

Solubilization of Polyelectrolytic Hairy-Rod Polyfluorene in Aqueous Solutions of Nonionic Surfactant

Matti Knaapila,^{*,†} László Almásy,[‡] Vasil M. Garamus,[§] Christopher Pearson,[⊥] Swapna Pradhan,^{||} Michael C. Petty,[⊥] Ullrich Scherf,^{||} Hugh D. Burrows,[#] and Andrew P. Monkman[†]

Department of Physics, University of Durham, South Road, Durham DH1 3LE, United Kingdom, Research Institute for Solid State Physics and Optics, P.O. Box 49, Budapest-1525, Hungary, GKSS Research Centre, Max-Planck-Strasse 1, D-21502 Geesthacht, Germany, School of Engineering, University of Durham, South Road, Durham DH1 3LE, United Kingdom, Makromolekulare Chemie und Institut für Polymertechnologie, Gauss-Str. 20, Bergische Universität Wuppertal, D-42097 Wuppertal, Germany, and Departamento de Química, Universidade de Coimbra, P-3004-535 Coimbra, Portugal

Received: October 22, 2005; In Final Form: March 24, 2006

We report on the solubilization, phase behavior, and self-organized colloidal structure of a ternary water–polyfluorene–surfactant (amphiphile) system comprised of polyelectrolytic poly{1,4-phenylene[9,9-bis(4-phenoxybutylsulfonate)]fluorene-2,7-diyl} (PBS–PFP) in nonionic pentaethylene glycol monododecyl ether (C₁₂E₅) at 20 °C. We show in particular how a high amount (milligrams per milliliter) of polyfluorene can be solubilized by aqueous C₁₂E₅ via aggregate formation. The PBS–PFP and C₁₂E₅ concentrations of 0.31×10^{-4} – 5×10^{-4} M and 2.5×10^{-4} – 75×10^{-4} M, respectively, were used. Under the studied conditions, the photoluminescence (PL), surface tension, static contact angle, and (π -A) isotherm measurements imply that D₂O–PBS–PFP(C₁₂E₅)_x realizes three phase regimes with an increasing molar ratio of surfactant over monomer unit (x). First, for $x \leq 0.5$, the mixture is cloudy. In this regime polymer is only partially dissolved. Second, for $1 \leq x \leq 2$, the solution is homogeneous. In this regime polymer is dissolved down to the colloidal level. Small-angle neutron scattering (SANS) patterns indicate rigid elongated (polymer–surfactant) aggregates with a diameter of 30 Å and mean length of ~ 900 Å. The ratio between contour length and persistence length is less than 3. Third, for $x \geq 4$, the solution is homogeneous and there is cooperative binding between polymer and surfactant. Surface tension, contact angle, and surface pressure remain essentially constant with increasing x . A PL spectrum characteristic of single separated polyfluorene molecules is observed. SANS curves show an interference maximum at $q \sim 0.015 \text{ \AA}^{-1}$, indicating an ordered phase. This ordering is suggested to be due to the electrostatic repulsion between polymer molecules adsorbed on or incorporated into the C₁₂E₅ aggregates (micelles). On dilution the distance between micelles increases via 3-dimensional packing. In this regime the polymer is potentially dissolved down to the molecular level. We show further that the aggregates ($x = 2$) form a floating layer at the air–water interface and can be transferred onto hydrophilic substrates.

1. Introduction

There are myriad reasons to develop water soluble π -conjugated polymers and tailor their phase behavior, structure, and supramolecules in water. Water soluble π -conjugated polymers form an option for interrogating biological substrates.^{1,2} The inkjet process and layer-by-layer self-assembly—the major advantages of polymers over oligomers in light emitting diode (LED) fabrication—benefit from electroluminescent materials in nonnoxious solvent, harmless for the cartridges.³ A tuning of the air–water interface of π -conjugated polymers⁴ is a base of Langmuir–Blodgett (LB) films⁵ having opportunities in sensors.⁶ Dissolving π -conjugated “model” polymers in water can also promote general understanding of the self-assembly of π -conjugated hairy-rod polymers.⁷

Control of the solubility of π -conjugated polymers is a nontrivial task. Few such polymers can be dissolved in water. A typical strategy to achieve water solution is to introduce neutral or charged hydrophilic functionalities to the terminal position of the polymer backbone. Examples include cationic^{8,9} and sulfonated¹⁰ poly(*p*-phenylene) (PPP) and sulfonated poly(*p*-phenylenevinylene) (PPV), poly[5-methoxy-2-(4-sulfoxybutoxy)-1,4-phenylene-vinylene] (MBL-PPV).¹¹ Water soluble polyfluorenes (PFs) used in DNA peptide nucleic acid detection¹² belong to this class of materials. However, the water solubility of PFs is difficult to achieve in *high* concentrations and cosolvents such as methanol are employed to ensure the uniformity of the solutions.¹³ Another strategy to assist solubility of π -conjugated polymers is a surfactant layer separating polymer and solvent. The interaction between polymer and surfactant can be due to either hydrophobic–hydrophilic effects¹⁴ or strong physical bonds.¹⁵ Examples include polyelectrolytic PPV¹⁶ and PF.^{14,15,17} This technique can also be used to induce liquid crystallinity as shown for PPP^{18,19} and poly(*p*-pyridine).²⁰

* Address correspondence to this author. Phone: +44-191-33-43558. Fax: +44-191-33-43585. E-mail: matti.knaapila@durham.ac.uk.

[†] Department of Physics, University of Durham.

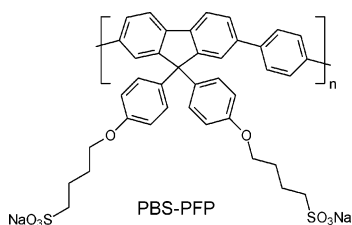
[‡] Research Institute for Solid State Physics and Optics.

[§] GKSS Research Centre.

[⊥] School of Engineering, University of Durham.

^{||} Bergische Universität Wuppertal.

[#] Departamento de Química, Universidade de Coimbra.

CHART 1: Chemical Structure of the PBS–PFP Copolymer

Oxyethylene glycol-monoethers (C_iE_j)^{21,22} are archetypical surfactants widely used in fundamental phase studies of microemulsions,^{23–27} block-*co*-polymers,²⁸ and micelles.^{29,30} The phase behavior of pentaethylene glycol monododecyl ether (*n*-dodecyl pentaerythritol glycol ether) ($C_{12}E_5$)^{31,32} is particularly well-known: Pure $C_{12}E_5$ forms cylindrical micelles but the oil-swollen $C_{12}E_5$ micelles are wormlike with a rather high flexibility and cannot be described as rigid rods.^{33,30}

Several C_iE_j s can be used to solubilize selected PPPs^{34,35} and PFS.^{14,36} A particular example¹⁴ concerns poly{[1,4-phenylene-[9,9-bis(4-phenoxybutylsulfonate)]fluorene-2,7-diyl} copolymer (or PBS–PFP), see Chart 1. PBS–PFP is not water soluble. However, when the concentration of PBS–PFP is 8.3×10^{-6} M and 0–10 mol of $C_{12}E_5$ with respect to the polymer's monomer unit ($0-6 \times 10^{-4}$ M) are added, the constituents form a transparent solution with optical changes at the threshold surfactant concentration of 5×10^{-5} M. This concentration is close to the critical micelle concentration (CMC) of binary aqueous $C_{12}E_5$, 5×10^{-5} M.²² It has been further demonstrated how this results in consequent fluorescence enhancement.¹⁴

In this paper we present a further investigation of the solubilization of an aqueous PBS–PFP/ $C_{12}E_5$ mixture at concentrations much higher than those used in the previous study.¹⁴ A feasibility study for thin film formation from a water subphase is also reported. A high PBS–PFP concentration of 5×10^{-4} M (i.e. 10 times that of the earlier study¹⁴) was specifically selected as it might better allow the formation of thin films from water. The studied surfactant concentration regime (2.5×10^{-4} – 100×10^{-4} M) and temperature (20 °C) would correspond to the L_1 phase (notation for isotropic liquid introduced in ref 37) of the binary water– $C_{12}E_5$ system, yet exceeding its CMC.

Three phase regimes with different characteristics were observed by photoluminescence (PL), surface tension, contact angle, and (π - A) isotherm measurements. The structural evolution of observed phases was probed by using small-angle neutron scattering (SANS). Ternary D_2O –PBS–PFP($C_{12}E_5$)_{*x*} solution proves to be homogeneous when the polymer concentration is 5×10^{-4} M and when the molar ratio of surfactant over monomer, *x*, is around 1, defining the first phase boundary. The SANS data of the ternary system differ from that of a binary aqueous surfactant. Elongated objects (mean length ~ 900 Å) with near circular cross section (diameter ~ 30 Å) are seen for $x = 1-2$. These objects are stiffer and thinner than $C_{12}E_5$ micelles in the same concentration regime. This observation indicates aggregate rather than micelle formation in the ternary system. Increasing the surfactant concentration, at $x = 4$, an interference maximum appears at $q \sim 0.015$ Å⁻¹ indicating ordering of the micelles with a characteristic separation distance of 40 nm, defining the second phase boundary. The ordering is suggested to be due to electrostatic repulsion rather than steric hindrance. The ternary aggregates form a floating layer at the air–water interface, which enables thin film formation from water.

2. Experimental Section

Materials. The preparation of PBS–PFP ($M_n = 6.5$ kg/mol) (Chart 1) has been described in refs 14 and 15. For studying the polymer–solvent system, PBS–PFP was dissolved in deuterated dimethyl sulfoxide (DMSO) (>99.8%; Euriso-top) to a concentration of 1.3 mg/mL. The ternary polymer–surfactant–water solutions were prepared with D_2O (>99.9%; GOSS Scientific Instruments Ltd) and $C_{12}E_5$ (Sigma) to form the D_2O –PBS–PFP($C_{12}E_5$)_{*x*} system where *x* is the molar ratio of $C_{12}E_5$ with respect to the polymer's monomer unit. The concentration of polymer in heavy water is given with respect to monomer units (not with respect to the whole polymer) and it varied from 0.3×10^{-4} to 5×10^{-4} M while *x* varied from 0.5 to 15 corresponding to the surfactant concentration of $\sim 2.5 \times 10^{-4}$ – 75×10^{-4} M. All samples were prepared by combining D_2O mixtures of the pure constituents. Stirring for 1 day and ultrasound agitation for 10 min were used to ensure uniformity of the solutions. To avoid any degradation, fresh surfactant was used and samples were measured within a few days of preparation.

Thin Films. The (π - A) isotherms were measured and nominal Langmuir monolayers formed with use of a LB715 Langmuir–Blodgett trough (Molecular Photonics). The samples were spread onto the surface of a pure water subphase with a microliter syringe. The films were compressed at a speed of 9.4×10^{-5} – 4.2×10^{-4} nm²/(s·molecule), and a dipping speed of 0.080 mm/s was used for transfer to solid substrates consisting of (hydrophilic) glass plates or glass plates made highly hydrophobic with a 2% solution of dimethyldichlorosilane in 1,1,1-trichloroethane (BDH). Silicon wafers were used to provide a parallel set of samples to glass. In every case the temperature was 20 ± 1 °C.

PL Measurements. PL measurements of solutions were performed with a Jobin-Yvon Fluoromax fluorimeter and quartz cell at 20 ± 1 °C. The excitation wavelength was 370 nm and monochromator resolution 1 nm. As the chromophore concentration was constant for the samples studied, the effect of self-absorption to the peak position could be ignored. PL measurements of films were done on a Jobin-Yvon Fluorolog spectrometer. The excitation wavelength was 350 nm and monochromator resolution 1 nm. Measurements were carried out either immediately or within 2 days after film preparation and the results were consistent within this period.

Surface Tension Measurements. Surface tension was measured with the microbalance of a LB715 Langmuir–Blodgett trough (Molecular Photonics), using the Wilhelmy plate method at 20 ± 1 °C. D_2O gave a surface tension of $\sigma = 72.5 \pm 1.0$ mN/m, corresponding to that of distilled water. Experiments were repeated at least three times to ensure consistent values.

Contact Angle Measurements. For contact angle measurements, solution drops were studied on glass plates whose surfaces had been made highly hydrophobic with a 2% solution of dimethyldichlorosilane in 1,1,1-trichloroethane (BDH). Measurements were performed with a horizontal Vickers microscope equipped with a Canon EO5 digital camera at 20 ± 1 °C. This method gave a contact angle of $88 \pm 0.5^\circ$ and $89 \pm 0.5^\circ$ for distilled water and D_2O , respectively. Measurements were reproduced several times. The saturation time for wetting was 60 s for each sample corresponding to that observed for $C_{12}E_5$ /oil systems on hydrophobic surfaces in ref 38.

SANS Measurements. SANS experiments were made with the *Yellow Submarine* instrument at the Budapest Neutron

Centre in Budapest, Hungary,³⁹ and with the SANS-1 instrument at the GKSS research center in Geesthacht, Germany.⁴⁰ Several sample-to-detector distances (from 0.7 to 9.7 m) and wavelengths (from 5.7 to 11.4 Å) were employed to cover the range of the magnitude of scattering vectors q from 0.005 to 0.3 Å⁻¹. The wavelength resolution of the *Yellow Submarine* is 20% (full-width at half-maximum) while that of the SANS-1 is 10%. Hellma quartz cells with 5 mm path length were filled with the sample solution and placed in a thermostated holder, kept at 20.0 ± 0.5 °C. The raw spectra were corrected for sample transmission, room background, and sample cell scattering by conventional procedures.⁴¹ The 2D isotropic scattering patterns were azimuthally averaged, converted to an absolute scale, and corrected for detector efficiency by dividing by the incoherent scattering spectra of 1 mm thick pure water. The scattering from D₂O used for the sample preparation was subtracted as a background; the small incoherent scattering due to the protonated surfactant and polymer was disregarded. The data for one sample were collected for 6 h on average.

3. Analysis of SANS Data

Local Structure. The absolute intensity SANS patterns were analyzed by using a model independent indirect Fourier transform method (IFT) developed by Glatter.⁴² Here we considered noninteracting micelles at the q -range 0.02–0.3 Å⁻¹ and took instrumental resolution into account according to Pedersen.⁴³ In the case of elongated particles, the IFT approach allows the decoupling of the model scattering function and fitting the cross-sectional structure at the q -regime dominated by local rigidity of aggregates. The asymptotic behavior of the scattering function for $q > 1/l_p$ (l_p is the persistence length) can be expressed as

$$I(q) = \frac{\pi}{q} 2\pi \int_0^\infty \tilde{p}_{CS}(r) J_0(qr) r dr = \frac{\pi}{q} I_{CS}(q) \quad (1)$$

where J_0 is the zeroth-order Bessel function and $I_{CS}(q)$ is the cross-sectional scattering intensity.

The normalized cross-sectional distance distribution function $\tilde{p}_{CS}(r)$ is given by

$$\tilde{p}_{CS}(r) = \frac{c}{2\pi M_L} \int \Delta\rho(\mathbf{r}) \Delta\rho(\mathbf{r} + \mathbf{r}') d\mathbf{r}' \quad (2)$$

where ρ is the scattering length density, c the overall concentration, and M_L the mass per length, and where the vectors \mathbf{r} and \mathbf{r}' are lying in the cross-sectional plane.

From $\tilde{p}_{CS}(r)$ we calculated the integral parameters of the aggregate cross section. The cross-sectional radius of gyration, $R_{CS,g}$, is given by

$$R_{CS,g} = \left[\frac{\int_0^\infty r^2 \tilde{p}_{CS}(r) dr}{\int_0^\infty \tilde{p}_{CS}(r) dr} \right]^{1/2} \quad (3)$$

In the case of homogeneous cylindrical object $R_{CS,g} = 2^{-1/2}R$, where R is the radius of cross section. The cross-sectional forward scattered intensity $I_{CS}(0)$ is given by

$$I_{CS}(0) = 2\pi \int_0^\infty \tilde{p}_{CS}(r) dr \quad (4)$$

which allows us to calculate the mass per length, M_L (in units g/cm), via

$$M_L = \frac{I_{CS}(0)}{\Delta\rho_m^2} \quad (5)$$

Overall Size and Flexibility of Aggregates. The length and flexibility of rodlike aggregates were obtained by fitting the scattering curves by two models: stiff and flexible cylinders. In both cases the scattering intensity is written as

$$I(q) \sim S_L(q, L, l_p) S_{CS}(q, R_{CS}) \quad (6)$$

where for the model of flexible cylinders $S_L(q, L, l_p)$ is the scattering function of a flexible cylinder of contour length L and persistence length l_p . Expression $S_L(q, L, l_p)$ contains the single-cylinder scattering function for a flexible cylinder with excluded-volume effects and is written as

$$S_L(q, L, l_p) = \{ [1 - \zeta(q, L, l_p)] S_{chain}(q, L, l_p) + \zeta(q, L, l_p) S_{rod}(q, L) \} G(q, L, l_p) \quad (7)$$

where $S_{chain}(q, L, l_p)$ is the scattering function of a flexible (cylindrical) chain with excluded-volume effects, $S_{rod}(q, L)$ the scattering function of a rod, and $\zeta(q, L, l_p)$ a crossover function, and where the function $G(q, L, l_p)$ corrects the crossover region. Detailed expressions can be found in refs 44–46.

In the case of a stiff rod $S_L(q, L, l_p)$ reduces to $S_{rod}(q, L)$, which represents the scattering of an infinitely thin cylinder of length L as

$$S_{rod}(q, L) = \frac{2Si(qL)}{qL} 4 \sin^2\left(\frac{qL/2}{q^2 L^2}\right) \quad (8)$$

where $Si(x) = \int_0^x t^{-1} \sin t dt$.

For some cases, the aggregates could not be modeled by cylindrical cross section. There we used models of elliptical cross section. The scattering function in this case is written as

$$S_{CS}(q, A, B) = \frac{2}{\pi} \int_0^{\pi/2} \left(\frac{2J_1(qr(A, B, \theta))}{qr(A, B, \theta)} \right)^2 d\theta \quad (9)$$

where J_1 is the first-order Bessel function, $r(A, B, \theta) = [A^2 \sin^2\theta + B^2 \cos^2\theta]^{1/2}$, and A and B are the semi-axes of the ellipse. A circular cross section corresponds to $A = B$.

Aggregation Number from Position of Maximum. In the case where an interference maximum is observed in the scattering curves, it is possible to obtain the aggregation number from the position of this maximum. Here, one assumes that the observed maximum corresponds to the (111) planes of a fcc lattice.⁴⁷ Aggregation number m is expressed via maximum position q_{max} as

$$m = \frac{1}{4} \left(\frac{2\pi 3^{1/2}}{q_{max}} \right)^3 N_A c \quad (10)$$

where N_A is Avogadro number and c is the concentration of molecules. Knowing the volume of one molecule, the size and volume of the aggregates can be obtained. If the aggregates have strong interaction, this approximation gives similar results to direct modeling.⁴⁸

4. Results

Solution Formation. Solution formation was studied by visual inspection. Figure 1 depicts essential visual examples of the solutions. In the studied concentration regimes (0.3×10^{-4} – 5×10^{-4} M) PBS–PFP is not soluble in water but precipitated

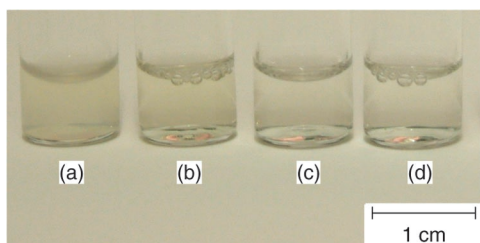


Figure 1. (a–d) A photograph of D_2O –PBS–PFP($C_{12}E_5$) $_x$ for $x = 0, 1, 2,$ and $10,$ respectively, suggesting solution formation. The PBS–PFP concentration is 5×10^{-4} M for all x .

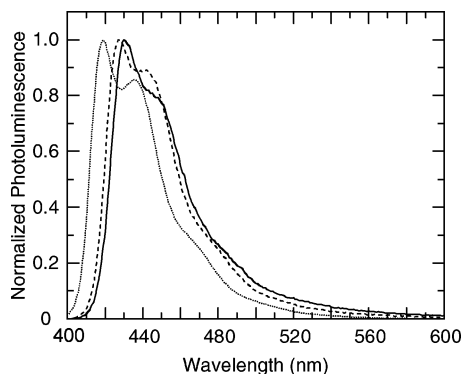


Figure 2. PL spectra of PBS–PFP (solid line) and PBS–PFP($C_{12}E_5$) $_x$ for $x = 2$ (dashed line) and 8 (dotted line) in D_2O at 20 °C. The PBS–PFP concentration is 5×10^{-4} M for all x .

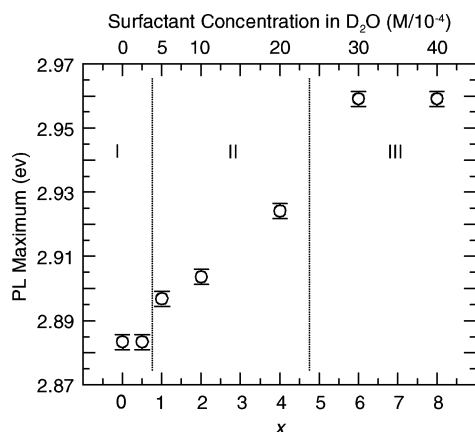


Figure 3. PL maximum of D_2O –PBS–PFP($C_{12}E_5$) $_x$ as a function of molar ratio x at 20 °C. The PBS–PFP concentration is 5×10^{-4} M for all x . The dotted lines distinguish tentative phase regimes I–III.

particles are always observed. Aqueous solutions of pure $C_{12}E_5$ (2.5×10^{-4} – 100×10^{-4} M) are naturally transparent. When mixing these constituents we see a clear transition so that PBS–PFP($C_{12}E_5$) $_x$ appeared visually cloudy for $x < 1$ and perfectly transparent (or slightly bluish due to the fluorescence) for $x \geq 1$.

Phase Boundaries. Phase behavior was probed by using PL, surface tension (σ), static contact angle, and (π – A) isotherm measurements. Figure 2 plots essential examples of the PL spectra of D_2O –PBS–PFP($C_{12}E_5$) $_x$ as a function of molar ratio x . Figure 3 plots in turn the position of the 0–0 maximum as a function of x . A progressive blue shift of the whole PL spectrum, with the spectral maximum shifting from 430 to 419 nm, occurs on surfactant addition.

Figure 4 plots surface tension and static contact angle on hydrophobic surfaces. When PBS–PFP is added to D_2O , σ drops some 5 mN/m. Adding surfactant causes σ to drop further until it approaches the level of the binary system at $x \sim 4$. On average,

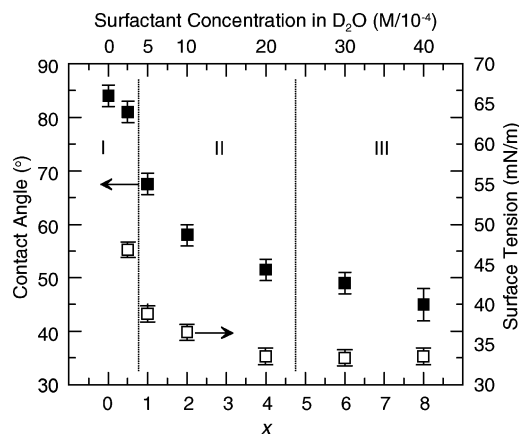


Figure 4. Contact angle (solid squares) and surface tension (open squares) of D_2O –PBS–PFP($C_{12}E_5$) $_x$ as a function of molar ratio x . The dotted lines distinguish tentative phase regimes I–III. $T = 20$ °C.

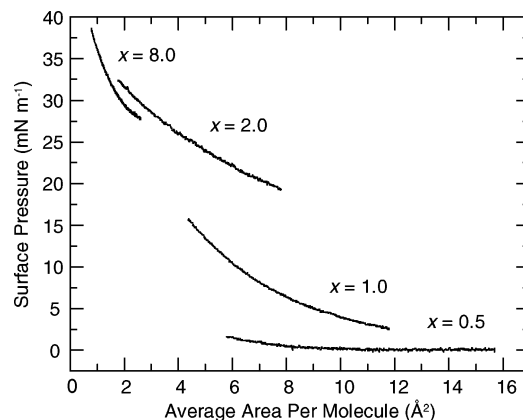


Figure 5. Characteristic (π – A) isotherms of PBS–PFP($C_{12}E_5$) $_x$ solutions for $x = 0.5, 1, 2,$ and 8 measured with a compression speed of 9.4×10^{-5} – 4.2×10^{-4} nm 2 /(s·molecule) at a temperature of 20 ± 1 °C.

the surface tension of the ternary system for $x > 4$ is somewhat higher than that of equally concentrated binary ones. Static contact angle behaves much like surface tension approaching that of pure D_2O for small x and that of binary solution for $x > 4$ yet remaining higher than those of equally concentrated binary systems. The results obtained are compiled in Table 1.

Figure 5 plots typical surface pressure–area (π – A) isotherms of PBS–PFP($C_{12}E_5$) $_x$ on a pure water subphase. For $x = 0.5, 1.0, 2.0,$ and $8.0,$ 0.22 mL of solution with concentrations of 0.46, 0.56, 0.77, and 1.99 mg/mL, respectively, was spread. Three phenomenological regimes can be recognized corresponding to those shown in Figures 3 and 4. When precipitated macroscopic particles of PBS–PFP in water are seen for $x = 0.5,$ the surface pressure approaches zero. The transparent mixture with higher surfactant fractions ($x = 1$) results in an expanded isotherm, with a distinct upturn with decreasing surface area. This indicates the formation of a floating layer at the water–air interface. The surface pressure increases when x is increased from 1 to 2 but essentially saturates for $x > 2$.

Inspecting Figures 3–5, three different regimes of dissolution of PBS–PFP in the $C_{12}E_5$ /water mixture are observed. At lower concentrations of $C_{12}E_5$ ($x < 1$) the rapidly decreasing surface tension and contact angle and approximately solid state PL indicate that interaction between polymer and surfactant is quite low and that surfactant molecules are predominantly free in the solution. We denote this as Regime I.

At intermediate concentration of $C_{12}E_5$ $x = 1$ – 4 the emerging blue shift in PL points to the gradual dissolution of the polymer,

TABLE 1: Materials Parameters of the Studied Samples in D₂O at 20 °C

material	monomer concn c_m (M)	surfactant concn c_s (M)	overall concn (mg/mL)	visual appearance	first PL max (nm)	surface tension σ (mN/m)	contact angle (deg)
C ₁₂ E ₅	0	2.5×10^{-4}	0.10 ± 0.02	transparent	n/a	32.5 ± 1.0	85.5 ± 1.0
C ₁₂ E ₅	0	10×10^{-4}	0.41 ± 0.02	transparent	n/a	30.6 ± 1.0	40.0 ± 2.0
C ₁₂ E ₅	0	30×10^{-4}	1.22 ± 0.02	transparent	n/a	31.0 ± 1.0	38.5 ± 2.0
C ₁₂ E ₅	0	100×10^{-4}	4.07 ± 0.02	transparent	n/a	31.1 ± 1.0	33.5 ± 2.0
PBS–PFP	5×10^{-4}	0	0.36 ± 0.02	precipitated particles	430	n/a	84.0 ± 0.5
PBS–PFP	2.5×10^{-4}	0	0.18 ± 0.02	precipitated particles		n/a	
PBS–PFP(C ₁₂ E ₅) _{0.5}	5×10^{-4}	2.5×10^{-4}	0.46 ± 0.02	cloudy but no particles	430	46.8 ± 1.0	81.0 ± 0.5
PBS–PFP(C ₁₂ E ₅) _{1.0}	5×10^{-4}	5×10^{-4}	0.56 ± 0.02	transparent	430	38.8 ± 1.0	67.5 ± 0.5
PBS–PFP(C ₁₂ E ₅) _{2.0}	5×10^{-4}	10×10^{-4}	0.77 ± 0.02	transparent	426	36.5 ± 1.0	58.0 ± 2.0
PBS–PFP(C ₁₂ E ₅) _{4.0}	5×10^{-4}	20×10^{-4}	1.17 ± 0.02	transparent	424	33.5 ± 1.0	51.5 ± 2.0
PBS–PFP(C ₁₂ E ₅) _{6.0}	5×10^{-4}	30×10^{-4}	1.58 ± 0.02	transparent	419	33.3 ± 1.0	49.0 ± 2.0
PBS–PFP(C ₁₂ E ₅) _{8.0}	5×10^{-4}	40×10^{-4}	1.99 ± 0.02	transparent	419	33.5 ± 1.0	45.0 ± 2.0
PBS–PFP(C ₁₂ E ₅) _{10.0}	5×10^{-4}	50×10^{-4}	2.39 ± 0.02	transparent		31.1 ± 1.0	42.0 ± 2.0

while the moderately decreasing surface tension and contact angle shows the saturation of the free surfactant concentration, i.e., that there is a strong polymer–surfactant interaction. This could be somewhat analogous to the critical aggregate concentration (CAC) of colloidal science. This is Regime II. In this regime, surfactant molecules are suggested to be partly located close to the polymer and partly at the interface. In other words, the interaction between polymer and surfactant may not be strong enough to catch all surfactant molecules and we may call this *noncooperative* binding of C₁₂E₅.

Finally at high $x > 4$ we observe constant values of all parameters, which indicates that the fraction of free C₁₂E₅ is constant and that the surplus C₁₂E₅ is bound at the aggregates (polymer–surfactant or pure surfactant). This means that we observe *cooperative* binding of C₁₂E₅. We denote this as Regime III. We note in the end that no macrophase separation occurred after 6 months so the transparent solutions seem to represent an equilibrium state.

Particle Structure in Aqueous Solution. The nanoscale structure of the particles in aqueous solution was studied with SANS. To obtain information on the conformation of the pure PBS–PFP, we dissolved it first in deuterated DMSO at 0.1 wt %, which resulted in a transparent solution. The SANS curve of this solution is, however, not distinguishable from the background. The small molecular weight of the polymer (~ 10 units) and the incorporation of the side chains into the solvent prevented measurements of its structure as could be done for a high molecular weight PF^{49,50} dissolved in *d*-toluene.

Cloudy Mixtures of D₂O–PBS–PFP(C₁₂E₅)_x at the Low Surfactant Fraction Limit $x \rightarrow 0$. As ternary D₂O–PBS–PFP–(C₁₂E₅)_x for $x \leq 0.5$ is visually cloudy, the SANS method will detect only that fraction of the colloidal objects which fall into

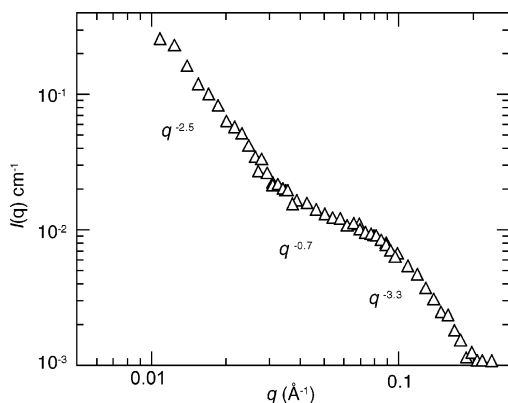


Figure 6. SANS data of D₂O–PBS–PFP(C₁₂E₅)_{0.5} for a monomer concentration of 5×10^{-4} M. $T = 20$ °C.

its observation window ($< 2\pi/q_{\min} \approx 1000$ Å). Figure 6 plots the scattering curves for the ternary system at low surfactant concentration, $x = 0.5$. The power law scattering at the lowest q -range indicates the presence of large aggregates. For $q > 0.03$ Å⁻¹, the scattering curve resembles those seen in Regime II (see later, Figure 7). This indicates the appearance of aggregates with characteristic sizes of the order of 100 Å. At the highest q , the intensity decreases as $\sim q^{-3.3}$. Altogether, we see that in Regime I the polymer crystals start to dissolve with the assistance of the surfactant.

Visually Transparent Solutions of D₂O–PBS–PFP(C₁₂E₅)_x at Intermediate Surfactant Fraction, $x = 1-4$. Figure 7 plots SANS data of D₂O–PBS–PFP(C₁₂E₅)_{2.0} and D₂O–C₁₂E₅ of the corresponding surfactant concentration as well as fits obtained from the IFT. The scattering intensity follows a power law $\sim q^{-1}$ over wide q -range. Figure 8 plots the cross-sectional pair correlation functions obtained from the IFT at $q > 0.02$ Å⁻¹. Table 2 summarizes the cross-sectional parameters. The upper limits for the cross-sectional pair correlation function are found to be 34 Å for the aggregates in the ternary system and 44 Å in the binary system. The radius of the cross section obtained by circular approximation is smaller for PBS–PFP–(C₁₂E₅)_x aggregates with $x = 1$ and 2 than for C₁₂E₅ micelles at a similar concentration. The values of mass per unit length obtained are in agreement with the values of radii and show that PBS–PFP(C₁₂E₅)_x aggregates with $x = 1$ and 2 consist of

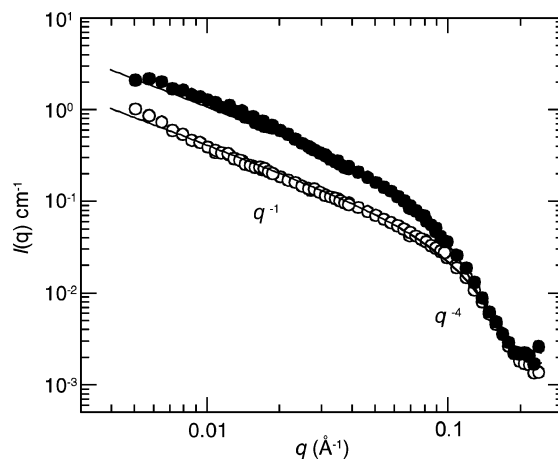


Figure 7. SANS data of D₂O–PBS–PFP(C₁₂E₅)_{2.0} with a monomer concentration of 5×10^{-4} M (open symbols) and D₂O–C₁₂E₅ with a concentration of 10×10^{-4} M (solid symbols, intensities were multiplied by 2 for clarity), which corresponds to the surfactant concentration of the case $x = 2$ as well as the corresponding fits obtained by IFT. $T = 20$ °C.

TABLE 2: Cross Sectional Parameters of Rodlike Aggregates of PBS–PFP(C₁₂E₅)_x and Micelles of C₁₂E₅ Obtained by Model Independent Analysis^a

material	monomer concn c_m (M)	surfactant concn c_s (M)	$R_{CS,g}$ (Å)	R (Å)	$M_L, 10^{-13}$ (g/cm)	no. of monomers/nm	no. of C ₁₂ E ₅ molecules/nm
C ₁₂ E ₅	0	2.5×10^{-4}	14.5 ± 0.2	20.5	0.99	0	15
C ₁₂ E ₅	0	10×10^{-4}	15.2 ± 0.3	21.5	1.01	0	15
C ₁₂ E ₅	0	30×10^{-4}	15.6 ± 0.3	22.1	1.02	0	15
PBS–PFP(C ₁₂ E ₅) _{1.0}	5×10^{-4}	5×10^{-4}	10.2 ± 0.3	14.1	0.389	2	2
PBS–PFP(C ₁₂ E ₅) _{2.0}	5×10^{-4}	10×10^{-4}	11.6 ± 0.2	16.4	0.513	2	4

^a R is the radius of the cross section in homogeneous and circular approximation of the cross section. M_L is the average mass of aggregates per 1 cm, also the present number of monomer of PBS–PFP(C₁₂E₅) per 1 nm and the number of C₁₂E₅ molecules per 1 nm in the case of aggregates and pure micelles.

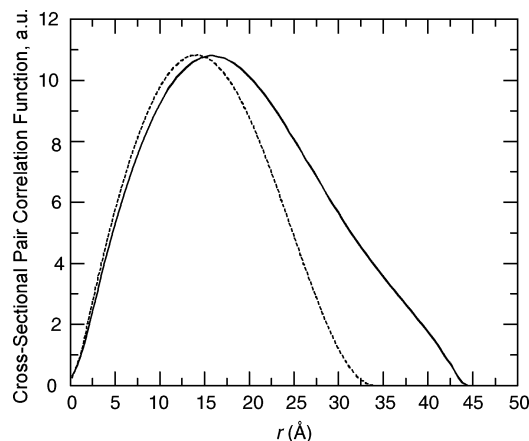


Figure 8. $\bar{p}_{CS}(r)$ obtained by IFT from the scattering curves: solid line for D₂O–C₁₂E₅ with a concentration of 10×10^{-4} M and dashed line for D₂O–PBS–PFP(C₁₂E₅)_{2.0} with a monomer concentration of 5×10^{-4} M. $T = 20$ °C.

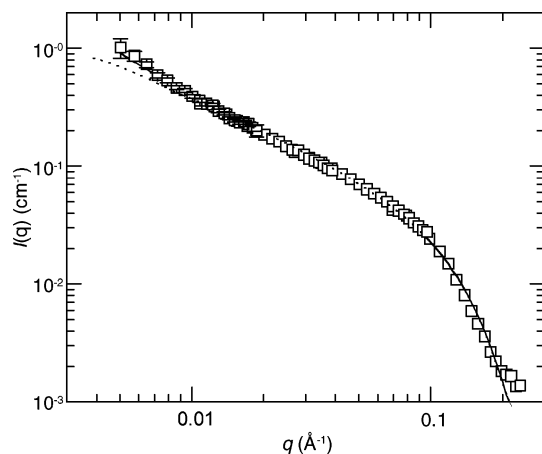


Figure 9. SANS data of PBS–PFP(C₁₂E₅)_{2.0} with a monomer concentration of 5×10^{-4} M and fit by flexible (solid line) and stiff cylinder models (dashed line). $T = 20$ °C.

2 repeat units and 2–4 surfactant molecules per nanometer. In contrast, C₁₂E₅ micelles consist of ~ 15 molecules per nanometer.

Figure 9 plots the data of D₂O–PBS–PFP(C₁₂E₅)_{2.0} and corresponding fits to the models of a stiff and flexible cylinder. The cross section is nearly circular. The flexible model results in a contour length of 900 ± 100 Å, a persistence length of 300 ± 30 Å, and a radius $R = 15.2 \pm 0.5$ Å while the stiff cylinder model yields a length of 880 ± 80 Å and a radius $R = 15.4 \pm 0.5$ Å for D₂O–PBS–PFP(C₁₂E₅)_{2.0}. The application of the flexible model to SANS data of equally dense D₂O–C₁₂E₅ (10×10^{-4} M) solution yields the values of 750 ± 80 and 120 ± 10 Å, respectively, for the contour and persistence lengths. The cross section is not circular. Fitting it with the

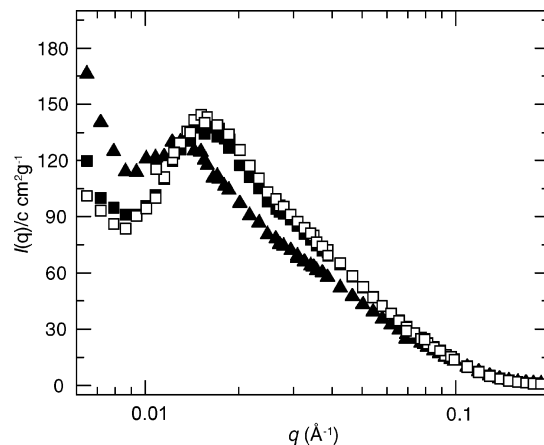


Figure 10. SANS data of D₂O–PBS–PFP(C₁₂E₅)_x for a monomer concentration of 5×10^{-4} M with $x = 4$ (solid triangles), 6 (solid squares), and 8 (open squares) normalized to a total concentration of PBS–PFP and C₁₂E₅. $T = 20$ °C.

ellipsoid model results in values of semi-axes $A = 26.0 \pm 0.5$ Å and $B = 16.0 \pm 0.5$ Å.

Overall, the solutions are transparent and SANS data suggest the formation of elongated aggregates which are not C₁₂E₅ micelles. The aggregates are thinner (with nearly circular cross section) and have lower mass per unit length than corresponding pure micelles. Two quite different models yield similar characteristics of the aggregates' size. The stiff cylinder model fits well apart from lowest q (cf. Figure 9) while the flexible cylinder model fits well over the whole q -range. However, the ratio between total length and persistence length is < 3 , which suggests that the aggregates are stiff rather than flexible.

Visually Transparent Solutions of D₂O–PBS–PFP(C₁₂E₅)_x at the High Surfactant Fraction, $x > 4$. Figure 10 plots SANS data of D₂O–PBS–PFP(C₁₂E₅)_x for $x > 2$. The scattering intensities are normalized to the total concentration of PBS–PFP and C₁₂E₅. A clear transition is seen when the molar ratio is increased to $x = 4$. A distinct interference maximum appears at 0.015 – 0.02 Å⁻¹ (the scattering angle increasing with x) and potentially a very broad maximum at 0.025 – 0.035 Å⁻¹. The position of this maximum is the same for $x = 6$ and 8, which implies that the intermicellar distance does not change much with increasing x . The values of normalized intensities are higher for $x = 8$ than for $x = 6$, which implies the growth of aggregates with increasing x . The determination of the number of PBS–PFP repeat units from the position of the interference maximum (eq 9) yields ~ 20 PBS–PFP repeat units and ~ 160 surfactant molecules per one aggregate at $x = 8$. This means that each aggregate contains two polymers on average. The characteristic dimension of the aggregates is of the order of 40 Å.

Figure 11 plots the SANS data of D₂O–PBS–PFP(C₁₂E₅)_{8.0} on dilution by D₂O. Figure 12 plots the position of the maximum as a function of concentration for $x = 8$ and 10. We observe

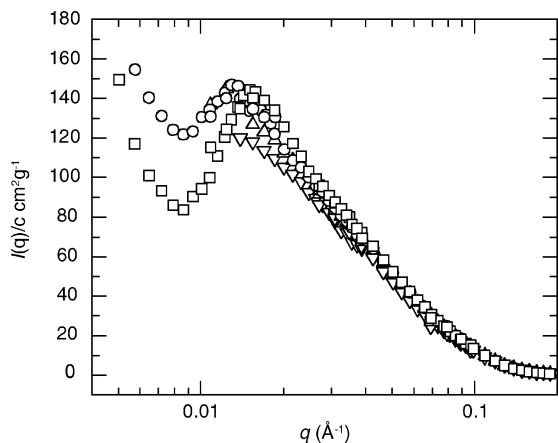


Figure 11. SANS data of D_2O -PBS-PFP($C_{12}E_5$) $_x$ for $x = 8$ with a PBS-PFP concentration of 5×10^{-4} (squares), 2.5×10^{-4} (spheres), 1.25×10^{-4} (upper triangles), and 0.625×10^{-4} M (lower triangles). $T = 20$ °C.

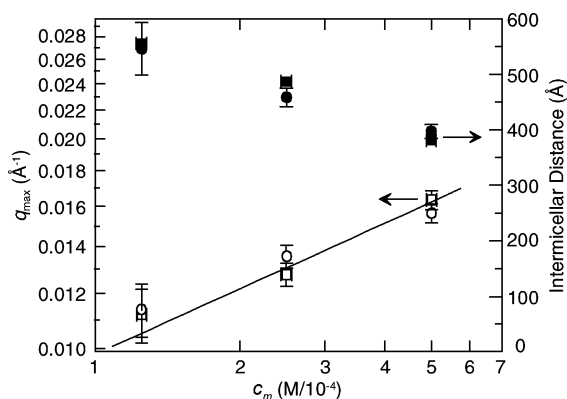


Figure 12. The q_{\max} (open symbols) and intermicellar distance (solid symbols) vs monomer concentration c_m corresponding to $x = 8$ (squares) and 10 (spheres). The solid line is a linear fit of q_{\max} for $x = 8$. $T = 20$ °C.

TABLE 3: The Intermicellar Distance and Surface Tension of D_2O -PBS-PFP($C_{12}E_5$) $_x$ as a Function of Molar Ratio x and Overall Concentration

molar ratio x	monomer concn c_m (M)	overall concn (mg/mL)	surface tension σ (mN/m)	interm. distance l (nm)
4.0	5×10^{-4}	1.17 ± 0.02	33.5 ± 1.0	48 ± 1
6.0	5×10^{-4}	1.58 ± 0.02	33.3 ± 1.0	42 ± 1
8.0	5×10^{-4}	1.99 ± 0.02	33.5 ± 1.0	38 ± 1
8.0	2.5×10^{-4}	0.99 ± 0.02	33.4 ± 1.0	49 ± 1
8.0	1.25×10^{-4}	0.50 ± 0.02	32.7 ± 1.0	55 ± 2
8.0	0.625×10^{-4}	0.25 ± 0.02	34.2 ± 1.0	n/a
8.0	0.3125×10^{-4}	0.12 ± 0.02	36.1 ± 1.0	n/a
10.0	5×10^{-4}	2.39 ± 0.02	31.1 ± 1.0	40 ± 1
10.0	2.5×10^{-4}	1.96 ± 0.02	32.2 ± 1.0	46 ± 1
10.0	1.25×10^{-4}	0.60 ± 0.02	32.3 ± 1.0	56 ± 2
10.0	0.625×10^{-4}	0.30 ± 0.02	32.9 ± 1.0	n/a

scaling $q_{\max} \sim c^{0.31 \pm 0.05} \approx c^{1/3}$, which suggests that the distance between micelles increases in a way of 3D packing.^{51,52} The interference maximum, which depends mainly on concentration of polymer according to 3D packing, implies that the concentration of $C_{12}E_5$ aggregates is determined by the number of active sites in polymer molecules. The intermicellar distance (long period) and surface tension as a function of molar ratio and concentration are displayed in Table 3. Surface tension stays constant on dilution, which further enforces the idea of cooperative binding.

Floating Aggregate Layer at the Air–Water Interface. The floating layers of PBS-PFP($C_{12}E_5$) $_x$ at the air–water interface

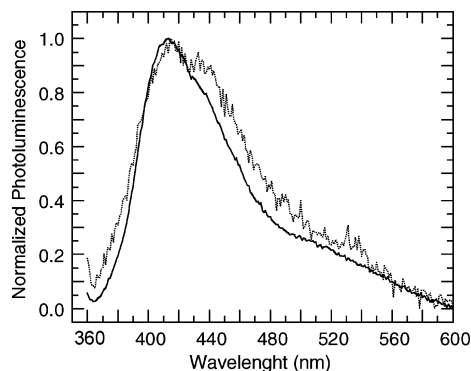


Figure 13. PL spectra of nominal PBS-PFP($C_{12}E_5$) $_{2.0}$ monolayers on hydrophilic (solid line) and hydrophobic glass (dashed line).

were studied by using Langmuir–Blodgett dipping with hydrophilic and hydrophobic glass substrates. Unsurprisingly, the dipping was unsuccessful for $x = 0.5$. When higher surfactant fractions were used and surface pressure kept constant, we observed a smooth kink in the slope of the interface area at the beginning of the dipping with hydrophilic substrate suggesting material transfer from the subphase. This effect was not seen with hydrophobic glass. As the highest amount of polymer was anticipated to be transferred by keeping the polymer fraction and surface pressure simultaneously high, we considered high surface pressure at the low surfactant fraction limit, i.e., $x = 2$. As the surface tension of PBS-PFP($C_{12}E_5$) $_{2.0}$ does not approach that of aqueous surfactant (cf. Figure 4), no substantial surfactant surplus was assumed. Figure 13 plots the PL spectra of nominal PBS-PFP($C_{12}E_5$) $_{2.0}$ monolayers on hydrophilic and hydrophobic glass showing the characteristic PL of polyfluorene.

5. Discussion

Solution Formation. In our work, the first “definition” of dissolution is the visual transparency and under the studied concentration regime this occurs for D_2O -PBS-PFP($C_{12}E_5$) $_x$ at $x \geq 1$. Formation of a transparent ternary system supports the previous assumption of the polymer particles broken by $C_{12}E_5$ surfactant.¹⁴ Nevertheless, bare visual consideration does not indicate that PBS-PFP would be dissolved down to the molecular level. It simply means that no gross macroscopic particles are seen.

Phase Boundaries. The binary water- $C_{12}E_5$ system is an appropriate starting point for our discussion. It reveals a CMC of around 5×10^{-5} M.²² Near room temperature it shows an isotropic liquid L_1 phase when the $C_{12}E_5$ fraction is below ~ 35 wt %, ³² which corresponds to a $C_{12}E_5$ concentration of $\sim 9 \times 10^{-1}$ M. The normal hexagonal H_1 phase is seen for higher surfactant concentrations. Transition from an L_1 to an $L_1' + L_1''$ phase occurs at $T = 35$ – 50 °C, increasing with $C_{12}E_5$ fraction.³² The present study was performed at 20 °C with the surfactant concentration between 2.5×10^{-4} and 75×10^{-4} M. Therefore, the studied “base” binary mixture at 20 °C would correspond to the L_1 phase above the CMC.

A progressive blue shift of the whole PL spectrum, with the spectral maximum shifting from 430 to 419 nm, occurs on suggestive polymer dissolution (Figure 2). The “solution PL” indicates that polymer chains are fully separated from each other but here its form does not depend on the nature of the surrounding media. This result differs slightly from the previous study that suggested a shift down to 412 nm,¹⁴ a fact that presumably stems from the lower overall concentration. The PL characteristics of water-PBS-PFP($C_{12}E_5$)¹⁴ can also be found elsewhere. The overall situation illustrated in Figure 3

gives us tentatively three phenomenological phase regimes: two possible plateaus for $x < 1$ (Regime I) and $x > 4$ (Regime III) where, respectively, the PL spectrum approaches that of the solid state and solution PL of PF, as well as the intermediate region for $x = 1-4$ (Regime II).

The idea of three phase regimes is supported by surface tension and static contact angle. Figure 4 illustrates regimes where σ approaches the values of pure D₂O and pure aqueous C₁₂E₅. In phase Regime II the values of σ are low but still higher than those in Regime III in which they are still slightly higher than those in binary systems. This suggests that the free surfactant molecules are present in Regime II at a concentration smaller than the CMC. These observations are in accordance with the changes in contact angle (Figure 4). As both surface tension and contact angle of aqueous PBS-PFP(C₁₂E₅)_x are consistently slightly larger than those of correspondingly concentrated aqueous C₁₂E₅, we expect that PBS-PFP binds part of C₁₂E₅. The surface tension²⁶ and contact angle⁵³ of C₁₂E₅ and C₁₂E₅/oil systems have been reported elsewhere.

Finally, three phenomenological regimes may be recognized in the (π -A) isotherms as well (Figure 5). When precipitated macroscopic particles of PBS-PFP in water are seen for $x = 0.5$, the surface pressure approaches zero indicating negligible film formation at the air-water interface. This supports the above-mentioned assumption that PBS-PFP is at or below the solubilization limit and binds the majority of surfactants for $x = 0.5$. The higher surfactant fraction ($x = 1$) leads to a floating layer at the water-air interface. The surface pressure increases rapidly with C₁₂E₅ fraction but saturates for $x \geq 2$, separating Regimes II and III. We observed some fluctuations when the measurements were repeated, which is potentially due to the instability of the layer. However, the gross deviation between the three regimes was reproduced.

In this framework, the solution structure of Regimes I-III was then studied by using SANS. We have shown in section 4 that the suggested regimes correspond to the SANS data, where the distinction between Regimes II and III is, however, observed between $x = 2$ and 4 rather than between $x = 1$ and 2. Below we discuss SANS data of D₂O-PBS-PFP(C₁₂E₅)_x for $x = 1-10$, show comparison to D₂O-C₁₂E₅ in similar conditions and provide an interpretation for the different regimes.

Particle Structure in Solution. Regime I, $x \leq 0.5$: Partial Solubilization. PBS-PFP is not water soluble but forms a cloudy mixture with a typical solid-state PL spectrum (cf. Figure 2). The existence of nondissolved particles prevents us from giving a reliable quantitative analysis. However, inspecting SANS data plotted in Figure 6, the following conclusions can be drawn. The slope -2.5 at smallest q can imply the presence of large, polydisperse or mass fractal objects ($> 1000 \text{ \AA}$). Assuming mass fractals, their fractal dimension D_m would be equal to the value of the slope.⁵⁴ The region with slope -0.7 appears to be crossover toward the scattering pattern from the smaller objects, which display a slope of -3.3 at highest q . This would correspond to a surface-fractal-like surface of these particles,⁵⁴ which are presumably aggregates formed by the dissolved polymers and surfactant molecules. Altogether, we propose that we observe large nondissolved polymer particles, and "mixed" surfactant-polymer aggregates, similar to what exist in Regime II (see below).

Regime II, $1 \leq x \leq 4$: Dissolved Aggregates. This regime is important as a low fraction of surfactant and potential solubilization are simultaneously achieved. We draw attention to the following features seen in Figures 7 and 8 following the classic texts.^{54,55} First, in contrast to the cloudy solutions at $x = 0.5$,

the increased scattering intensity seen in Figure 6 at $q < 0.02 \text{ \AA}^{-1}$ completely disappears. Consequently, the large polymer aggregates are completely dissolved for $x > 1$. Second, the SANS curves follow q^{-1} behavior over a wide range of q , which is characteristic of rodlike objects. At corresponding concentrations binary aqueous C₁₂E₅ follows roughly q^{-1} behavior as well, but the scattering curves clearly differ from those of the ternary system, the difference becoming apparent in the structural parameters obtained by the fitting procedures. Third, the reasonable fits in Figure 7 obtained with the IFT approach justify the modeling by elongated particles in both the binary and ternary mixtures. Small deviations at the lowest q reflect flexibility of elongated objects or the presence of significantly larger aggregates which are not taken into account by IFT. Fourth, Figure 8 indicates that the aggregates in the ternary system possess a smaller cross-section than the micelles in the binary system.

The scattering data were fitted to the models of stiff and flexible cylinders (Figure 9). The choice of the model of a stiff cylinder is justified by distinctive -1 scaling at the intermediate q -range and by the successful application of model independent IFT analysis. As this model can be an oversimplification and as the oil swollen surfactant micelles are known to be wormlike, we applied the flexible cylinder model as well. The flexible cylinder model gives a visually better fit at lowest q values (Figure 9), but this agreement does not improve the overall accuracy of the fit, considering the resulting χ^2 values ($\chi^2 = 2.44$ for the stiff cylinder model and 2.2 for the flexible cylinder model). As described above, the parameters obtained are similar as well. It is also noteworthy that the persistence length of PBS-PFP(C₁₂E₅)_{2.0} is actually larger than the overall length of PBS-PFP ($< 20 \text{ nm}$), a similar result as has been reported for the PPP-C₇E₃ system.³⁵

Furthermore, the data suggest that the aggregates in the ternary system are thinner than the pure surfactant micelles indicating that they do not consist of well-defined C₁₂E₅ micelles with a PBS-PFP core. We find that this is plausible, because there are only a very few surfactants per repeat unit (1-2). Therefore we believe that C₁₂E₅ will mainly aggregate with the polymer rather than form micelles and suggest that there is a *noncooperative* binding between PBS-PFP and C₁₂E₅. Fiber aggregates of PPP within surfactant micelles have been found elsewhere.³⁵ However, in that work, the mass ratio of surfactant over monomer unit was 10, i.e., much higher than in Regime II.

Regime III, $4 \leq x \leq 15$: Interacting Micelles and Cooperative Binding. As shown in Figures 10-12, an interference maximum at $\sim 0.015 \text{ \AA}^{-1}$ is seen for the ternary D₂O-PBS-PFP(C₁₂E₅)_x for $x \geq 4$. This peak corresponds to the maximum of the structure factor of the ordered micellar system. Upon dilution, the interference maximum shifts predictably toward lower q , and remains observable down to $\sim 1.25 \times 10^{-4} \text{ M}$. Essentially the same behavior is observed for all $x \geq 4$. We observe scaling $q_{\max} \sim c^{1/3}$ indicating 3D packing. This scaling behavior differs from that of aqueous poly(sodium *p*-phenylenesulfonate)¹⁰ and MBL-PPV¹¹ which show an interference maximum scaling $q_{\max} \sim c^{1/2}$.

We note that the H₁ phase of the binary system is formed only when the surfactant fraction is as high as 35 wt %.³² Therefore, the observed ordering in the considered dilute solutions ($\sim 1 \text{ wt } \%$) is certainly not caused by steric effects but rather by the electrostatic repulsive interactions induced by the charged PBS-PFP molecules. This ordering should not be a surprise as the counterion sensitive order and a reflection at

low q due to electrostatic repulsion have been reported also for example for aqueous PPP^{8–10} and MBL–PPV¹¹ polyelectrolytes.

A simple structure that could be considered for Regime III is that where the polymer is included in the hydrocarbon core of the C₁₂E₅ micelle, similar to the wormlike micelles in water–C₁₂E₅–decane³⁰ or water–alkyl-substituted PPP–C_iE_j³⁵ systems. Nevertheless, in this case the charges would be rather screened by the outer part of the micelle, thus diminishing the intermicellar repulsion, which is contrary to our observation. A further argument against this structure is that the polymer–surfactant aggregates observed in Regime II are thinner than the surfactant micelles. Another possibility for PBS–PFP is to be located at the outer surface of the micelle. This would better explain electrostatic repulsion between the micelles and be in agreement with the observed reflection. A rigorous answer cannot be given at present.

Floating Aggregate Layer at the Air–Water Interface. A natural application of the aqueous π -conjugated system is the controlled formation of Langmuir–Blodgett films from an aqueous subphase. Nondissolved PBS–PFP does not allow film formation but when surfactant is added, films on both hydrophobic and hydrophilic glass show a similar PL spectrum of PBS–PFP (Figure 13) suggesting that not only surfactant but also PBS–PFP has been transferred onto the surface. The blue shift in PL suggests that the PBS–PFP chains are separated, which is plausible assuming that the films are very thin. The PL intensity from the film on hydrophobic glass is considerably weaker than that from the otherwise identically dipped hydrophilic glass, which indicates in turn that the polymer-containing aggregates are attached much better on the hydrophilic than the highly hydrophobic surface. The observation of a floating layer of PBS–PFP(C₁₂E₅)_x is in accordance with the corresponding layer of alkyl-substituted PPP–C_iE_j.¹⁸

6. Conclusions

In summary, the solubilization of PBS–PFP polymer by C₁₂E₅ surfactant in water has been studied. The dissolution of the polymer was initially suggested by the visual transparency and a blue shift in the PL spectrum upon mixing the constituents. As indicated by this consideration, the polymer itself is not water soluble but can be solubilized by aqueous C₁₂E₅. The phase behavior of D₂O–PBS–PFP(C₁₂E₅)_x was then detailed by using the PL, surface tension, static contact angle, (π – A) isotherms, and SANS. A feasibility study of the film formation was also realized.

The D₂O–PBS–PFP(C₁₂E₅)_x system was specifically considered when the surfactant concentration was above the CMC of aqueous C₁₂E₅ at the temperature (20 °C) and concentration which correspond to the isotropic liquid L₁ phase regime of the binary D₂O–C₁₂E₅ system. Under these conditions, three ternary phase regimes were observed. First, the cloudy phase is seen for $x \leq 0.5$ (Regime I, partial solubilization) indicating partial dissolution of the polymer. Second, the D₂O–PBS–PFP(C₁₂E₅)_x solution turns homogeneous for $x \geq 1$ (Regime II, dissolved aggregates). The q^{-1} behavior of all the measured SANS curves indicates rodlike aggregates for $x = 1–2$, resembling the wormlike micelles of binary surfactant solutions. However, the scattering characteristics of the ternary system differ from those of the binary ones and the persistence length was found to be larger (up to 30 nm). Fitted data to the model of stiff and flexible cylinders indicate rodlike conformation for which the ratio between length and persistence length is < 3 . Interestingly, the aggregates in the ternary system were found to be less flexible and *thinner* than the micelles in the binary

ones. Third, for $x = 4–15$ (Regime III, interacting micelles), the SANS curve shows an interference peak at $q \sim 0.015 \text{ \AA}^{-1}$. This ordering is suggested to be due to the repulsion between the charged polymers attached to or incorporated into the micelles.

Altogether these results show that a high amount (milligrams per milliliter) of otherwise insoluble polyfluorene can be solubilized in water by using C₁₂E₅ via complicated aggregate formation. These aggregates form a floating layer at the air–water interface and can be transferred onto the hydrophilic substrate with use of Langmuir dipping.

Acknowledgment. This work has been funded by One North East (UK) UIC Nanotechnology Grant. L.A. thanks the Széchenyi István Scholarship Foundation for support. M.K. and L.A. thank the European Commission for support under the 6th Framework Program through the Key Action: Strengthening the European Research Area, Research Infrastructures (Contract No. RII3-CT-2003-505925).

References and Notes

- Heeger, P. S.; Heeger, A. J. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 12219–12221.
- Chen, L.; McBranch, D. W.; Wang, H.-L.; Helgeson, R.; Wudl, F.; Whitten, D. G. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 12287–12292.
- Sirringhaus, H. *Adv. Mater.* **2005**, *17*, 2411–2425.
- Breiby, D. W.; Samuelsen, E. J.; Kononov, O.; Struth, B. *Langmuir* **2004**, *20*, 4116–4123.
- Bjørnholm, T.; Hassenkam, T.; Reitzel, N. *J. Mater. Chem.* **1999**, *9*, 1975–1990.
- Levitsky, I. A.; Kim, J.; Swager, T. M. *Macromolecules* **2001**, *34*, 2315–2319.
- Knaapila, M.; Stepanyan, R.; Lyons, B. P.; Torkkeli, M.; Monkman, A. P. *Adv. Funct. Mater.* **2006**, *16*, 599–609.
- Guillaume, B.; Blaul, J.; Wittemann, M.; Rehahn, M.; Ballauff, M. *J. Phys.: Condens. Matter* **2000**, *12*, A245–A251.
- Guillaume, B.; Blaul, J.; Ballauff, M.; Wittemann, M.; Rehahn, M.; Goerigk, G. *Eur. Phys. J. E* **2002**, *8*, 299–309.
- Zaroslav, Yu. D.; Gordeliy, V. I.; Kuklin, A. I.; Islamov, A. H.; Philippova, O. E.; Khokhlov, A. R.; Wegner, G. *Macromolecules* **2002**, *35*, 4466–4471.
- Wang, D.; Lal, J.; Moses, D.; Bazan, G. C.; Heeger, A. J. *Chem. Phys. Lett.* **2001**, *348*, 411–415.
- Gaylord, B. S.; Heeger, A. J.; Bazan, G. C. *Proc. Natl. Acad. Sci. U.S.A.* **2002**, *99*, 10954–10957.
- Ma, W.; Iyer, P. K.; Gong, X.; Liu, B.; Moses, D.; Bazan, G. C.; Heeger, A. J. *Adv. Mater.* **2005**, *17*, 274–277.
- Burrows, H. D.; Lobo, V. M. M.; Pina, J.; Ramos, M. L.; Seixas de Melo, J.; Valente, A. J. M.; Tapia, M. J.; Pradhan, S.; Scherf, U. *Macromolecules* **2004**, *37*, 7425–7427.
- Tapia, M. J.; Burrows, H. D.; Valente, A. J. M.; Pradhan, S.; Scherf, U.; Lobo, V. M. M.; Pina, J.; Seixas de Melo, J. *J. Phys. Chem. B* **2005**, *109*, 19108–19115.
- Chen, L.; Xu, S.; McBranch, D.; Whitten, D. *J. Am. Chem. Soc.* **2000**, *122*, 9302–9303.
- Chen, L.; Xu, S.; McBranch, D.; Whitten, D. Enhanced photo-physics of conjugated polymers, complex with surfactants, their manufacture and fluorescent properties. U.S. Patent Application 6569952, 2003, 12 pp.
- Fütterer, T.; Hellweg, T.; Findenegg, G. H.; Frahn, J.; Schlüter, A. D.; Böttcher, C. *Langmuir* **2003**, *19*, 6537–6544.
- Fütterer, T.; Hellweg, T.; Findenegg, G. H. Particle characterization by scattering methods in systems containing different types of aggregates: Aggregation of an amphiphilic poly(*p*-phenylene) in micellar surfactant. In *ACS Symposium Series: Mesoscale Phenomena in Fluid Systems*; Case, F., Alexandris, P., Eds.; American Chemical Society: Washington, DC, 2003; Vol. 861, p 117.
- Knaapila, M.; Stepanyan, R.; Horsburgh, L. E.; Monkman, A. P.; Serimaa, R.; Ikkala, O.; Subbotin, A.; Torkkeli, M.; ten Brinke, G. *J. Phys. Chem. B* **2003**, *107*, 14199–14203.
- Hellweg, T. *Curr. Opin. Colloid Interface Sci.* **2002**, *7*, 50–56.
- Holmberg, K.; Jönsson, B.; Kronberg, B.; Lindman, B. *Surfactants and Polymers in Aqueous Solutions*, 2nd ed.; Wiley: Chichester, UK, 2003.
- Sprunt, S.; Zhang, C. Y.; Litster, J. D. *J. Chem. Phys.* **1995**, *103*, 10325–10331.
- Salamat, G.; de Vries, R.; Kaler, E. W.; Satija, S.; Sung, L. *Langmuir* **2000**, *16*, 102–107.

- (25) Maccarini, M.; Briganti, G. *J. Phys. Chem. A* **2000**, *104*, 11451–11458.
- (26) Stoyanov, S. D.; Rehage, H.; Paunov, V. N. *Phys. Rev. Lett.* **2003**, *91*, 086102.
- (27) D'Arrigo, G.; Briganti, G.; Maccarini, M. *J. Phys. Chem. B* **2006**, *110*, 4612–4620.
- (28) Fritz, G.; Scherf, G.; Glatter, O. *J. Phys. Chem. B* **2000**, *104*, 3463–3470.
- (29) Lichtenfeld, F.; Schmeling, T.; Strey, R. *J. Phys. Chem.* **1986**, *90*, 5762–5766.
- (30) Menge, U.; Lang, P.; Findenegg, G. H.; Strunz, P. *J. Phys. Chem. B* **2003**, *107*, 1316–1320.
- (31) Kato, T.; Anzai, S.; Seimiya, T. *J. Phys. Chem.* **1990**, *94*, 7255–7259.
- (32) Strey, R.; Schomäcker, R.; Roux, D.; Nallet, F.; Olsson, U. *J. Chem. Soc., Faraday Trans.* **1990**, *86*, 2253–2261.
- (33) Hellweg, T.; von Klitzing, R. *Phys. A* **2000**, *283*, 349–358.
- (34) Fütterer, T.; Hellweg, T.; Findenegg, G. H.; Frahn, J.; Schlüter, A. D. *Macromolecules* **2005**, *38*, 7443–7450.
- (35) Fütterer, T.; Hellweg, T.; Findenegg, G. H.; Frahn, J.; Schlüter, A. D. *Macromolecules* **2005**, *38*, 7451–7455.
- (36) Burrows, H. D.; Lobo, V. M. M.; Pina, J.; Ramos, M. L.; Seixas de Melo, J.; Valente, A. J. M.; Tapia, M. J.; Pradhan, S.; Scherf, U.; Hintschich, S. I.; Rothe, C.; Monkman, A. P. *Colloids Surf. A* **2005**, *270–271*, 61–66.
- (37) Tiddy, G. J. T. *Phys. Rep.* **1980**, *57*, 1–46.
- (38) Dutschk, V.; Sabbatovskiy, K. G.; Stolz, M.; Grundke, K.; Rudoy, V. M. *J. Colloid Interface Sci.* **2003**, *267*, 456–462.
- (39) Rosta, L. *Appl. Phys. A* **2002**, *74*, S52.
- (40) Stuhrmann, H. B.; Burkhardt, N.; Dietrich, G.; Jünemann, R.; Meerwinck, W.; Schmitt, M.; Wadzack, J.; Willumeit, R.; Zhao, J.; Nierhaus, K. H. *Nucl. Instrum. Methods Phys. Res., Sect. A* **1995**, *A356*, 124–132.
- (41) Wignall, G. D.; Bates, F. S. *J. Appl. Crystallogr.* **1987**, *20*, 28–40.
- (42) Glatter, O. *J. Appl. Crystallogr.* **1977**, *10*, 415–421.
- (43) Pedersen, J. S.; Posselt, D.; Mortensen, K. *J. Appl. Crystallogr.* **1990**, *23*, 321–333.
- (44) Pedersen, J. S.; Schurtenberger, P. *J. Appl. Crystallogr.* **1996**, *29*, 646–661.
- (45) Pedersen, J. S.; Schurtenberger, P. *Macromolecules* **1996**, *29*, 7602–7612.
- (46) Jerke, G.; Pedersen, J. S.; Egelhaaf, S. U.; Schurtenberger, P. *Phys. Rev. E* **1997**, *56*, 5772–5788.
- (47) Holmes, M. C.; Reynolds, D. J.; Boden, N. *J. Phys. Chem.* **1987**, *91*, 5257–5262.
- (48) Iijima, H.; Kato, T.; Yoshida, H.; Imai, M. *J. Phys. Chem. B* **1998**, *102*, 990–995.
- (49) Fytas, G.; Nothofer, H. G.; Scherf, U.; Vlassopoulos, D.; Meier, G. *Macromolecules* **2002**, *35*, 481–488.
- (50) Knaapila, M.; Stepanyan, R.; Torckeli, M.; Lyons, B. P.; Ikonen, T. P.; Almásy, L.; Foreman, J. P.; Serimaa, R.; Güntner, R.; Scherf, U.; Monkman, A. P. *Phys. Rev. E* **2005**, *71*, 041802.
- (51) Chen, S. H.; Sheu, E. Y.; Kalus, J.; Hoffmann, H. *J. Appl. Crystallogr.* **1988**, *21*, 751–769.
- (52) Boden, N. Statistical Thermodynamics of Amphiphilic Self-Assembly: Structure and Phase Transitions in Micellar Solutions. In *Micelles, Membranes, Microemulsions, and Monolayers*; Gelbart, W. M., Ben-Shaul, A., Roux, D., Eds.; Springer: New York, 1994; pp 153–211.
- (53) Fletcher, P. D. I.; Nicholls, R. J. *Phys. Chem. Chem. Phys.* **2000**, *2*, 361–365.
- (54) Schmidt, P. W. Some Fundamental Concepts and Techniques Useful in Small-Angle Scattering Studies of Disordered Solids. In *Modern Aspects of Small-Angle Scattering*; Brumberger, H., Ed.; Kluwer Academic Publishers: Dordrecht, The Netherlands, 1995; pp 1–56.
- (55) Feigin, L. A.; Svergun, D. I. *Structure Analysis by Small-Angle X-ray and Neutron Scattering*; Plenum Press: New York, 1987.