, I.M. Temperature-dependent effects of cadmium and purine nucleotides on mitochondrial aconitase from a marine ectotherm, *Crassostrea virginica*: A role of temperature in oxidative stress and allosteric enzyme regulation. *J. Exp. Biol.* **2007**, *210 Pt 1*, 46–55. [[CrossRef](#)]
162. Cannino, G.; Ferruggia, E.; Luparello, C.; Rinaldi, A.M. Cadmium and mitochondria. *Mitochondrion* **2009**, *9*, 377–384. [[CrossRef](#)]
163. Jomova, K.; Valko, M. Advances in metal-induced oxidative stress and human disease. *Toxicology* **2011**, *283*, 65–87. [[CrossRef](#)] [[PubMed](#)]
164. Al-Malki, A.L.; Moselhy, S.S. Impact of pesticides residue and heavy metals on lipids and fatty acids composition of some seafoods of Red Sea (KSA). *Hum. Exp. Toxicol.* **2011**, *30*, 1666–1673. [[CrossRef](#)]
165. Petroody, S.S.A.; Hamidian, A.H.; Ashrafi, S.; Eagderi, S.; Khazae, M. Investigation of Body Size Effect on Bioaccumulation Pattern of Cd, Pb and Ni in the Soft Tissue of Rock Oyster *Saccostrea Cucullata* from Laft Port Alavian. *J. Persian Gulf Mar. Sci.* **2013**, *4*, 39–45.
166. MacFarlane, G.R.; Schreider, M.; McLennan, B. Biomarkers of Heavy Metal Contamination in the Red Fingered Marsh Crab, *Parasesarma erythodactyla*. *Arch. Environ. Contam. Toxicol.* **2006**, *51*, 584–593. [[CrossRef](#)]
167. Sharma, R.; Agrawal, M. Biological effects of heavy metals: An overview. *J. Environ. Biol.* **2005**, *26*, 301–313. [[PubMed](#)]
168. Rainbow, P. Ecophysiology of Trace Metal Uptake in Crustaceans. *Estuar. Coast. Shelf Sci.* **1997**, *44*, 169–176. [[CrossRef](#)]
169. Krishnamurti, C.R.; Viswanathan, P. *Toxic Metals in the Indian Environment*; Tata McGraw-Hill Limited: New Delhi, India, 1991.
170. McLaughlin, M.J.; Parker, D.R.; Clarke, J.M. Metals and micronutrients—food safety issues. *Field Crop. Res.* **1999**, *60*, 143–163. [[CrossRef](#)]
171. U.S. Department of Health and Human Services. *Toxicological Profile for Copper*; U.S. Department of Health and Human Services: Washington, DC, USA, 2004. [[CrossRef](#)]
172. Francisco, C.D.M.; Bertolino, S.M.; Júnior, R.J.D.O.; Morelli, S.; Pereira, B.B. Genotoxicity assessment of polluted urban streams using a native fish *Astyanax altiparanae*. *J. Toxicol. Environ. Health Part A* **2019**, *82*, 514–523. [[CrossRef](#)] [[PubMed](#)]
173. Vázquez-Boucard, C.; Anguiano-Vega, G.; Mercier, L.; Del Castillo, E.R. Pesticide Residues, Heavy Metals, and DNA Damage in Sentinel Oysters *Crassostrea Gigas* From Sinaloa and Sonora, Mexico. *J. Toxicol. Environ. Health Part A* **2014**, *77*, 169–176. [[CrossRef](#)] [[PubMed](#)]
174. Jeong, C.-B.; Kang, H.-M.; Lee, M.-C.; Byeon, E.; Park, H.G.; Lee, J.-S. Effects of polluted seawater on oxidative stress, mortality, and reproductive parameters in the marine rotifer *Brachionus koreanus* and the marine copepod *Tigriopus japonicus*. *Aquat. Toxicol.* **2018**, *208*, 39–46. [[CrossRef](#)]

175. Gabryelak, T.; Filipiak, A.; Brichon, G. Effects of zinc on lipids of erythrocytes from carp (*Cyprinus carpio* L.) acclimated to different temperatures. *Comp. Biochem. Physiol. Part C Pharmacol. Toxicol. Endocrinol.* **2000**, *127*, 335–343. [[CrossRef](#)] [[PubMed](#)]
176. Weis, J.S.; Windham, L.; Santiago-Bass, C.; Weis, P. Growth, survival, and metal content of marsh invertebrates fed diets of detritus from *Spartina alterniflora* Loisel. and *Phragmites australis* Cav. Trin. ex Steud. from metal-contaminated and clean sites. *Wetl. Ecol. Manag.* **2002**, *10*, 71–84. [[CrossRef](#)]
177. Kwok, K.W.; Leung, K.M.; Bao, V.W.; Lee, J.-S. Copper toxicity in the marine copepod *Tigropus japonicus*: Low variability and high reproducibility of repeated acute and life-cycle tests. *Mar. Pollut. Bull.* **2008**, *57*, 632–636. [[CrossRef](#)] [[PubMed](#)]
178. Petersen, R.C. Population and Guild Analysis for Interpretation of Heavy Metal Pollution in Streams. In *Community Toxicity Testing*; ASTM International: West Conshohocken, PA, USA, 1986; pp. 180–198. [[CrossRef](#)]
179. Luoma, S.N.; Rainbow, P.S. *Metal Contamination in Aquatic Environments: Science and Lateral Management*; Cambridge University Press: Cambridge, UK, 2008.
180. Krupanidhi, S.; Sreekumar, A.; Sanjeevi, C.B. Copper & biological health. *Indian J. Med. Res.* **2008**, *128*, 448–461. [[PubMed](#)]
181. Cotou, E.; Henry, M.; Zeri, C.; Rigos, G.; Torreblanca, A.; Catsiki, V.-A. Short-term exposure of the European sea bass *Dicentrarchus labrax* to copper-based antifouling treated nets: Copper bioavailability and biomarkers responses. *Chemosphere* **2012**, *89*, 1091–1097. [[CrossRef](#)] [[PubMed](#)]
182. Eskandari, S.; Mozaffari, V. Interactive Effect of Soil Salinity and Copper Application on Growth and Chemical Composition of Pistachio Seedlings (cv. Badami). *Commun. Soil Sci. Plant Anal.* **2014**, *45*, 688–702. [[CrossRef](#)]
183. Uriu-Adams, J.Y.; Rucker, R.B.; Commisso, J.F.; Keen, C.L. Diabetes and dietary copper alter ⁶⁷Cu metabolism and oxidant defense in the rat. *J. Nutr. Biochem.* **2005**, *16*, 312–320. [[CrossRef](#)]
184. Barman, T.E. *Enzyme Handbook*; Springer: Berlin/Heidelberg, Germany, 1974. [[CrossRef](#)]
185. Santore, R.C.; Di Toro, D.M.; Paquin, P.R.; Allen, H.E.; Meyer, J.S. Biotic ligand model of the acute toxicity of metals. 2. application to acute copper toxicity in freshwater fish and daphnia. *Environ. Toxicol. Chem.* **2001**, *20*, 2397–2402. [[CrossRef](#)]
186. Zeri, C.; Hatzianestis, I. Distribution of total dissolved and C18 extractable copper and nickel in relation to dissolved organic matter sources, in the Thermaikos Gulf (eastern Mediterranean). *J. Mar. Syst.* **2005**, *58*, 143–152. [[CrossRef](#)]
187. Thomas, K.V.; Brooks, S. The environmental fate and effects of antifouling paint biocides. *Biofouling* **2010**, *26*, 73–88. [[CrossRef](#)]
188. Gaetke, L.M. Copper toxicity, oxidative stress, and antioxidant nutrients. *Toxicology* **2003**, *189*, 147–163. [[CrossRef](#)] [[PubMed](#)]
189. Viarengo, A.; Canesi, L.; Pertica, M.; Poli, G.; Moore, M.; Orunesu, M. Heavy metal effects on lipid peroxidation in the tissues of *mytilus galloprovincialis* lam. *Comp. Biochem. Physiol. Part C Comp. Pharmacol.* **1990**, *97*, 37–42. [[CrossRef](#)]
190. Frasco, M.F.; Fournier, D.; Carvalho, F.; Guilhermino, L. Do metals inhibit acetylcholinesterase (AChE)? Implementation of assay conditions for the use of AChE activity as a biomarker of metal toxicity. *Biomarkers* **2005**, *10*, 360–375. [[CrossRef](#)]
191. Muhvich, A.G.; Jones, R.T.; Kane, A.S.; Anderson, R.S.; Reimscheuessel, R. Effects of chronic copper exposure on the macrophage chemiluminescent response and gill histology in goldfish (*Carassius auratus* L.). *Fish Shellfish. Immunol.* **1995**, *5*, 251–264. [[CrossRef](#)]
192. Canesi, L.; Ciacci, C.; Piccoli, G.; Stocchi, V.; Viarengo, A.; Gallo, G. In vitro and in vivo effects of heavy metals on mussel digestive gland hexokinase activity: The role of glutathione. *Comp. Biochem. Physiol. Part C Pharmacol. Toxicol. Endocrinol.* **1998**, *120*, 261–268. [[CrossRef](#)]
193. de Almeida, E.A.; Miyamoto, S.; Bainy, A.C.D.; de Medeiros, M.H.G.; Di Mascio, P. Protective effect of phospholipid hydroperoxide glutathione peroxidase (PHGPx) against lipid peroxidation in mussels *Perna perna* exposed to different metals. *Mar. Pollut. Bull.* **2004**, *49*, 386–392. [[CrossRef](#)]
194. Doyotte, A.; Cossu, C.; Jacquín, M.-C.; Babut, M.; Vasseur, P. Antioxidant enzymes, glutathione and lipid peroxidation as relevant biomarkers of experimental or field exposure in the gills and the digestive gland of the freshwater bivalve *Unio tumidus*. *Aquat. Toxicol.* **1997**, *39*, 93–110. [[CrossRef](#)]
195. Maria, V.; Bebianno, M. Antioxidant and lipid peroxidation responses in *Mytilus galloprovincialis* exposed to mixtures of benzo(a)pyrene and copper. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **2011**, *154*, 56–63. [[CrossRef](#)]
196. Hall, L.W.H.; Scott, M.C.; Killen, W.D. Ecological risk assessment of copper and cadmium in surface waters of Chesapeake Bay watershed. *Environ. Toxicol. Chem.* **1998**, *17*, 1172–1189. [[CrossRef](#)]
197. Xie, Z.-C.; Wong, N.-C.; Qian, P.-Y.; Qiu, J.-W. Responses of polychaete *Hydroides elegans* life stages to copper stress. *Mar. Ecol. Prog. Ser.* **2005**, *285*, 89–96. [[CrossRef](#)]
198. Piola, R.F.; Johnston, E.L. Pollution reduces native diversity and increases invader dominance in marine hard-substrate communities. *Divers. Distrib.* **2008**, *14*, 329–342. [[CrossRef](#)]
199. Lawes, J.C.; Clark, G.F.; Johnston, E.L. Contaminant cocktails: Interactive effects of fertiliser and copper paint on marine invertebrate recruitment and mortality. *Mar. Pollut. Bull.* **2016**, *102*, 148–159. [[CrossRef](#)]
200. Sierra, M.; Sanhueza, A.; Alcántara, R.; Sánchez, G. Antimicrobial evaluation of copper sulfate (II) on strains of *Enterococcus faecalis*. In vitro study. *J. Oral Res.* **2013**, *2*, 114–118. [[CrossRef](#)]
201. Sun, Q.; Hu, K.; Yang, X. The Efficacy of Copper Sulfate in Controlling Infection of *Saprolegnia parasitica*. *J. World Aquac. Soc.* **2014**, *45*, 220–225. [[CrossRef](#)]
202. Abramova, A.; Gedanken, A.; Popov, V.; Ooi, E.-H.; Mason, T.J.; Joyce, E.M.; Beddow, J.; Perelshtein, I.; Bayazitov, V. A sonochemical technology for coating of textiles with antibacterial nanoparticles and equipment for its implementation. *Mater. Lett.* **2013**, *96*, 121–124. [[CrossRef](#)]

203. Das, D.; Nath, B.C.; Phukon, P.; Dolui, S.K. Synthesis and evaluation of antioxidant and antibacterial behavior of CuO nanoparticles. *Colloids Surfaces B Biointerfaces* **2013**, *101*, 430–433. [[CrossRef](#)]
204. Geng, J.-J.; Dimkpa, C.O.; Calder, A.; Britt, D.W.; McLean, J.E.; Anderson, A.J. Responses of a soil bacterium, *Pseudomonas chlororaphis* O6 to commercial metal oxide nanoparticles compared with responses to metal ions. *Environ. Pollut.* **2011**, *159*, 1749–1756. [[CrossRef](#)]
205. İpeksaç, T.; Kaya, F.; Kaya, C. Template-free hydrothermal method for the synthesis of multi-walled CuO nanotubes. *Mater. Lett.* **2014**, *130*, 68–70. [[CrossRef](#)]
206. Mageshwari, K.; Sathyamoorthy, R. Flower-shaped CuO Nanostructures: Synthesis, Characterization and Antimicrobial Activity. *J. Mater. Sci. Technol.* **2013**, *29*, 909–914. [[CrossRef](#)]
207. Sathyamoorthy, R.; Mageshwari, K. Synthesis of hierarchical CuO microspheres: Photocatalytic and antibacterial activities. *Phys. E Low Dimens. Syst. Nanostructures* **2013**, *47*, 157–161. [[CrossRef](#)]
208. Sohrabnezhad, S.; Moghaddam, M.M.; Salavatiyan, T. Synthesis and characterization of CuO–montmorillonite nanocomposite by thermal decomposition method and antibacterial activity of nanocomposite. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **2014**, *125*, 73–78. [[CrossRef](#)]
209. Sonia, S.; Jayram, N.D.; Kumar, P.S.; Mangalaraj, D.; Ponpandian, N.; Viswanathan, C. Effect of NaOH concentration on structural, surface and antibacterial activity of CuO nanorods synthesized by direct sonochemical method. *Superlattices Microstruct.* **2014**, *66*, 1–9. [[CrossRef](#)]
210. Dewez, D.; Geoffroy, L.; Vernet, G.; Popovic, R. Determination of photosynthetic and enzymatic biomarkers sensitivity used to evaluate toxic effects of copper and fludioxonil in alga *Scenedesmus obliquus*. *Aquat. Toxicol.* **2005**, *74*, 150–159. [[CrossRef](#)]
211. Subramanian, B.; Priya, K.A.; Rajan, S.T.; Dhandapani, P.; Jayachandran, M. Antimicrobial activity of sputtered nanocrystalline CuO impregnated fabrics. *Mater. Lett.* **2014**, *128*, 1–4. [[CrossRef](#)]
212. Zabrieski, Z.; Morrell, E.; Hortin, J.; Dimkpa, C.O.; McLean, J.E.; Britt, D.; Anderson, A.J. Pesticidal activity of metal oxide nanoparticles on plant pathogenic isolates of *Pythium*. *Ecotoxicology* **2015**, *24*, 1305–1314. [[CrossRef](#)]
213. Song, L.-Y.; Wang, Y.-Q. Investigation of microbial community structure of a shallow lake after one season copper sulfate algacide treatment. *Microbiol. Res.* **2015**, *170*, 105–113. [[CrossRef](#)] [[PubMed](#)]
214. Stevens, M.; Doran, G.; Mo, J. Efficacy and environmental fate of copper sulphate applied to Australian rice fields for control of the aquatic snail *Isidorella newcombi*. *Crop. Prot.* **2014**, *63*, 48–56. [[CrossRef](#)]
215. Sengco, M.R. Prevention and control of *Karenia brevis* blooms. *Harmful Algae* **2009**, *8*, 623–628. [[CrossRef](#)]
216. Dumme, V.; Tanhan, P.; Kruatrachue, M.; Damrongphol, P.; Pokethitoyook, P. Histopathological changes in snail, *Pomacea canaliculata*, exposed to sub-lethal copper sulfate concentrations. *Ecotoxicol. Environ. Saf.* **2015**, *122*, 290–295. [[CrossRef](#)]
217. Nguyen, T.T.H.; Li, S.; Li, J. The combined effects of copper sulfate and rosin sizing agent treatment on some physical and mechanical properties of poplar wood. *Constr. Build. Mater.* **2013**, *40*, 33–39. [[CrossRef](#)]
218. Halpern, M.; Gasith, A.; Teltsch, B.; Porat, R.; Broza, M. Chloramine and Copper Sulphate as Control Agents of Planktonic Larvae of *Chironomus luridus* in Water Supply Systems. *J. Am. Mosq. Control. Assoc.* **1999**, *15*, 453–457.
219. Straus, D.L.; Farmer, B.D.; Ledbetter, C.K.; Beck, B.H.; Williams, R.S.; Clark, M.L.; Freeze, T.M. Use of Copper Sulfate to Control Egg Saprolegniasis at a Commercial Sunshine Bass Hatchery. *N. Am. J. Aquac.* **2016**, *78*, 243–250. [[CrossRef](#)]
220. Amorim, M.J.B. The Daunting Challenge of Ensuring Sustainable Development of Nanomaterials. *Int. J. Environ. Res. Public Health* **2016**, *13*, 245. [[CrossRef](#)] [[PubMed](#)]
221. Vance, M.E.; Kuiken, T.; Vejerano, E.P.; McGinnis, S.P.; Hochella, M.F., Jr.; Rejeski, D.; Hull, M.S. Nanotechnology in the real world: Redeveloping the nanomaterial consumer products inventory. *Beilstein J. Nanotechnol.* **2015**, *6*, 1769–1780. [[CrossRef](#)] [[PubMed](#)]
222. Pang, C.; Selck, H.; Misra, S.K.; Berhanu, D.; Dybowska, A.; Valsami-Jones, E.; Forbes, V.E. Effects of sediment-associated copper to the deposit-feeding snail, *Potamopyrgus antipodarum*: A comparison of Cu added in aqueous form or as nano- and micro-CuO particles. *Aquat. Toxicol.* **2012**, *106–107*, 114–122. [[CrossRef](#)] [[PubMed](#)]
223. Ramskov, T.; Selck, H.; Banta, G.; Misra, S.K.; Berhanu, D.; Valsami-Jones, E.; Forbes, V.E. Bioaccumulation and effects of different-shaped copper oxide nanoparticles in the deposit-feeding snail *Potamopyrgus antipodarum*. *Environ. Toxicol. Chem.* **2014**, *33*, 1976–1987. [[CrossRef](#)] [[PubMed](#)]
224. Keller, A.A.; Adeleye, A.S.; Conway, J.R.; Garner, K.L.; Zhao, L.; Cherr, G.N.; Hong, J.; Gardea-Torresdey, J.L.; Godwin, H.A.; Hanna, S.; et al. Comparative environmental fate and toxicity of copper nanomaterials. *Nanoimpact* **2017**, *7*, 28–40. [[CrossRef](#)]
225. Scanes, P. ‘Oyster watch’: Monitoring trace metal and organochlorine concentrations in Sydney’s coastal waters. *Mar. Pollut. Bull.* **1996**, *33*, 226–238. [[CrossRef](#)]
226. Dafforn, K.A.; Lewis, J.A.; Johnston, E.L. Antifouling strategies: History and regulation, ecological impacts and mitigation. *Mar. Pollut. Bull.* **2011**, *62*, 453–465. [[CrossRef](#)]
227. Johnston, E.; Marzinelli, E.; Wood, C.; Speranza, D.; Bishop, J. Bearing the burden of boat harbours: Heavy contaminant and fouling loads in a native habitat-forming alga. *Mar. Pollut. Bull.* **2011**, *62*, 2137–2144. [[CrossRef](#)]
228. WHO. *Guidelines for Drinking-Water Quality: Fourth Edition Incorporating the First and Second Addenda*; WHO: Geneva, Switzerland, 2022.
229. Mebane, C.A.; Schmidt, T.S.; Miller, J.L.; Balistrieri, L.S. Bioaccumulation and Toxicity of Cadmium, Copper, Nickel, and Zinc and Their Mixtures to Aquatic Insect Communities. *Environ. Toxicol. Chem.* **2020**, *39*, 812–833. [[CrossRef](#)]
230. Grosell, M. Copper. *Fish Physiol.* **2011**, *31*, 53–133. [[CrossRef](#)]

231. Shioi, Y.; Tamai, H.; Sasa, T. Inhibition of Photosystem II in the Green Alga *Ankistrodesmus falcatus* by Copper. *Physiol. Plant.* **1978**, *44*, 434–438. [[CrossRef](#)]
232. Küpper, H.; Šetlík, I.; Spiller, M.; Küpper, F.C.; Prášil, O. Heavy metal-induced inhibition of photosynthesis: Targets of in vivo heavy metal chlorophyll formation. *J. Phycol.* **2002**, *38*, 429–441. [[CrossRef](#)]
233. McElroy, D.J.; Doblin, M.A.; Murphy, R.J.; Hochuli, D.F.; Coleman, R.A. A limited legacy effect of copper in marine biofilms. *Mar. Pollut. Bull.* **2016**, *109*, 117–127. [[CrossRef](#)] [[PubMed](#)]
234. Nguyen, L.T.; Janssen, C.R. Embryo-Larval Toxicity Tests with the African Catfish (*Clarias gariepinus*): Comparative Sensitivity of Endpoints. *Arch. Environ. Contam. Toxicol.* **2002**, *42*, 256–262. [[CrossRef](#)]

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